

Original Communication

Selenium-Enriched Yeast Supplementation and Its Influence on Nerve Function Parameters and Oxidative Stress Markers in Diabetic Peripheral Neuropathy: A Randomized Placebo-Controlled Study

Mahsa Ghazisoltani¹, Sevil Ghaffarzadeh Rad¹, Payam Bahari-Moghaddam², Alireza Ostadrahimi³, Majid Mobasseri¹,*, Azizeh Farshbaf-Khalili⁴,*

Academic Editor: Torsten Bohn

Submitted: 10 July 2025 Revised: 21 September 2025 Accepted: 24 September 2025 Published: 15 December 2025

Abstract

Background: Diabetic peripheral neuropathy (DPN), a prevalent microvascular complication of diabetes mellitus, poses a substantial clinical burden and has a detrimental impact on quality of life. This triple-blind, randomized, placebo-controlled trial investigated the effects of an 8-week supplementation with selenium-enriched yeast on DPN symptoms, neuropathy severity, pro-oxidant-antioxidant balance (PAB), and sexual satisfaction in individuals aged 40-70 years with DPN. Methods: Fifty participants were randomized in a 1:1 ratio to receive either a daily 200 µg dose of Saccharomyces cerevisiae yeast-derived selenium (in a 500 mg capsule) or a placebo. Outcomes were assessed using validated tools: The Michigan Neuropathy Screening Instrument (MNSI) for symptoms, the Toronto Clinical Scoring System (TCSS) for severity, the Larson Sexual Satisfaction Questionnaire (LSSQ), and serum PAB levels via a specialized assay. Analyses followed a modified intention-to-treat approach, with ANCOVA and logistic regression used to adjust for confounders. **Results**: Post-intervention, both groups exhibited significant reductions in neuropathy symptoms (selenium: p < 0.001; placebo: p =0.001), though intergroup differences were non-significant [adjusted mean difference (aMD): -0.92; 95% CI: -1.9 to 0.10]. Neuropathy severity decreased significantly in the selenium group (p = 0.002) but not in the placebo group. While PAB levels declined markedly with selenium (p = 0.001), the between-group difference was non-significant (aMD: -32.1; 95% CI: -66.02 to 1.87). Sexual satisfaction scores improved significantly in the selenium group versus the placebo group (aMD: 8.51; 95% CI: 0.74 to 16.28). Conclusion: These findings suggest that selenium-enriched yeast supplementation may enhance biochemical markers (PAB) and quality-of-life parameters (sexual satisfaction) in DPN. However, its limited efficacy in improving neuropathy-specific outcomes underscores the need for larger trials to clarify its therapeutic potential. Clinical Trial Registration: This trial was registered at the Iranian Registry of Clinical Trials (IRCT20131009014957N10, https://trialsearch.who.int/Trial2.aspx?TrialID=IRCT20131009014957N10).

Keywords: diabetic neuropathy; selenium; supplementation; symptom assessment; severity of illness; oxidative stress

Main Points

- Diabetic Peripheral Neuropathy (DPN) is a common microvascular complication of diabetes mellitus.
- Some studies revealed that blood selenium levels and DPN are closely related.
- We evaluated the effect of organic selenium on symptoms and severity of DPN, oxidative stress, and sexual satisfaction.
- We observed the significant effect of selenium on improving the severity of DPN, oxidative stress, and sexual satisfaction.
- Further investigations are suggested to confirm the therapeutic effects of selenium on neuropathy outcomes.

1. Introduction

Diabetic peripheral neuropathy (DPN), a prevalent microvascular complication of diabetes mellitus (DM) [1], is characterized by significant clinical implications and substantial deterioration in patient quality of life [2]. This condition elevates risks of morbidity and disability, primarily through complications such as neuropathic ulceration and subsequent limb amputation [1]. Pathophysiologically, DPN manifests as symmetric sensorimotor polyneuropathy mediated by metabolic and microvascular alterations arising from chronic hyperglycemia and associated metabolic dysregulation [3].

The condition typically originates in the toes and progresses proximally [2], presenting clinically with both positive and negative sensory symptoms. Positive manifestations include paresthesia, burning pain, and neuropathic

¹Endocrine Research Center, Tabriz University of Medical Sciences, 5165665931 Tabriz, Iran

²Department of Occupational Therapy, Faculty of Rehabilitation Science, Tabriz University of Medical Sciences, 5165665931 Tabriz, Iran

³Nutrition Research Center, Tabriz University of Medical Sciences, 5165665931 Tabriz, Iran

⁴Physical Medicine and Rehabilitation Research Center, Aging Research Institute, Tabriz University of Medical Sciences, 5165665931 Tabriz, Iran

discomfort, while negative features encompass hypoesthesia (reduced tactile sensation), hypoalgesia (diminished pain perception), and palhypesthesia (decreased vibratory sensitivity) [4]. These sensory deficits contribute to impaired balance, gait instability, and elevated fall risk, potentially precipitating fractures and hospitalization [5]. Motor complications such as muscle weakness and spasms occur less frequently [6]. DPN is strongly associated with increased risks of lower extremity ulceration, limb amputation, and significant healthcare expenditures [7–9]. Early-stage pathophysiology involves segmental demyelination impairing nerve conduction velocity [10], progressing to axonal degeneration and myelin sheath damage. Diagnosis relies on nerve conduction studies (NCS) to differentiate DPN from other neuropathies [2].

Epidemiological studies reveal substantial global heterogeneity in DPN prevalence, attributed to variability in diagnostic criteria, diabetes subtypes, patient selection methodologies, and cohort sizes [11]. Reported prevalence rates demonstrate wide geographical disparities: 8.8% in China [12], 48.1% in Sri Lanka [13], 29.2% in India [14], 56.2% in Yemen [15], 39.5% in Jordan [16], 71.7% in Nigeria [17], 16.6% in Ghana [18], 26.5% in Malaysia [19], and 48.2% in Ethiopia [20]. Peripheral nerve degeneration in diabetes is typically irreversible [21,22], underscoring the critical need for early intervention. Established risk factors include advanced age, male sex, prolonged diabetes duration, concurrent microvascular complications, hypertension, rural residency, elevated body mass index (BMI), poor glycemic control (indicated by elevated Hemoglobin A1C (HbA1c) levels), alcohol consumption, chronic hyperglycemia, tobacco use, sedentary lifestyle, and marital status [1].

Current pharmacological management of neuropathic pain and paresthesia utilizes anticonvulsants, tricyclic antidepressants, or serotonin-noradrenaline reuptake inhibitors [23]. However, therapeutic strategies remain limited by the ineffectiveness of existing treatments for sensory loss [2], highlighting a critical gap in clinical management. The pathogenesis of DPN extends beyond hyperglycemiamediated neuronal damage, involving complex interactions between molecular mechanisms [6]. Key dysregulated pathways include enhanced polyol pathway flux, advanced glycation end-product (AGE) accumulation, oxidative stress, altered growth factor signaling, impaired insulin/C-peptide axis function, and protein kinase C (PKC) overactivation [24,25]. These processes simultaneously disrupt neuronal homeostasis and microvascular integrity within the vasa nervorum. Emerging pathophysiological models emphasize the convergence of multiple signaling networks and transcriptional regulators in DPN progression. Notably, extensive research has established oxidative stress, neuroinflammation, and programmed cell death as pivotal mechanisms in diabetic nerve injury through interconnected biochemical cascades [26]. This multifactorial

pathogenesis complicates the development of targeted neuroprotective therapies.

Selenium (Se) serves as an essential cofactor for antioxidant enzymes such as glutathione peroxidase (GPx), which neutralize reactive oxygen species (ROS) and prevent oxidative damage [27]. This micronutrient also contributes to the structural integrity of selenoprotein enzymes including deiodinase iodothyronine and thioredoxin reductase, which regulate thyroid hormone synthesis [28]. Clinical investigations have demonstrated an inverse relationship between blood selenium levels and DPN severity in type 2 diabetes patients [29]. Preclinical evidence from a 2011 rodent study (n = 40) revealed that sodium selenite supplementation mitigated diabetes-induced alterations in nerve conduction velocity distribution, restoring active nerve fiber parameters to age-matched control levels [30]. Similarly, selenate administration exhibited protective effects on electrophysiological parameters in diabetic models [30].

Selenoproteins, the biologically active forms of selenium, play critical antioxidant roles in numerous physiological processes [31]. Selenomethionine, the most bioavailable selenium species for human metabolism, accumulates efficiently in Saccharomyces cerevisiae yeast cultured in selenium-enriched media. Unlike inorganic forms, organic selenium derived from yeast undergoes site-specific amino acid absorption without mineral interactions, achieving complete bioavailability and systemic retention [32–34]. Selenium yeast supplementation is recognized as a safe, effective method for dietary fortification due to its superior absorption profile [35,36]. Currently, Saccharomyces cerevisiae represents the most efficient organism for selenomethionine accumulation and selenium biotransformation [37].

Given the escalating global prevalence of type 2 diabetes and its debilitating complications, novel therapeutic strategies for managing DPN require urgent investigation. Our extensive search of the database identified no prior clinical trials examining organic selenium's effects on DPN severity or symptomatology. This randomized controlled trial aimed to evaluate the efficacy of Saccharomyces cerevisiae-derived selenium over eight weeks on primary outcomes including neuropathic symptom severity and prooxidant-antioxidant balance (PAB) status, and a secondary outcome consisting of sexual satisfaction. This investigation seeks to address a critical gap in DPN management while elucidating selenium's potential therapeutic mechanisms through oxidative stress modulation.

2. Materials and Methods

2.1 Study Design and Participants

This triple-blind, parallel-group randomized controlled trial (RCT) enrolled participants aged 40–70 years with diagnosed DPN from outpatient clinics at Imam Reza and Sina Hospital in Tabriz, Iran. Ethical approval was ob-



tained from the Research Ethics Committee of Tabriz University of Medical Sciences (IR.TBZMED.REC.1401.374), and the trial was registered at the Iranian Registry of Clinical Trials (IRCT20131009014957N10; March 8, 2020). Researchers ensured protocol adherence through institutional authorization and strict compliance with the Declaration of Helsinki (1964) and its subsequent amendments. Prior to the intervention, participants received full disclosure of the study objectives, methodologies, potential risks/benefits, and withdrawal rights through direct researcher communication, followed by the acquisition of written informed consent.

Inclusion criteria encompassed: adults aged 40-70 years; a diagnosis of type 2 diabetes mellitus by an endocrinologist, including individuals with controlled (HbA1c <7%) and uncontrolled diabetes (HbA1c $\ge 7\%$) under stable antidiabetic regimens (≤20% insulin dose variation) and metabolic stability per medical records for ≥ 3 months; DPN confirmation via the Michigan Neuropathy Screening Instrument (MNSI); and independent ambulation capacity (>10 minutes). Exclusion criteria excluded participants with: non-diabetic neuropathy etiologies (alcoholassociated, chemotherapy-induced, congenital, inflammatory, thyroid dysfunction-related, vitamin B12 deficiency [serum < 200 pg/mL], HIV-associated, or idiopathic); coagulopathies; active malignancies; chronic renal failure (estimated glomerular filtration rate (eGFR) < 30 mL/min/1.73 m2); cerebrovascular accidents; neurodegenerative disorders (Parkinson's, Alzheimer's); severe depression; recent selenium supplementation (within the past 90 days); pregnancy/lactation (self-reported and physician-confirmed); or concurrent clinical trial participation.

Diabetes mellitus diagnosis followed WHO criteria: random plasma glucose ≥ 11.1 mmol/L, fasting plasma glucose ≥ 7.0 mmol/L, or a 2-hour oral glucose tolerance test result ≥ 11.1 mmol/L. Metabolic stability required documented glycemic control stability (HbA1c <9% at screening) with $\leq 20\%$ insulin dose adjustments and consistent oral hypoglycemic regimens for ≥ 3 months prior to enrollment.

2.2 Randomization and Intervention

Initial eligibility screening incorporated completion of the Michigan Neuropathy Symptom Questionnaire. Qualified volunteers who provided written informed consent underwent randomization via block-based allocation (1:1 ratio) using Random Allocation Software (RAS 2.0, M. Saghaei, Isfahan, Iran), which assigned participants to either an organic selenium group or a placebo arm. Participants received sequentially numbered opaque containers (1–50) containing either 60 capsules of 500 mg selenium-enriched yeast (200 µg selenium) or visually indistinguishable placebo capsules, both prepared under third-party supervision at the Nutrition Research Center. A triple-blind randomization scheme, generated by an independent re-

searcher, governed group allocation, with consecutive container distribution preserving allocation concealment. Researchers and participants remained blinded throughout the data collection and analysis phases, ensuring complete randomization integrity. All participants received standard clinical care alongside the intervention. The 500 mg capsules were identical across groups in physical characteristics (shape, color, weight, odor), maintaining triple-blind conditions between participants, researchers, and statistical analysts. Participants were instructed to ingest one capsule daily for two months.

2.2.1 Intervention Formulation

The selenium-enriched yeast (Se-yeast) capsules contained 500 mg of Saccharomyces cerevisiae biomass incorporating 200 µg of organic selenium. Placebo capsules consisted exclusively of 500 mg of non-enriched Saccharomyces cerevisiae cultivated under identical growth conditions (media composition, incubation temperature, pH parameters, harvest phase) without selenium supplementation. Both formulations were synthesized in the laboratory of the Tabriz Nutrition Research Center, affiliated with Tabriz University of Medical Sciences, following established protocols from Rahimi et al. [38]. In brief, Se-yeast was produced through the aerobic fermentation of Saccharomyces cerevisiae in a selenium-supplemented medium. The base medium consisted of sugarcane or beet molasses, supplemented with vitamins, nutrients, and growth factors to maximize biomass production. Selenium was introduced in the form of sodium selenite. Several parameters influenced both yeast growth and selenium uptake efficiency, including temperature, pH, selenium concentration, timing of selenium addition, aeration rate, inoculum volume, inoculation timing, and medium composition. During fermentation, selenium was incorporated into the yeast in organic form. Following fermentation, the yeast cells were separated via centrifugation, thoroughly washed, and dried to remove residual inorganic selenium bound to the cell walls. The final product was formulated as a 500 mg capsule containing 200 µg of organic selenium. Inductively coupled plasma mass spectrometry (ICP-MS) analysis verified a selenium concentration of 12,600 ppm (12,600 µg/kg) within the active compound.

2.2.2 Data Collection Procedures

Participants received standardized training in capsule administration and maintained daily medication adherence logs. Baseline evaluations included completion of demographic surveys and neuropathy severity and sexual satisfaction questionnaires. Anthropometric measurements employed calibrated instrumentation: body weight was recorded to a precision of ± 0.1 kg using a Seca digital scale (participants wearing light clothing), and height was measured to an accuracy of ± 0.1 cm via a wall-mounted stadiometer with participants positioned in an anatomical



stance (head, shoulders, buttocks, and heels contacting vertical reference planes). Venous blood samples (5 mL) were collected from the antecubital fossa using standardized phlebotomy techniques, processed in serum separation tubes containing clot activator gel, centrifuged at 3500 $\times g$ for 10 minutes, and cryopreserved at $-70~^{\circ}\mathrm{C}$ pending biochemical analysis. The subjects were referred to a single laboratory for fasting blood sugar measurement the next morning, and the intervention began after the sample was given.

The research team implemented weekly telephone follow-ups with participants to monitor medication adherence and protocol compliance. A dedicated contact number was provided to facilitate bidirectional communication, enabling subjects to address any study-related inquiries or concerns. Given that selenium functions as a nutritional supplement and considering the administered safe dosage—determined through preliminary safety assessments—no adverse effects were anticipated across medical, physical, mental, or psychological domains. Nevertheless, participants were explicitly informed that any suspected supplement-related complications would trigger immediate discontinuation of the intervention, accompanied by the provision of complementary medical management and follow-ups without any charge.

2.2.3 Post-Intervention Protocols

Following the 60-day intervention period, participants underwent repeat questionnaire administration and physiological assessments. The questionnaires on symptoms and severity of neuropathy and the sexual satisfaction questionnaire were completed again, and assessments were performed. The satisfaction with medication form was completed by asking patients on the pertinent sheet, and the drug side effects checklist was collected. A daily medication checklist and the used drug container were collected to count the remaining medication capsules. Fasting blood samples were collected the morning after intervention completion at a designated laboratory prior to initiating biochemical testing. The PAB assay was conducted using methodology adapted from Zahedi Avval et al. [39], involving sequential preparation of chromogenic substrates: a standard solution gradient combining 250 µM H₂O₂ (0-100%) with 3 mM uric acid in 10 mM NaOH; tetramethylbenzidine (TMB) dissolved in dimethyl sulfoxide (60 mg/10 mL) subsequently mixed with acetate buffer (0.05 M, pH 4.5) and chloramine T (70 μL, 100 mM) to create a TMB+ substrate. After vortexing and dark incubation (2 hours, 25 °C), peroxidase enzyme (25 U) was added before aliquoting and storage at -20 °C. Working solutions combined TMB+ with acetic acid buffer (0.05 M, pH 5.8) in darkness. Serum samples (10 µL) were reacted with 200 μL of working solution in ELISA plates during 12-minute 37 °C incubations, terminated by the addition of 2N HCl. Optical density measurements at 450 nm (reference wavelengths 570/620 nm) were converted to Hamidi-Koliakos (HK) units through interpolation from H_2O_2 -derived standard curves.

2.3 Assessment of Study Variables

The investigation evaluated primary outcomes encompassing DPN symptomatology, neuropathy severity, and serum PAB, with sexual satisfaction serving as the secondary endpoint. Data collection utilized an eight-section instrument comprising (1) eligibility criteria verification; (2) demographic profiling (age, gender, marital status, income level, education level, employment status, number of family members, place of residence, medication use, medical history, weight, height, and BMI); (3) the Michigan Neuropathy Screening Instrument (MNSI); (4) the Toronto Clinical Scoring System (TCSS) for neuropathy severity; (5) the Larson Sexual Satisfaction Questionnaire (LSSQ); (6) biochemical test documentation; (7) medication adherence through daily intake logs; and (8) adverse event monitoring. Psychometric validation of the demographic instrument involved iterative refinement through content and face validity assessments by a 10-expert panel, ensuring construct alignment and cultural appropriateness. Analytical standardization was achieved through uniform protocols: single-operator biochemical assays using identical equipment and reagent batches (Lot-Controlled Single-Kit Methodology) to minimize inter-assay variability.

2.3.1 Michigan Neuropathy Screening Instrument

The MNSI operationalizes DPN assessment through dual modalities: a 15-item self-reported symptom inventory (a score ≥ 7 indicates neuropathy) and a clinician-administered examination generating a maximum score of 8 through four items: (1) assessing pedal deformities; (2) dermatological pathology (xerosis, hyperkeratosis, and fissures), infectious lesions, ulceration status; (3) Achilles tendon reflexes; and (4) vibratory perception thresholds (128 Hz tuning fork application to the hallux); a score ≥ 2.5 confirms neuropathy [40,41].

2.3.2 Toronto Clinical Scoring System (TCSS)

The TCSS, originally developed by Perkins *et al.* [42], has demonstrated cross-cultural validity through prior psychometric evaluations in Iranian populations. This instrument utilizes a 19-point scale to grade diabetic neuropathy severity. Diagnostic thresholds are stratified into three categories: 0–6 (mild neuropathy), 7–13 (moderate neuropathy), and >13 (severe sensory impairment), with stratification criteria specifically calibrated to vibratory and proprioceptive deficits in the hallux region during neurological assessment [42].

2.3.3 Larson Sexual Satisfaction Questionnaire (LSSQ)

The instrument comprises 25 items employing a 5-point Likert scale (1: never to 5: always), yielding a total



score range of 25–125. Sexual satisfaction is stratified into four categories: dissatisfaction (\leq 50), low (51–75), moderate (76–100), and high satisfaction (101–125) [43]. Psychometric validation was conducted in Shams Mofarrahe's study [44], "Investigating the Effect of Marital Counseling on Couples' Sexual Satisfaction", which established face and content validity through expert appraisal. Test-retest reliability analysis demonstrated exceptional consistency, with a coefficient of 0.98.

2.4 Sample Size Determination

A priori power analysis via G*Power 3.1.2 incorporated parameters from Ahrary *et al.* [40]: neuropathy symptom reduction (Δ = 2.74, SD = 2.94, α = 0.05, β = 0.1) necessitated n = 20 per group, while severity score differences (Δ = 1.5, SD = 2.89, α = 0.05, β = 0.1) required n = 23 per group. Accounting for 10% attrition, the final sample comprised 25 participants per arm (50 total), ensuring adequate statistical power across outcome measures.

2.5 Statistical Analysis

Statistical analyses were performed using SPSS version 21 (IBM Corporation, Armonk, NY, USA) through a hierarchical analytical framework adhering to modified intention-to-treat principles. Normality assumptions were verified for each interventional group using Kolmogorov-Smirnov and Shapiro-Wilk tests, with Levene's test confirming variance homogeneity. Analytical procedures encompassed: intergroup comparisons via χ^2 /Fisher's exact tests for categorical variables, Mann-Whitney U tests for ordinal data, and independent t-tests/ANCOVA (adjusted for baseline measures and confounders) for continuous outcomes; intragroup longitudinal changes assessed through paired t-tests; and adjusted odds ratios with 95% confidence intervals (CIs) derived from logistic regression models, validated through Hosmer-Lemeshow goodness-of-fit testing. All inferential analyses maintained a two-tailed significance threshold of $\alpha = 0.05$.

3. Results

Participant recruitment occurred between April 21, 2021, and October 22, 2024, with the follow-up period concluding on December 22, 2024. A total of 335 individuals with diabetes were screened. Of these, 253 did not meet the inclusion criteria, and 32 were excluded due to unwillingness to participate (n = 19), digestive disorders (n = 4), coagulation risks (n = 5), rheumatological conditions (n = 3), or chronic renal failure (n = 1). The remaining 50 eligible participants were randomized into two groups (25 per group). During the 8-week intervention, two participants in the organic selenium group and one in the placebo group withdrew due to unwillingness to continue. Final analysis included 23 participants in the selenium group and 24 in the placebo group (Fig. 1).

Table 1 summarizes demographic and clinical characteristics. No statistically significant differences were observed between groups in terms of sex, age, marital status, residence, education, income, employment, weight, or BMI (all p > 0.05). All participants had type 2 diabetes, with four insulin users per group and three participants using insulin combined with gliclazide. Mean diabetes duration exceeded 10 years (13.5 years in the intervention group vs. 14.6 in the placebo group; p = 0.571). Hypertension (prevalence >70% in both groups; p = 1.000), hyperlipidemia (80% in both groups; p = 1.000), and smoking rates (p = 0.167) were comparable. Baseline fasting blood glucose levels differed significantly between groups (p = 0.035), with both exceeding normal ranges.

Table 2 details diabetic peripheral neuropathy (DPN) symptom scores using the Michigan Neuropathy Screening Instrument (MNSI). Baseline total scores were similar (8.6 \pm 2.1 in the selenium group vs. 8.2 \pm 1.9 in the placebo group; p = 0.271), both above the diagnostic threshold of 7. Post-intervention reductions were significant in both groups (selenium: p < 0.001; placebo: p = 0.001), with a greater decline in the selenium group (adjusted mean difference [aMD]: -0.92; 95% CI: -1.9 to 0.10), which resulted in a mean score lower than the borderline of 7. However, intergroup differences remained nonsignificant (p = 0.077), resulting in reducing mean scores below the threshold (6.5 \pm 2.0) [Clarified that the scores were reduced, not that the lack of significance reduced them]. There was no significant difference in the mean score of the MNSI examination section between the two groups before the intervention (p =0.667). MNSI examination scores decreased significantly within groups (selenium: p = 0.004; placebo: p = 0.008) but not between groups (p = 0.969). Intragroup improvements occurred in foot appearance (selenium: p = 0.029; placebo: p = 0.008) and hallux vibratory perception (selenium: p = 0.016), though intergroup comparisons showed no significant differences (all p > 0.05).

Table 3 presents neuropathy severity outcomes using the Toronto Clinical Scoring System (TCSS). Baseline scores indicated moderate severity in both groups (12.2 \pm 3.2 vs. 11.2 \pm 2.9; p=0.259), with >60% of participants classified as moderate in both groups (p=0.311). Post-intervention, the selenium group showed significant improvement (p=0.002), though intergroup differences remained nonsignificant (p=0.214). Intragroup analysis revealed reductions in symptom (p=0.015) and sensory scores (p=0.017) for the selenium group, and in sensory scores for the placebo group (p=0.045), with no significant intergroup differences in symptom (p=0.992), reflex (p=0.319), or sensory (p=0.456) domains.

Table 4 outlines PAB results. Baseline serum PAB levels were comparable (p=0.424). Post-intervention, selenium supplementation significantly reduced PAB levels (p=0.001), though intergroup differences were nonsignificant (aMD: -32.1; 95% CI: -66.02 to 1.87; p=0.063).



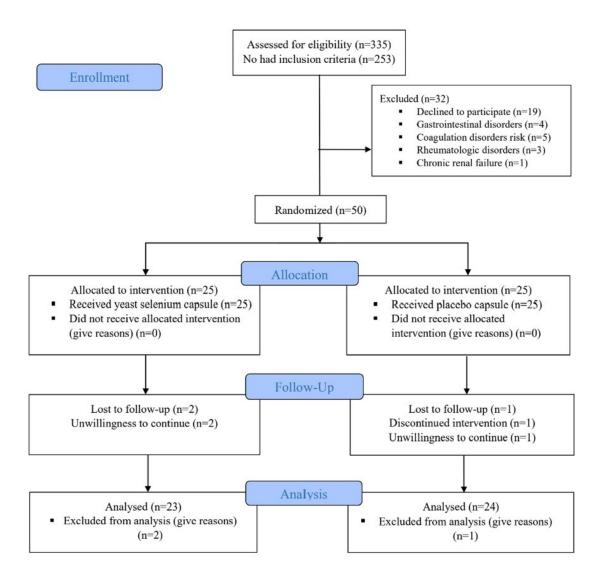


Fig. 1. The flow diagram of study.

Table 5 details sexual satisfaction outcomes. Baseline scores reflected moderate satisfaction in both groups (p=0.964). Post-intervention, placebo group scores declined to low satisfaction (p=0.004), while the selenium group maintained moderate levels (p=0.277). Final satisfaction scores were higher in the selenium group (aMD: 8.51; 95% CI: 0.74–16.28; p=0.033), with 84.2% of selenium participants reporting moderate satisfaction and 75% of placebo participants reporting low satisfaction (p=0.001).

Intervention satisfaction differed marginally: 43.4% of selenium and 25% of placebo participants reported being satisfied, while 52.2% vs. 58.3% were neutral, and 8.7% vs. 16.7% were dissatisfied (p = 0.445). Adverse events included mild diarrhea, abdominal pain, and dizziness (4 in the selenium group, 3 in the placebo group), none requiring discontinuation. Capsule counting from the returned

medications and review of the controlling daily medication checklist showed that medication compliance exceeded 90% in both groups

4. Discussion

The analysis revealed that both groups demonstrated comparable baseline total MNSI scores exceeding established clinical thresholds. Post-intervention assessment at eight weeks showed statistically significant reductions in total scores for both the selenium-enriched yeast-supplemented and placebo groups. While the selenium group exhibited greater mean decreases, achieving post-treatment scores below clinical cutoffs, intergroup comparisons revealed no statistically significant differential effects. Longitudinal evaluation of four MNSI subdomains (symptom inventory, appearance, sensory perception, and



Table 1. Demographic & baseline characteristics of participants with diabetic peripheral neuropathy (DPN).

	Selenium	Placebo	
Characteristics	n = 25 n (%)	n = 25	p
		n (%)	
Age (years)			0.316 [¥]
40–50	7 (28.0)	6 (24.0)	
51–60	9 (36.0)	5 (20.0)	
61–70	9 (36.0)	14 (56.0)	
Gender			$0.551^{\rm f}$
Female	15 (60.0)	18 (72.0)	
Male	10 (40.0)	7 (28.0)	
Habitation			$1.000^{\rm f}$
City	23 (92.0)	24 (96.0)	
Country	2 (8.0)	1 (4.0)	
Marriage			$1.000^{\rm f}$
Single	5 (20.0)	6 (24.0)	
Married	20 (80.0)	19 (76.0)	
Education	` ′		0.180€
Illiterate	5 (20.0)	11 (44.0)	
Under diploma	15 (60.0)	11 (44.0)	
Diploma and higher	5 (20.0)	3 (12.0)	
Occupation	, ,	, ,	0.498^{Ψ}
Home wife	14 (56.0)	18 (72.0)	
Unoccupied	3 (12.0)	2 (8.0)	
Occupied	8 (32.0)	5 (20.0)	
Family income	,	,	0.382€
Sufficient	3 (12.0)	1 (4.0)	
Rather sufficient	9 (36.0)	9 (36.0)	
Insufficient	13 (52.0)	15 (60.0)	
Weight (kg)/Mean \pm SD	78.6 ± 14.9	74.6 ± 15.0	0.353*
BMI (kg/m ²)			$0.697^{ ext{\text{$\frac{1}{2}}}}$
Normal (18.5–25.9 kg/m ²)	3 (12.0)	5 (20.0)	
Overweight (25–29.9 kg/m ²)	10 (40.0)	8 (32.0)	
Obese ($>30 \text{ kg/m}^2$)	12 (48.0)	12 (48.0)	
Type of diabetes/Type II	25 (100.0)	25 (100.0)	$1.000^{\rm f}$
FBS (mg/dL)	180.3 (51.2)	150.0 (47.7)	0.035*
Duration of DPN (years)/Mean \pm SD	13.5 ± 5.8	14.6 ± 7.6	0.571*
Taking diabetic medications	15.6 ± 5.6	1110 1 710	0.889 ^f
Gloripa	6 (24.0)	6 (24.0)	
Gliclazide	5 (20.0)	6 (24.0)	
Metformin+gloripa	4 (16.0)	3 (12.0)	
Metformin	4 (16.0)	5 (20.0)	
Insulin	4 (16.0)	4 (16.0)	
Insulin+gliclazide	2 (8.0)	1 (4.0)	
Smoking	2 (0.0)	1 (1.0)	0.167^{4}
Never	19 (76.0)	23 (92.0)	0.107
Current smoker	3 (12.0)	(- -)	
Past smoker	3 (12.0)	2 (8.0)	
Having hypertension/yes	19 (76.0)	18 (72.0)	$1.000^{\rm f}$
Having hyperlipidemia/yes	20 (80.0)	20 (80.0)	1.000 ^f

DPN, diabetic peripheral neuropathy; BMI, body mass index; MNSI, Michigan neuropathy screening instrument; A score of ≥ 7 was considered abnormal. The total possible score of MNSI examination (1. foot appearance, 2. ulceration, 3. ankle reflexes, 4. vibration perception) is 8 points and a score ≥ 2.5 is considered abnormal, ^f Fishers exact test, [§] Chi-square test, [©] Chi-square by trend, * Independent *t*-test.



Table 2. The score of Michigan neuropathy screening instrument (MNSI) symptoms among participants by study groups.

Variable	Selenium $(n = 25)$	Placebo $(n = 25)$	aMD (95% CI)	<i>p</i> -value
Variable	Mean (SD)	Mean (SD)	- alviD (9370 CI)	p-value
Total MNSI (0–15)				
Baseline	8.6 (1.5)	8.2 (1.2)	-	0.271*
After 2 month	6.5 (2.0)	7.1 (1.7)	-0.92 (-1.9 to 0.10)	0.077§
$p ext{-value}^{\Psi}$	< 0.001	0.001		
Total MNSI examination (0-8)				
Baseline	2.6 (1.1)	2.5 (0.9)	-	0.667*
After 2 month	1.8 (1.2)	1.8 (1.0)	-0.01 (-0.65 to 0.63)	0.969§
$p ext{-value}^{\Psi}$	0.004	0.008		
MNSI exam 1/Disordered	n (%)	n (%)	OR (95% CI)	<i>p</i> -value
Baseline	13 (52.0)	15 (60.0)	-	$0.776^{\rm f}$
After 2 month	7 (28.0)	9 (36.0)	0.84 (0.23 to 3.02)	0.792^{\pm}
p -value $^{\Psi}$	0.029	0.008		
MNSI exam 2/Disordered			OR (95% CI)	<i>p</i> -value
Baseline	7 (28.0)	5 (20.0)	-	$0.742^{\rm f}$
After 2 month	3 (12.0)	3 (12.0)	0.79 (0.12 to 4.98)	0.792^{\pm}
$p ext{-value}^{\Psi}$	0.125	0.688		
MNSI exam 3/Disordered			OR (95% CI)	<i>p</i> -value
Baseline	20 (80.0)	19 (76.0)	-	$1.000^{\rm f}$
After 2 month	15 (60.0)	16 (64.0)	0.78 (0.24 to 2.58)	0.687^{\pm}
$p ext{-value}^{\Psi}$	0.125	0.508		
MNSI exam 4/Disordered			OR (95% CI)	<i>p</i> -value
Baseline	25 (100)	23 (92.0)	-	$0.490^{\rm f}$
After 2 month	18 (72.0)	18 (72.0)	1.12 (0.32 to 3.91)	0.853^{\pm}
p -value $^{\Psi}$	0.016	0.180		

^{*} Independent *t*-test, aMD (95% CI): adjusted Mean difference (95% confidence interval). § ANCOVA adjusted for baseline values, age, and sex, * Paired Samples *t*-test, ± Logistic regression adjusted for baseline values, age, and sex, f Fishers exact test. The bold *p*-values indicate the statistical significance. MNSI is a two-part instrument, with the first part consisting of 15 questions to be completed by the individual and the second part consisting of 4 questions to be completed by a professional after examination. In the first part, the responses are summed to obtain a total score. A score greater than or equal to 7 is considered abnormal. The range of scores for Total MNSI and Total MNSI examination is 0–15 and 0–8, respectively.

reflex testing) similarly showed non-significant between-group differences at baseline and post-intervention. These findings suggest comparable therapeutic trajectories despite numerical advantages in the experimental group. Neuropathy severity evaluation using TCSS demonstrated moderate mean total scores at baseline in both groups. Post-intervention, the selenium group showed significant intragroup reductions in neuropathy severity. The organic selenium group exhibited significant decreases in PAB index values during treatment and statistically greater improvements in sexual satisfaction metrics compared to placebo controls.

Pain pathophysiology involves complex interactions between oxidative stress and Ca²⁺ signaling. Mitochondrial dysfunction caused by Ca²⁺-induced membrane depolarization generates ROS, which is modulated by antioxidants including selenium. As a GPx cofactor and selenoprotein component, selenium exerts neuroprotective effects by regulating ROS overproduction, inflammation, and Ca²⁺

overload in conditions like diabetic neuropathy, allodynia, and nociception. Ca²⁺ influx occurs through transient receptor potential (TRP) channels (TRPA1, TRPM2, TRPV1, TRPV4), many of which are activated by oxidative stress to induce peripheral pain. Recent studies indicate that selenium modulates peripheral pain via TRP channel inhibition in dorsal root ganglia [45]. Song and Lin [46] demonstrated that selenium upregulates the nuclear factor erythroid 2-related factor 2/heme oxygenase-1Nrf2/HO-1 pathway to inhibit oxidative stress and p38 MAPK phosphorylation, enhancing Schwann cell proliferation and peripheral nerve regeneration.

Current models characterize pain as a multifactorial process involving oxidative stress-mediated neural cascades. Mitochondrial dysfunction from Ca²⁺-induced ROS generation is central to nociceptive processing. Selenium's therapeutic potential arises from its dual role in GPx activation and selenoprotein synthesis, mitigating oxidative damage in chronic pain [47]. Altuhafi *et al.* [48] delin-



Table 3. The score of Toronto Clinical Scoring System (TCSS) among participants by study groups.

Variable	Selenium (n = 25)	Placebo (n = 25)	- aMD (95% CI)	p -value $^\pm$	
variable	Mean (SD)	Mean (SD)	alviD (9370 C1)	p-value-	
Total TCSS (0-19)					
Baseline	12.2 (3.2)	11.2 (2.9)	-	0.259*	
After 2 month	9.5 (4.8)	10.2 (4.0)	-1.34 (-3.49 to 0.81)	0.214§	
p -value $^{\Psi}$	0.002	0.106			
Symptom scores (0–6)					
Baseline	4.5 (1.5)	4.5 (1.2)	-	0.992*	
After 2 month	3.4 (2.2)	4.2 (1.7)	-0.71 (-1.68 to 0.26)	0.147§	
p -value $^{\Psi}$	0.015	0.181			
Reflex scores (0–8)					
Baseline	4.3 (2.5)	3.6 (2.5)	-	0.319*	
After 2 month	2.9 (2.6)	3.6 (2.6)	-0.71 (-2.19 to 0.77)	0.337^{\S}	
p -value $^{\Psi}$	0.092	1.0			
Sensory test scores (0–5)					
Baseline	3.4 (1.1)	3.2 (0.9)	-	0.456*	
After 2 month	2.6 (1.2)	2.7 (1.2)	-0.17 (-0.87 to 0.54)	0.636^{\S}	
p -value $^{\Psi}$	0.017	0.045			
TCSS severity	n (%)	n (%)		$p ext{-value}^\pm$	
Baseline					
Mild	1 (4.0)	2 (8.0)	-	0.311	
Moderate	15 (60.0)	17 (68.0)			
Severe	9 (36.0)	6 (24.0)			
After 2 month					
Mild	6 (25.0)	4 (17.4)	-	0.658	
Moderate	13 (54.2)	15 (62.2)			
Severe	5 (20.8)	4 (17.4)			

TCSS is a screening tool for diabetic peripheral neuropathy, and correlates with diabetic neuropathy severity. Maximum TCNS is 19 points. Symptom scores graded as 0 = absent, 1 = present; Sensory test scores graded as 0 = normal, 1 = abnormal, Reflexes graded as 0 = normal, 1 = reduced, 2 = absent. Scores of 0-6 indicate mild neuropathy, 7-13 indicate moderate neuropathy, and scores above 13 indicate severe sensory disturbances.

Table 4. The score of pro-oxidant-antioxidant balance (PAB) among participants by study groups.

Selenium $(n = 25)$	Placebo $(n = 25)$	aMD (95% CI)	<i>p</i> -value
Mean (SD)	Mean (SD)	aiviD (5570 C1)	
192.9 (34.5)	182.4 (55.6)	-	0.424*
134.3 (49.6)	163.7 (45.7)	-32.07 (-66.02 to 1.87)	0.063§
-58.6 (-89.3 to -28.1)	-18.6 (-45.0 to 7.8)		
0.001	0.159		
	Mean (SD) 192.9 (34.5) 134.3 (49.6) -58.6 (-89.3 to -28.1)	Mean (SD) Mean (SD) 192.9 (34.5) 182.4 (55.6) 134.3 (49.6) 163.7 (45.7) -58.6 (-89.3 to -28.1) -18.6 (-45.0 to 7.8)	Mean (SD) Mean (SD) 192.9 (34.5) 182.4 (55.6) - 134.3 (49.6) 163.7 (45.7) -32.07 (-66.02 to 1.87) -58.6 (-89.3 to -28.1) -18.6 (-45.0 to 7.8)

^{*} Independent t-test, aMD (95% CI): adjusted Mean difference (95% confidence interval). § ANCOVA adjusted for baseline values, age, and sex. § Paired Samples t-test. The bold p-value indicates the statistical significance. The bold p-values indicate the statistical significance.

eated oxidative stress mechanisms in type 2 diabetes mellitus (T2DM) pathogenesis, where chronic hyperglycemia drives free radical production via glucose autoxidation and impairs antioxidant systems. This redox imbalance causes cellular organelle damage, elevated lipid peroxidation, and insulin resistance [49].

Diabetic metabolic dysregulation increases mitochondrial ROS in pancreatic β -cells while suppressing GPx activity [50]. Persistent hyperglycemia elevates superoxide production [45], with organic hydroperoxide accumulation correlating with membrane pathophysiology and diabetic complications. Clinical data show inverse correla-



^{*} Independent *t*-test, aMD (95% CI): adjusted Mean difference (95% confidence interval). § ANCOVA adjusted for baseline values, age, and sex, $^{\text{\frac{4}}}$ Paired Samples *t*-test, $^{\pm}$ Mann-Whitney U. The bold *p*-values indicate the statistical significance.

Table 5. The score of Larson's sexual satisfaction among participants by study groups.

Variable	Selenium $(n = 20)$	Placebo $(n = 19)$	aMD (95% CI)	<i>p</i> -value
variable	Mean (SD)	Mean (SD) Mean (SD)		p-value
Sexual satisfaction score (25–125)				
Baseline	85.1 (13.2)	84.9 (16.2)	-	0.964*
After 2 month	81.1 (13.4)	73.5 (9.2)	8.51 (0.74 to 16.28)	0.033§
$p ext{-value}^{ooagle}$	0.277	0.004		
Sexual satisfaction rate	n (%)	n (%)		p -value $^\pm$
Baseline				
No (<50)	-	-		0.771
Low (51–75)	3 (15.0)	6 (31.6)		
Moderate (76–100)	15 (75.0)	9 (47.4)		
High (101–125)	2 (10.0)	4 (21.1)		
After 2 months				
No (<50)	-	-		$\textbf{0.001}^{\pm}$
Low (51–75)	4 (20.0)	16 (84.2)		
Moderate (76–100)	15 (75.0)	2 (10.5)		
High (101–125)	1 (5.0)	1 (5.3)		

The higher the total score, the more sexual satisfaction, aMD (95% CI): mean difference (95% confidence interval), * Independent *t*-test, \S ANCOVA adjusted for baseline, age, BMI, \S paired Samples *t*-test, \S Mann-Whitney U. The bold *p*-values indicate the statistical significance.

tions between lipid peroxidation biomarkers and erythrocyte GPx activity in diabetics [51]. Multiple studies document compromised selenium status in diabetic populations. Altuhafi's work [48] identifies positive correlations between serum selenium concentrations and GPx activity, suggesting that selenoprotein deficiency exacerbates prooxidative states, thereby accelerating complications. Systemic antioxidant depletion in diabetes reflects profound redox dysregulation, where chronic ROS overproduction impairs defense mechanisms, creating a self-perpetuating cycle of oxidative damage. This underscores the importance of therapeutic strategies targeting selenium-dependent antioxidant pathways in diabetes management.

Huo et al. [52] conducted a meta-analysis of 14 randomized controlled trials (n = 1384 participants) investigating nutraceutical interventions for DPN. Their systematic review identified three high-quality trials demonstrating that vitamin and antioxidant supplementation significantly enhanced sural nerve sensory nerve conduction velocity (SNCV). Furthermore, pooled data from seven trials (n = 758) suggested improved peroneal nerve motor nerve conduction velocity (MNCV), while four studies reported analogous enhancements in median nerve MNCV. These findings align empirically with our observations regarding selenium's capacity to mitigate DPN severity. Complementary mechanistic evidence emerges from Aydın and Nazıroğlu [53], who demonstrated that streptozotocin (STZ)-induced TRPM7 channel activation exacerbates neuropathic pain and apoptotic pathways in murine models. Crucially, selenium administration attenuated TRPM7-mediated neurotoxicity, mirroring our findings of selenium's neuroprotective efficacy. Jafari et al. [54] further corroborated this therapeutic potential through *in vitro* experiments showing that selenium co-treatment significantly reduced vincristine-induced cytotoxicity in PC12 neuronal cells (p < 0.01 vs. vincristine-only controls).

Ayaz and Kaptan [30] provided electrophysiological evidence that sodium selenite intervention ameliorates diabetes-induced neural conduction abnormalities, including prolonged signal transmission latency, increased action potential amplitude (32% vs. diabetic controls), and restoration of fast/slow fiber conduction velocity distributions to age-matched normative ranges. These data synergize with Xu et al.'s clinical correlation analysis [29], which identified low selenium status as an independent risk factor for diabetic polyneuropathy progression in type 2 diabetes cohorts. At the molecular level, Nazıroğlu et al. [47] postulated that selenium's antinociceptive mechanism involves TRP channel modulation in dorsal root ganglia, potentially disrupting nociceptive signaling cascades. Collectively, these multidisciplinary findings substantiate selenium's dual role as both a redox modulator and ion channel regulator in diabetic neuropathy pathogenesis, providing mechanistic context for its observed clinical benefits in DPN management.

Teimouri *et al.* [55] conducted a double-blind, randomized, placebo-controlled clinical trial to investigate the therapeutic efficacy of selenium supplementation on psychosexual outcomes. Their findings demonstrated clinically significant amelioration of core premature ejaculation symptomatology in the intervention group compared to placebo controls (p < 0.05), with particular improvements observed in sexual satisfaction metrics. The intervention demonstrated favorable tolerability profiles, with



adverse event incidence comparable to placebo. This empirical evidence substantiates the therapeutic potential of selenium in addressing sexual dysfunction parameters, particularly through modulation of ejaculatory latency mechanisms. These findings exhibit notable congruence with our experimental observations regarding selenium's putative role in enhancing psychosexual outcomes via redox homeostasis regulation and neurotransmitter modulation pathways. However, the research by Rahimi et al. [38], which reported results contrary to our findings, stated that no significant difference in sexual satisfaction was observed in the groups consuming selenium compared to the placebo at the end of the intervention [Improved flow and conciseness]. Given that the drug dose and selenium (selenoprotein) composition were identical in both studies, the discrepancy may relate to differences in treatment duration and participant characteristics, as Rahimi et al.'s study [38] included postpartum women without diabetes.

One of the limitations of our study was the restricted sample size, which future research should address through larger cohorts. A key strength was the triple-blind design, enhancing methodological rigor. Additional strengths included standardized examinations performed by an experienced specialist, precise follow-up timing to ensure timely patient referrals, and the use of organic selenium with high bioavailability and lower toxicity. However, the absence of speciation analysis introduces uncertainty regarding the supplement's composition and bioactivity, necessitating such analyses in future trials to clarify bioavailability and therapeutic efficacy. Larger studies should also characterize selenium species and investigate dose-response relationships. This study did not evaluate key inflammatory biomarkers, such as IL-6, IL-8, and TNF- α , in either serum or tissue samples. Considering the established association between chronic inflammation and the progression of DPN, the inclusion of such biomarkers in future investigations would substantiate mechanistic interpretations. Therefore, it is recommended that subsequent research incorporate these analyses. Furthermore, the results cannot be generalized to patients with type 1 diabetes or those with metabolic instability or unstable antidiabetic regimens.

5. Conclusion

The 8-week intervention yielded significant therapeutic outcomes across both groups. Post-treatment evaluation revealed clinically meaningful reductions in MNSI scores for both groups, though intergroup comparisons showed no statistically significant differential effect. Neuropathic severity assessments using the TCSS demonstrated a 38% baseline-to-post-intervention score reduction exclusively in the selenium group. Psychosexual outcomes diverged markedly between groups: placebo recipients exhibited a progressive decline in sexual satisfaction metrics to subclinical thresholds by week 8, whereas the selenium group maintained baseline satisfaction levels, achieving statisti-

cally superior terminal scores. Selenium supplementation also significantly reduced the PAB index. These findings suggest that selenium may simultaneously mitigate neuropathic progression and preserve sexual function in diabetic populations, though its therapeutic effect on neuropathy outcomes requires further investigation through larger trials with speciation analysis, extended durations, and doseresponse characterization.

Availability of Data and Materials

The datasets supporting this study's findings are available from corresponding author per request.

Author Contributions

MG and AFK were responsible for drafting the protocol, data collection, data analysis, and interpretation and writing of the final manuscript. MM was responsible for drafting the protocol, project administration, data analysis and interpretation, and editing the final version of the manuscript. SGR, PBM and AO were involved in the acquisition of data and critically reviewed the manuscript for intellectual content. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Ethical approval was obtained from the Research Ethics Committee of Tabriz University of Medical Sciences (IR.TBZMED.REC.1401.374). Researchers ensured protocol adherence through institutional authorization and strict compliance with the Declaration of Helsinki (1964) and its subsequent amendments. Prior to the intervention, participants received full disclosure of the study objectives, methodologies, potential risks/benefits, and withdrawal rights through direct researcher communication, followed by the acquisition of written informed consent. This trial was registered at the Iranian Registry of Clinical Trials (IRCT20131009014957N10, https://trialsearch.who.int/Trial2.aspx?TrialID=IRCT20131009014957N10).

Acknowledgment

The authors extend their gratitude to all study participants for their contributions. Additionally, they acknowledge the institutional support provided by the Research Department of Tabriz University of Medical Sciences, which facilitated the completion of this research endeavor.

Funding

The Vice-Chancellor for Research and Technology, Tabriz University of Medical Sciences has funded the original research (Grant No: 69271).



Conflict of Interest

The authors declare no conflict of interest.

Declaration of AI and AI-Assisted Technologies in the Writing Process

The authors utilized DeepSeek (DeepSeek-V3) for scientific English language editing to improve the clarity and grammar of this manuscript. Following AI-assisted editing, the authors thoroughly reviewed and edited the content as needed and take full responsibility for the published work.

References

- [1] Khan S, Ahmad S, Khan M, Lohani M, Khan MS, Haneef M. Diabetic Peripheral Neuropathy: Navigating Controversies and Pioneering Advances. Advancements in Life Sciences. 2025; 12: 01–12. http://dx.doi.org/10.62940/als.v12i1.3627.
- [2] Yang Y, Zhao B, Wang Y, Lan H, Liu X, Hu Y, et al. Diabetic neuropathy: cutting-edge research and future directions. Signal Transduction and Targeted Therapy. 2025; 10: 132. https://doi. org/10.1038/s41392-025-02175-1.
- [3] Tesfaye S, Boulton AJM, Dyck PJ, Freeman R, Horowitz M, Kempler P, et al. Diabetic neuropathies: update on definitions, diagnostic criteria, estimation of severity, and treatments. Diabetes Care. 2010; 33: 2285–2293. https://doi.org/10.2337/dc 10-1303.
- [4] Callaghan BC, Price RS, Feldman EL. Distal Symmetric Polyneuropathy: A Review. JAMA. 2015; 314: 2172–2181. https://doi.org/10.1001/jama.2015.13611.
- [5] Moseley KF. Type 2 diabetes and bone fractures. Current Opinion in Endocrinology, Diabetes, and Obesity. 2012; 19: 128–135. https://doi.org/10.1097/MED.0b013e328350a6e1.
- [6] Boulton AJM, Malik RA, Arezzo JC, Sosenko JM. Diabetic somatic neuropathies. Diabetes Care. 2004; 27: 1458–1486. https://doi.org/10.2337/diacare.27.6.1458.
- [7] Alleman CJM, Westerhout KY, Hensen M, Chambers C, Stoker M, Long S, et al. Humanistic and economic burden of painful diabetic peripheral neuropathy in Europe: A review of the literature. Diabetes Research and Clinical Practice. 2015; 109: 215–225. https://doi.org/10.1016/j.diabres.2015.04.031.
- [8] Gordois A, Scuffham P, Shearer A, Oglesby A. The healthcare costs of diabetic peripheral neuropathy in the UK. Diabetic Foot Journal. 2003; 6: 62–73.
- [9] Happich M, John J, Stamenitis S, Clouth J, Polnau D. The quality of life and economic burden of neuropathy in diabetic patients in Germany in 2002–results from the Diabetic Microvascular Complications (DIMICO) study. Diabetes Research and Clinical Practice. 2008; 81: 223–230. https://doi.org/10.1016/j.diab res.2008.03.019.
- [10] American Diabetes Association. 4. Comprehensive Medical Evaluation and Assessment of Comorbidities: Standards of Medical Care in Diabetes-2020. Diabetes Care. 2020; 43: S37– S47. https://doi.org/10.2337/dc20-S004.
- [11] Edwards JL, Vincent AM, Cheng HT, Feldman EL. Diabetic neuropathy: mechanisms to management. Pharmacology & Therapeutics. 2008; 120: 1–34. https://doi.org/10.1016/j.pharmt hera.2008.05.005.
- [12] Lu B, Hu J, Wen J, Zhang Z, Zhou L, Li Y, et al. Determination of peripheral neuropathy prevalence and associated factors in Chinese subjects with diabetes and pre-diabetes Shang-Hai Diabetic neuRopathy Epidemiology and Molecular Genetics Study (SH-DREAMS). PloS One. 2013; 8: e61053. https://doi.org/10.1371/journal.pone.0061053.

- [13] Katulanda P, Ranasinghe P, Jayawardena R, Constantine GR, Sheriff MHR, Matthews DR. The prevalence, patterns and predictors of diabetic peripheral neuropathy in a developing country. Diabetology & Metabolic Syndrome. 2012; 4: 21. https://doi.org/10.1186/1758-5996-4-21.
- [14] Bansal D, Gudala K, Muthyala H, Esam HP, Nayakallu R, Bhansali A. Prevalence and risk factors of development of peripheral diabetic neuropathy in type 2 diabetes mellitus in a tertiary care setting. Journal of Diabetes Investigation. 2014; 5: 714–721. https://doi.org/10.1111/jdi.12223.
- [15] Al Washali A, Azuhairi A, Hejar A, Amani Y. Prevalence and associated risk factors of diabetic peripheral neuropathy among diabetic patients in national center of diabetes in Yemen. International Journal of Public Health and Clinical Sciences. 2014; 1: 141–150.
- [16] Khawaja N, Abu-Shennar J, Saleh M, Dahbour SS, Khader YS, Ajlouni KM. The prevalence and risk factors of peripheral neuropathy among patients with type 2 diabetes mellitus; the case of Jordan. Diabetology & Metabolic Syndrome. 2018; 10: 8. https://doi.org/10.1186/s13098-018-0309-6.
- [17] Owolabi MO, Ipadeola A. Total vascular risk as a strong correlate of severity of diabetic peripheral neuropathy in Nigerian Africans. Ethnicity & Disease. 2012; 22: 106–112.
- [18] Yeboah K, Agyekum JA, Owusu Mensah RNA, Affrim PK, Adu-Gyamfi L, Doughan RO, et al. Arterial Stiffness Is Associated with Peripheral Sensory Neuropathy in Diabetes Patients in Ghana. Journal of Diabetes Research. 2018; 2018: 2320737. https://doi.org/10.1155/2018/2320737.
- [19] Ching SM, Lee KW, Yusof Khan AHK, Devaraj NK, Cheong AT, Yap SF, et al. Prevalence and factors associated with peripheral neuropathy in a setting of retail pharmacies in Malaysia-A cross-sectional study. PloS One. 2024; 19: e0307093. https://doi.org/10.1371/journal.pone.0307093.
- [20] Salih MH, Tsega SS, Alemayehu BT, Ferede AJ. Peripheral neuropathy and associated factors among type 2 diabetic patients attending referral hospitals in the Amhara region, a multi-center cross-sectional study in Ethiopia. Scientific Reports. 2024; 14: 19662. https://doi.org/10.1038/s41598-024-70314-9.
- [21] Worku D, Hamza L, Woldemichael K. Patterns of diabetic complications at jimma university specialized hospital, southwest ethiopia. Ethiopian Journal of Health Sciences. 2010; 20: 33–39. https://doi.org/10.4314/ejhs.v20i1.69424.
- [22] Won JC, Park TS. Recent Advances in Diagnostic Strategies for Diabetic Peripheral Neuropathy. Endocrinology and Metabolism (Seoul, Korea). 2016; 31: 230–238. https://doi.org/10.3803/En M.2016.31.2.230.
- [23] Ziegler D, Fonseca V. From guideline to patient: a review of recent recommendations for pharmacotherapy of painful diabetic neuropathy. Journal of Diabetes and its Complications. 2015; 29: 146–156. https://doi.org/10.1016/j.jdiacomp.2014.08.008.
- [24] Van Dam PS, Cotter MA, Bravenboer B, Cameron NE. Pathogenesis of diabetic neuropathy: focus on neurovascular mechanisms. European Journal of Pharmacology. 2013; 719: 180–186. https://doi.org/10.1016/j.ejphar.2013.07.017.
- [25] Cameron NE, Eaton SE, Cotter MA, Tesfaye S. Vascular factors and metabolic interactions in the pathogenesis of diabetic neuropathy. Diabetologia. 2001; 44: 1973–1988. https://doi.org/10. 1007/s001250100001.
- [26] Negi G, Nakkina V, Kamble P, Sharma SS. Heme oxygenase-1, a novel target for the treatment of diabetic complications: focus on diabetic peripheral neuropathy. Pharmacological Research. 2015; 102: 158–167. https://doi.org/10.1016/j.phrs.2015.09. 014.
- [27] Pieczyńska J, Grajeta H. The role of selenium in human conception and pregnancy. Journal of Trace Elements in Medicine and Biology: Organ of the Society for Minerals and Trace Elements



- (GMS). 2015; 29: 31–38. https://doi.org/10.1016/j.jtemb.2014.
- [28] Kieliszek M, Błażejak S. Current Knowledge on the Importance of Selenium in Food for Living Organisms: A Review. Molecules (Basel, Switzerland). 2016; 21: 609. https://doi.org/10.3390/molecules21050609.
- [29] Xu H, Wen Q, Ye M. Correlating Blood Selenium Levels in Type 2 Diabetes Mellitus with Peripheral Neuropathy and Factors Contributing to Associated Lesion Development. Alternative Therapies in Health and Medicine. 2024; 30: 180–184.
- [30] Ayaz M, Kaptan H. Effects of selenium on electrophysiological changes associated with diabetic peripheral neuropathy. Neural Regeneration Research. 2011; 6: 617–622.
- [31] Ceko MJ, O'Leary S, Harris HH, Hummitzsch K, Rodgers RJ. Trace Elements in Ovaries: Measurement and Physiology. Biology of Reproduction. 2016; 94: 86. https://doi.org/10.1095/biolreprod.115.137240.
- [32] Kieliszek M, Błażejak S, Bzducha-Wróbel A. Influence of Selenium Content in the Culture Medium on Protein Profile of Yeast Cells Candida utilis ATCC 9950. Oxidative Medicine and Cellular Longevity. 2015; 2015: 659750. https://doi.org/10.1155/2015/659750.
- [33] Lavu RVS, Van De Wiele T, Pratti VL, Tack F, Du Laing G. Selenium bioaccessibility in stomach, small intestine and colon: Comparison between pure Se compounds, Se-enriched food crops and food supplements. Food Chemistry. 2016; 197: 382–387. https://doi.org/10.1016/j.foodchem.2015.08.001.
- [34] Fisinin VI, Papazyan TT, Surai PF. Producing selenium-enriched eggs and meat to improve the selenium status of the general population. Critical Reviews in Biotechnology. 2009; 29: 18–28. https://doi.org/10.1080/07388550802658030.
- [35] Khosravi-Darani K, Shojaosadati S. Solid State Fermentation of Apple Pomace for Production of Ethanol by Saccharomyces. Modares Journal of Electrical Engineering. 2002; 10: 55–60.
- [36] Chasteen TG, Bentley R. Biomethylation of selenium and tellurium: microorganisms and plants. Chemical Reviews. 2003; 103: 1–26. https://doi.org/10.1021/cr010210+.
- [37] Yoshinaga M, How S, Blanco D, Murdoch IS, Grudny M, Powers SL, *et al.* Directed Evolution of *Saccharomyces cerevisiae* for Increased Selenium Accumulation. Microorganisms. 2018; 6: 81. https://doi.org/10.3390/microorganisms6030081.
- [38] Rahimi P, Farshbaf-Khalili A, Shahnazi M, Ostarahimi A, Mohammadi M. Comparing the Effects of Selenium-Enriched Yeast and Sodium Selenite Supplementation on Postpartum Depression and Sexual Satisfaction: A Triple-Blind Controlled Clinical Trial. Journal of Caring Sciences. 2024; 13: 197–206. https://doi.org/10.34172/jcs.33218.
- [39] Zahedi Avval F, Mahmoudi N, Nosrati Tirkani A, Jarahi L, Hamidi Alamdari D, Sadjadi SA. Determining Pro-Oxidant Antioxidant Balance (PAB) and Total Antioxidant Capacity (TAC) in Patients with Schizophrenia. Iranian Journal of Psychiatry. 2018; 13: 222–226.
- [40] Ahrary Z, Khosravan S, Alami A, Najafi Nesheli M. The effects of a supportive-educational intervention on women with type 2 diabetes and diabetic peripheral neuropathy: a randomized controlled trial. Clinical Rehabilitation. 2020; 34: 794–802. https://doi.org/10.1177/0269215520914067.
- [41] Herman WH, Pop-Busui R, Braffett BH, Martin CL, Cleary PA, Albers JW, et al. Use of the Michigan Neuropathy Screening Instrument as a measure of distal symmetrical peripheral neuropathy in Type 1 diabetes: results from the Diabetes Control and Complications Trial/Epidemiology of Diabetes Interventions and Complications. Diabetic Medicine: a Journal of

- the British Diabetic Association. 2012; 29: 937–944. https://doi.org/10.1111/j.1464-5491.2012.03644.x.
- [42] Perkins BA, Olaleye D, Zinman B, Bril V. Simple screening tests for peripheral neuropathy in the diabetes clinic. Diabetes Care. 2001; 24: 250–256. https://doi.org/10.2337/diacare.24.2.250.
- [43] Larson JH, Anderson SM, Holman TB, Niemann BK. A longitudinal study of the effects of premarital communication, relationship stability, and self-esteem on sexual satisfaction in the first year of marriage. Journal of Sex & Marital Therapy. 1998; 24: 193–206. https://doi.org/10.1080/00926239808404933.
- [44] Shams Mofaraheh Z, Shahsiah M, Mohebi S, Tabaraee Y. The effect of marital counseling on sexual satisfaction of couples in Shiraz city. Journal of Health System Research. 2011; 6: 0–0.
- [45] Karunakaran U, Park KG. A systematic review of oxidative stress and safety of antioxidants in diabetes: focus on islets and their defense. Diabetes & Metabolism Journal. 2013; 37: 106– 112. https://doi.org/10.4093/dmj.2013.37.2.106.
- [46] Song J, Lin H. Experimental study on promotion of peripheral nerve regeneration by selenium-methylselenocysteine. Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi = Zhongguo Xiufu Chongjian Waike Zazhi = Chinese Journal of Reparative and Reconstructive Surgery. 2024; 38: 598–607. https://doi.org/10.7507/1002-1892.202402031.
- [47] Nazıroğlu M, Öz A, Yıldızhan K. Selenium and Neurological Diseases: Focus on Peripheral Pain and TRP Channels. Current Neuropharmacology. 2020; 18: 501–517. https://doi.org/ 10.2174/1570159X18666200106152631.
- [48] Altuhafi A, Altun M, Hadwan MH. The Correlation between Selenium-Dependent Glutathione Peroxidase Activity and Oxidant/Antioxidant Balance in Sera of Diabetic Patients with Nephropathy. Reports of Biochemistry & Molecular Biology. 2021; 10: 164–172. https://doi.org/10.52547/rbmb.10.2.164.
- [49] Henriksen EJ. Role of oxidative stress in the pathogenesis of insulin resistance and type 2 diabetes. In Bioactive food as dietary interventions for diabetes (pp. 3–17). Academic Press: CA, USA. 2019. https://doi.org/10.1016/B978-0-12-813822-9. 00001-1.
- [50] Likidlilid A, Patchanans N, Peerapatdit T, Sriratanasathavorn C. Lipid peroxidation and antioxidant enzyme activities in erythrocytes of type 2 diabetic patients. Journal of the Medical Association of Thailand = Chotmaihet Thangphaet. 2010; 93: 682–693.
- [51] Low PA. Oxidative stress and excitatory neurotoxins in neuropathy. In Peripheral Neuropathy: 2-Volume Set with Expert Consult Basic (pp. 509–533). Elsevier: Amsterdam, Netherlands. 2005
- [52] Huo J, Xue Y, Dong X, Lv J, Wu L, Gao H, et al. Efficacy of vitamin and antioxidant supplements for treatment of diabetic peripheral neuropathy: systematic review and meta-analysis of randomized controlled trials. Nutritional Neuroscience. 2023; 26: 778–795. https://doi.org/10.1080/1028415X.2022.2090606.
- [53] Aydın B, Nazıroğlu M. Involvement of TRPM7 Channel on the Induction of Diabetic Neuropathic Pain in Mice: Protective Role of Selenium and Curcumin. Biological Trace Element Research. 2023; 201: 2377–2395. https://doi.org/10.1007/ s12011-022-03518-7.
- [54] Jafari D, Eslami SS, Malih S, Tarighi P. Protective effect of selenium on vincristine-induced peripheral neuropathy in PC12 cell line. Cytotechnology. 2022; 74: 539–547. https://doi.org/10.1007/s10616-022-00543-1.
- [55] Teimouri A, Shivanian V, Elist JJ. A Double-Blind, Placebo-Controlled Study of the Effectiveness of Mate Endurance TM Dietary Supplement on Sexual Satisfaction, Ejaculatory Control, and Distress in Men with Premature Ejaculation. Open Journal of Urology. 2020; 10: 275–283.

