


## Research Article

# Comparative Study of Natural *Aloe vera* Gel, Sitagliptin and Glibenclamide as Diabetes Medications on Streptozotocin-Induced Male Rat Models

Ahlam A. Harasani<sup>1</sup>, Rokayya Sami<sup>2,\*</sup>, Rasha A. Al-Eisa<sup>3</sup>, Reham M. Algheshairy<sup>4</sup>,  
Hend F. Alharbi<sup>4</sup>, Abeer G. Almasoudi<sup>5</sup>, Suzan A. Abushal<sup>5</sup>, Siraj B. AlHarthi<sup>6</sup>,  
Suad H. Almasoudi<sup>7</sup>, Sameer H. Qari<sup>8</sup>

<sup>1</sup>Department of Biological Sciences, College of Science, University of Jeddah, 21959 Jeddah, Saudi Arabia

<sup>2</sup>Department of Food Science and Nutrition, College of Sciences, Taif University, 21944 Taif, Saudi Arabia

<sup>3</sup>Department of Biology, College of Sciences, Taif University, 21944 Taif, Saudi Arabia

<sup>4</sup>Department of Food Science and Human Nutrition, College of Agriculture and Food, Qassim University, 51452 Buraydah, Saudi Arabia

<sup>5</sup>Program of Food Sciences and Nutrition, Turabah University College, Taif University, 21944 Taif, Saudi Arabia

<sup>6</sup>Molecular Diagnostic Unit, Alhada Armed Forces Hospital, 26792 Taif, Saudi Arabia

<sup>7</sup>Department of Biology, College of Sciences, Umm Al-Qura University, 21955 Makkah, Saudi Arabia

<sup>8</sup>Department of Biology, Al-Jumum University College, Umm Al-Qura University, 21955 Makkah, Saudi Arabia

\*Correspondence: [rokayya.d@tu.edu.sa](mailto:rokayya.d@tu.edu.sa) (Rokayya Sami)

Academic Editor: Mehmet Ozaslan

Submitted: 16 March 2025   Revised: 8 April 2025   Accepted: 14 August 2025   Published: 8 January 2026

## Abstract

**Background and Objective:** *Aloe vera* gel is well-known as a beauty product, nutrient and herbal medicine due to its therapeutic properties. The current research aimed to make a comparative study of natural *Aloe vera* gel (AG), sitagliptin (SIAG) and glibenclamide (GLIB) as diabetes medications on streptozotocin (STZ)-induced albino Wistar male rat models. **Materials and Methods:** Rat models (60) were grouped equally into control, diabetic, normal+AG, STZ+AG, STZ+GLIB and STZ+SIAG. After 3 weeks of treatments, body weights, fasting blood glucose levels, blood analyses, fat profile and biochemical and antioxidant enzymes were evaluated. Data were analyzed in SPSS and all values are reported as Mean  $\pm$  SD. The *post hoc* multiple comparison tests revealed significant differences among rat groups at  $p \leq 0.05$ . **Results:** The oral administration of *Aloe vera* gel was efficient in the STZ+AG group compared to STZ+GLIB and STZ+SIAG. Treatments of the STZ+AG group caused a remarkable reduction of fasting blood glucose level 100.01 mg/dL. The STZ+AG group and STZ+GLIB group detected the same hemoglobin and glycosylated hemoglobin values. The STZ+GLIB group reported the lowest values for Homeostatic Model Assessment of Insulin Resistance (HOMA IR and  $\beta$  cell) 46.73 and 474.02, respectively. **Conclusion:** The effect of *Aloe vera* gel was more pronounced for the biochemical and antioxidant enzymes. These results involved that, when compared to sitagliptin and glibenclamide as diabetes medications, *Aloe vera* gel ingestion significantly enhanced the diabetic rat' antioxidant defense system.

**Keywords:** *Aloe vera*; streptozotocin-induced diabetes; antidiabetic activity; diabetes medications; antioxidant enzymes

## 1. Introduction

The diverse condition known as diabetes is typified by hyperglycemia, which has been accomplished by epidemic proportions in the current century [1]. It is one of the major healthcare problems of mortality worldwide [2]. Numerous complications, such as diabetic retinopathy, sorbitol pathway metabolic disorders, autooxidation of glycosylated proteins and glycation of body proteins, can occur from hyperglycemia [3]. These complications can lead to secondary problems that affect membrane damage and oxidation of cellular lipids in the kidneys, nerves and arteries [4]. Several medications such as biguanides, sulfonylureas sitagliptin and glibenclamide are well-known to lower hyperglycemia in diabetic patients [5]. Some of these drugs are expensive with poor availability for numerous countryside populations and developing countries

[6]. However, such medications can have some side effects [7]. There is an essential need in the medical system for the identification of antidiabetic plants, herbal medicines, natural compounds and alternative medications which shall have the therapeutic potential to overcome and reduce those side effects [8]. Numerous plant products and herbs have been demonstrated to have hypoglycemic effects [9]. *Aloe vera* (L.) plant has many medical benefits against stomach ailments, influenza, constipation, psoriasis, burns, lichen planus, healing wounds, plaque increase, gingivitis, gastrointestinal, ulcer and intestinal problems [10]. At present, it is utilized in nutraceuticals, skin care products and cosmetics [11]. It is considered an oral hypoglycaemic compound to reduce blood sugar levels and regulate plasma lipid levels [12]. Ghannam *et al.* [13] reported the positive effects of *Aloe* gel to lower blood glucose levels in



the streptozotocin (STZ) induced type-II diabetic animal model. The STZ may cause diabetes by selectively degenerating pancreatic B-cells [14]. In diabetic rat models, Noor *et al.* [15] reported that *Aloe vera* restored the bulk and function of pancreatic islet cells, demonstrating the protective benefits of the plant on the organ. Additionally, from *Aloe vera* extracts, a water-soluble compound with *Dipeptidyl Peptidase-IV* inhibitory action was extracted and some of their constituents can alter the expression of *Glucose Transporter Type 4* [16–18].

Nevertheless, more thorough research is required to determine the safety and effectiveness of plant extracts. In the current investigation, natural *Aloe vera* gel, sitagliptin and glibenclamide as diabetes medications on streptozotocin-induced male rat models' comparison with healthy and diabetic groups were studied.

## 2. Materials and Methods

### 2.1 Aloe Vera Plant and Extraction

Specimens *Aloe vera* (L.) plant, commonly known as *Aloe vera*, was cultivated and purchased in June, 2024 from a garden in Taif, Kingdom of Saudi Arabia. In this investigation, fresh leaves from this cultivated plant were used. The *Aloe vera* powder was prepared according to the protocol mentioned by Can *et al.* [9]. In brief, approximately 70 cm length of mature, healthy and fresh green leaves were cut transversely into several parts and then washed thoroughly with clean tap water and air-dried after the thick epidermis removal. The fleshy, colorless, semi-solid mucilaginous gel in the center of the leaves was extracted with the help of a sterilized knife and cut into small parts. The homogenate, which was thick and straw in color, was lyophilized, which was extracted with ethanol (95%) and water, filtrated, collected and freeze-dried by (ALPHA 1-4 LSC, Germany) to dryness. A non-metallic electric grinder (JH-E01, China) was used to crush the dried gel into powder and it was sieved through 300  $\mu\text{m}$ . The gel powder was stored in dry sterilized vacuumed tubes and kept at 4 °C until further use.

### 2.2 Animal Study

Approximately 3 month old male albino 150–200 g Wistar strain rats were employed. Rats were obtained from the Animal House in Jeddah, Kingdom of Saudi Arabia and selected as the animal models. Rats were maintained under the optimal laboratory conditions at 25 °C, 12 hrs light/dark cycle and had unlimited access to rat chow and tap water. The Gulf Countries Association of Sciences in Animal Experimentation granted Ethics Committee Number (8901/3151) approval for the 30 days animal study. Streptozotocin (STZ) a glucose analogue (2-deoxy-2-(3-methyl-3-nitrosouredio)-D-glucopyranose), sitagliptin and glibenclamide was obtained from Sigma, USA. All other chemicals, reagents and kits used were of analytical grade.

### 2.3 Experimental Procedure

Fig. 1 shows the design of the protocol and the treatment schedule. After acclimatization for a week for any observation of untoward symptoms, the rat models were randomly distributed into six main groups of ten rats each in separate cages as follows:

- **Vehicle group:** Healthy rat models.
- **Diabetic group:** Diabetic rat models Type 1 (induced with 30 mg/kg of STZ).
- **Normal+*Aloe vera* gel (AG) group:** Healthy rat models were given *Aloe vera* gel extract (300 mg/kg) daily by using gavage needles for 21 days.
- **STZ+AG group:** Diabetic rat models were given *Aloe vera* gel extract (300 mg/kg b.wt.) daily by using gavage needles for 21 days.
- **STZ+glibenclamide (GLIB) group:** Diabetic rat models were given glibenclamide (600  $\mu\text{g/kg}$ ) daily by using gavage needles for 21 days.
- **STZ+sitagliptin (SIAG) group:** Diabetic rat models were given sitagliptin (10 mg/kg) daily by using gavage needles for 21 days.

### 2.4 Induction of Diabetes

After the first seven days of the examination of rat models, the weight of rats was recorded after overnight fasting. Consequently, approximately 30 mg/kg of STZ in citrate buffer (0.1 M) with acidity (pH 4.5) was managed by a single intraperitoneal injection. The vehicle group of healthy rat models received an equal intraperitoneal dose of citrate buffer [19]. Blood glucose was assessed following a 16 hrs fast with free access to tap water. Overnight, the rats were given a 5% glucose solution to help them overcome the hypoglycemia caused by the medication. A glucometer (Bionime®, Switzerland) was used to measure fasting blood glucose levels and confirm the hyperglycemia before and 24 hrs following injection for diabetes approval and 7 days following injection for diabetes consolidation from the tails of the rat models. Fasting blood glucose levels were detected at weekly intervals on the 7th and 14th day with the tail prick assay. The body weight of each rat was then recorded every on the (1st, 7th, 14th and 21st) day by an electronic balance. The current study included STZ-treated animals that were diagnosed with diabetes if their fasting plasma levels were greater than 200 mg/dL with glucosuria. In order to prepare the *Aloe vera* gel extract, 300 mg of powdered *Aloe vera* gel was combined with 5 mL of water to create a solution. After the standardizing of the STZ dose, the experiments were conducted on animal groups to see the effect of natural *Aloe vera* gel and other diabetes medications, such as sitagliptin and glibenclamide, on rat models. Rat models in all groups were treated orally by using gavage needles every day in a single dose for 21 days.



**Fig. 1. Design of the protocol and the treatment schedule.** AG, *Aloe vera* gel; STZ, streptozotocin; GLIB, glibenclamide; SIAG, sitagliptin.

## 2.5 Administration of Samples

After a continuous three-week treatment period, the animals were fasted overnight to reduce experimental variability and normalize any possible changes in feeding behavior. The purpose of this fasting period was to improve the consistency of experimental outcomes by preventing recent food intake from influencing metabolic and physiological parameters. Necropsy was carried out right after euthanasia of thiopental sodium (40 mg/kg) in order to gather the organs and tissues required for further examination [20].

## 2.6 Blood Determinations

Blood samples were drawn from the tail vein following 24 hrs fasting with unrestricted access to water. A sharp, sterilized blade was used to cut the tail tip in a single stroke. Heparinized tubes were used to collect blood, the plasma was separated from the blood to detect the essential measurements by centrifuging it for 15 min at 2500 rpm. Approximately, 40  $\mu$ L was taken to measure glucose concentration according to the glucose oxidase assay (Reflotron 1, Boehringer, Germany). While hemoglobin and glycosy-

lated hemoglobin were detected [21]. Homeostatic Model Assessment of Insulin Resistance (HOMA IR and  $\beta$  cell) functions were detected [22]. The Dipeptidyl Peptidase-4 (DPP-4) enzyme activity was evaluated against the pNA standard graph after the incubation for 30 min at 37  $^{\circ}$ C to release the p-Nitroaniline. DPP IV enzyme activity was calculated as (U/min/mL) after absorbance at 405 nm by using a thermo scientific microplate (357) [23].

## 2.7 Fat Profile Determinations

The fat profile of blood, such as triglycerides (TG) and total cholesterol, was determined by the enzymatic colorimetric assays. The High-Density Lipoprotein (HDL) and Very Low-Density Lipoprotein (VLDL) of cholesterol were detected [24].

## 2.8 Biochemical and Antioxidant Enzyme Determinations

The biochemical and antioxidant enzyme evaluations for plasma and liver samples were detected [20]. Kits for blood and liver markers were purchased from Sigma, Darmstadt, Germany. Alkaline phosphatase (ALP) and Ala-

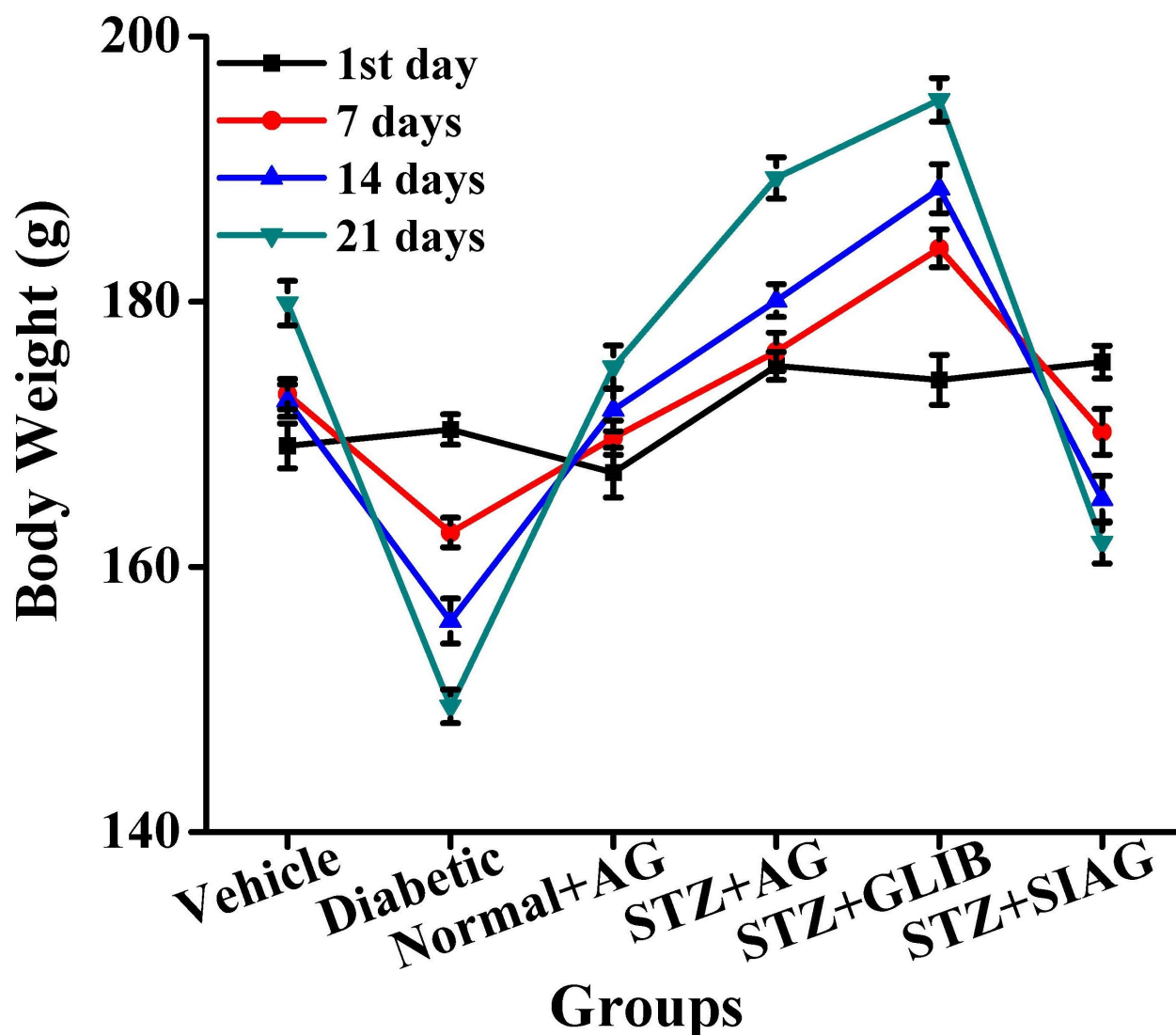


Fig. 2. Effects of various medications on body weights in rat models.  $n = 60$ ; each group  $n = 10$ .

nine Transaminase (ALT) were expressed as (U/L); Glutathione (GSH) and malondialdehyde (MDA) were expressed as (nmol/mg); Thiobarbituric Acid Reactive Substances (TBARS) and hydroperoxides were expressed as (mM/100 g); while antioxidant enzyme activities as glutathione peroxidase (GPx), superoxide dismutase (SOD), catalase (CAT) and glutathione-S-transferase (GST) were expressed as (U/mg).

### 2.9 Statistical Analysis

ANOVA test was used as the primary statistical analysis to compare differences between groups, followed by post-hoc test (Bonferroni) to identify specific group differences, with significance set at  $p < 0.05$ . The SPSS program version 26.0 (IBM Corp., Armonk, NY, USA) was applied for the statistical analysis. The results for all rat groups were presented as Mean  $\pm$  SD.

## 3. Results

### 3.1 Body Weights in Rat Models

Body weights were severely decreased in the diabetic group and STZ+SIAG group compared to the vehicle group on the 21st day (149.48, 161.84 and 179.89 g), respectively. Increases in body weights were improved by STZ+AG group or STZ+GLIB group treatments (189.33 g and 195.22 g) (Fig. 2).

### 3.2 Fasting Blood Glucose Levels in Rat Models

Fasting blood glucose levels were increased during the 21 days of post-diabetic inductions after the injection of the STZ dose. The level of the STZ+AG group consequently dropped down to reach 128.14 mg/dL on the 7th day, which further decreased to normal levels on the 14th and 21st day. Treatments of the STZ+AG group caused a remarkable reduction level of 100.01 mg/dL compared to the diabetic group 277.13 mg/dL (Fig. 3). On the other hand, the other



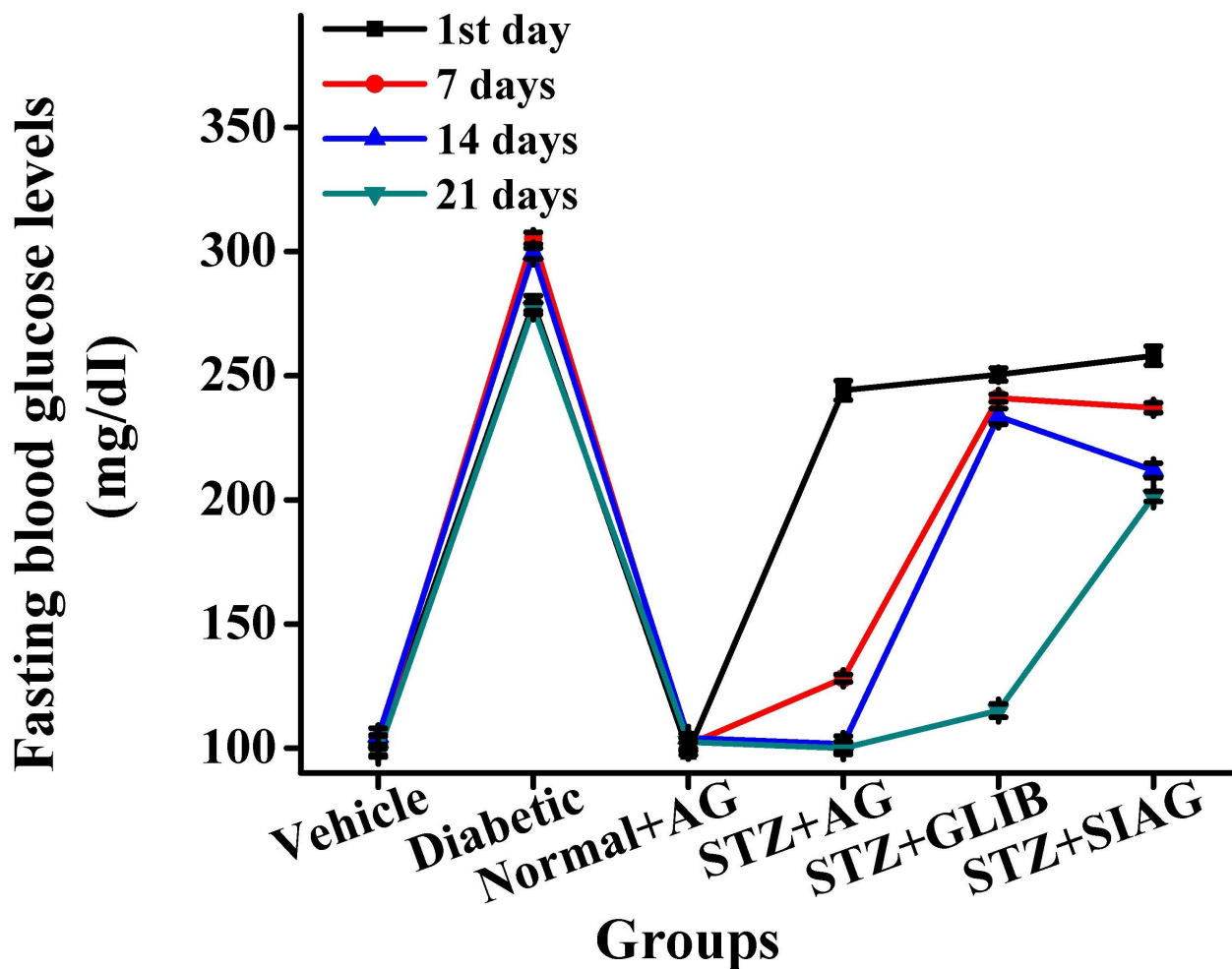


Fig. 3. Effects of various medications on fasting blood glucose levels in rat models.  $n = 60$ ; each group  $n = 10$ .

Table 1. Effects of various medications on HOMA IR and  $\beta$  cell functions in rat models.

Group	HOMA IR	HOMA $\beta$
Vehicle	$74.44 \pm 2.02^a$	$1010.02 \pm 20.13^a$
Diabetic	$58.76 \pm 2.07^b$	$955.32 \pm 20.89^b$
Normal+AG	$53.12 \pm 0.23^c$	$889.08 \pm 1.97^e$
STZ+AG	$49.03 \pm 1.91^d$	$600.07 \pm 21.14^c$
STZ+GLIB	$46.73 \pm 1.16^d$	$474.02 \pm 21.89^d$
STZ+SIAG	$48.46 \pm 0.21^d$	$488.76 \pm 10.54^d$

Different letters mean significant differences among rat models group. HOMA IR, Homeostatic Model Assessment of Insulin Resistance.

two diabetes medications sitagliptin and glibenclamide as did not show the ability to decrease the fasting blood glucose levels 201.42 and 115.14 mg/dL after 3 weeks compared with the vehicle group, respectively.

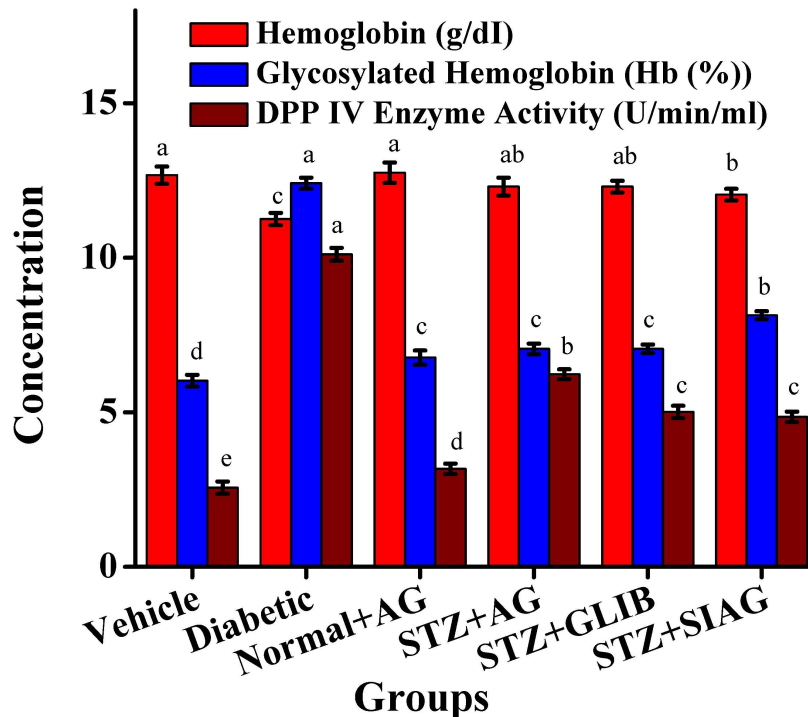
### 3.3 Blood Analysis in Rat Models

Hemoglobin levels rise in a glucose level-dependent manner in response to the severity of hyperglycemia. After 21 days, the administration of natural *Aloe vera* gel and other diabetes medications such as sitagliptin and glibenclamide (STZ+SIAG group) caused a reduction in hemoglobin level (12.04 g/dL), while STZ+AG group and STZ+GLIB group detected the same value and tended to reach the vehicle group (12.3 g/dL) compared to the diabetic group (11.25 g/dL) (Fig. 4). On the same trend, STZ+AG group and STZ+GLIB group detected the same value of glycosylated hemoglobin and tended to reach the vehicle group (7.05% Hb) compared to the diabetic group (12.41% Hb), while STZ+SIAG group had higher values (8.14% Hb) at the end of the experimental period (Table 1). The DPP IV enzyme activity demonstrated an increase in diabetic rat models from 4.85 U/min/mL in the STZ+SIAG group to 10.11 U/min/mL in the diabetic group compared to the vehicle group recorded 2.56 U/min/mL. While administration of glibenclamide decreased the DPP IV enzyme activity to reach 5.01 U/min/mL followed by the STZ+AG

**Table 2. Effects of various medications on fat profile in rat models.**

Group	Triglycerides (mg/dL)	Total cholesterol (mg/dL)	HDL (mg/dL)	VLDL (mg/dL)
Vehicle	568.32 ± 3.89 <sup>b</sup>	76.13 ± 3.18 <sup>a</sup>	28.14 ± 3.58 <sup>a</sup>	82.34 ± 3.81 <sup>b</sup>
Diabetic	802.00 ± 3.16 <sup>a</sup>	74.22 ± 3.53 <sup>a</sup>	27.13 ± 3.76 <sup>a</sup>	115.41 ± 3.94 <sup>a</sup>
Normal+AG	510.75 ± 3.81 <sup>c</sup>	81.04 ± 3.34 <sup>a</sup>	29.62 ± 3.21 <sup>a</sup>	78.03 ± 3.52 <sup>b</sup>
STZ+AG	420.51 ± 3.94 <sup>d</sup>	76.96 ± 3.42 <sup>a</sup>	27.03 ± 3.71 <sup>a</sup>	59.12 ± 3.78 <sup>c</sup>
STZ+GLIB	289.40 ± 3.17 <sup>e</sup>	75.87 ± 3.86 <sup>a</sup>	21.45 ± 3.34 <sup>a</sup>	50.02 ± 3.73 <sup>d</sup>
STZ+SIAG	245.42 ± 3.34 <sup>f</sup>	83.30 ± 3.97 <sup>a</sup>	22.54 ± 3.85 <sup>a</sup>	56.32 ± 3.83 <sup>cd</sup>

Different letters mean significant differences among rat models group. HDL, High-Density Lipoprotein; VLDL, Very Low-Density Lipoprotein.



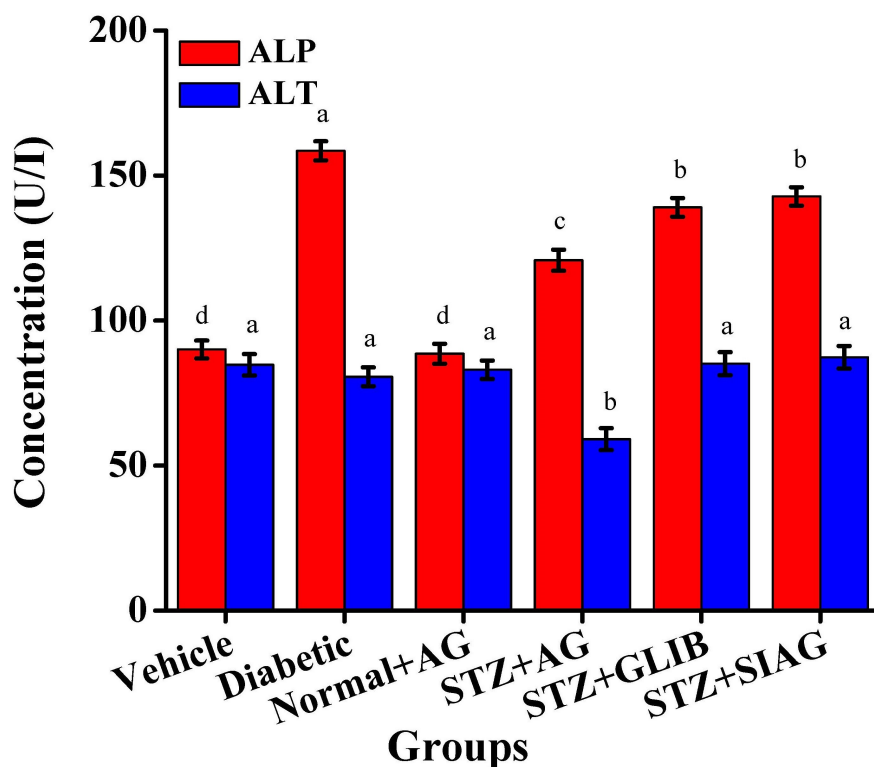
**Fig. 4. Effects of various medications on blood analysis in rat models.** Different letters mean significant differences among rat models group. Dipeptidyl Peptidase-4 (DDP IV), n = 60; each group n = 10.

group (6.23 U/min/mL). The increases in (HOMA IR and  $\beta$  cell) functions ranged as (74.44–1010.02) and (58.76–955.32) in both vehicle and diabetic groups, respectively. STZ+AG group detected a higher HOMA IR value (49.03) followed by the STZ+SIAG group which detected (48.46). On the same trend, the STZ+AG group detected a higher  $\beta$  cell value (600.07) followed by the STZ+SIAG group which detected (488.76) (Table 1). The STZ+GLIB group reported the lowest values for HOMA IR and  $\beta$  cell (46.73 and 474.02), respectively. Moreover, a clear reduction in HOMA  $\beta$  cells was detected in the normal+AG group to reach (889.08).

### 3.4 Fat Profile in Rat Models

The fat profile values among all rat models are presented in Table 2. An increase in TG and VLDL values in the diabetic group (802.00–115.41 mg/dL) compared with

that in the vehicle group (568.32–82.34 mg/dL), respectively. The STZ+AG group maintained TG and VLDL values (420.51–59.12 mg/dL) than the STZ+GLIB group (289.40–50.02 mg/dL) and STZ+SIAG group (245.42–56.32 mg/dL), respectively. The STZ+AG group preserved HDL value to reach 27.03 mg/dL compared with the vehicle group's 28.14 mg/dL. The STZ+SIAG group normal+AG group reported higher values of total cholesterol at 83.30 mg/dL followed by the normal+AG group, which reached 81.04 mg/dL. In the current work, supplementation of *Aloe vera* decreased the TG and VLDL levels; however, no obvious changes occurred in total cholesterol and HDL levels, indicating that lipid metabolism may be improved by gel. The highest values for TG and VLDL were noticed in the normal+AG group, which fed on *Aloe vera* only at 510.75 and 78.03 mg/dL, respectively (Table 2).



**Fig. 5.** Effects of various medications on liver tissue as Alkaline phosphatase (ALP) and Alanine Transaminase (ALT) activities in rat models. Different letters mean significant differences among rat models group, n = 60; each group n = 10.

### 3.5 Biochemical Evaluation in Rat Models

Some biochemical parameters for the liver functions were determined such as ALP and ALT activities (Fig. 5). The ALP activity was increased in the diabetic group, compared to the vehicle group. The ALT activity was reduced in the diabetic rat models given *Aloe vera* gel to reach 59.10 U/L. It was noticed that the STZ+AG group got protective effects comparable to STZ+GLIB and STZ+SIAG groups against hepatotoxicity. Fig. 6 presents liver tissue GSH and MDA values in the liver for all rat models. There was a great reduction in GSH values among diabetes rats compared to the vehicle group. The Treatments with *Aloe vera* and glibenclamide medication raised the GSH values to reach 32.57 and 29.03 nmol/mg, respectively. As shown in Fig. 6, the diabetic group detected an increase in MDA level 13.23 nmol/mg compared to the vehicle group 7.16 nmol/mg. It is known that higher levels of MDA are an indicator of increased oxidative stress. The administration of natural *Aloe vera* gel, sitagliptin and glibenclamide as diabetes medications to the diabetic rat models resulted in a great reduction of MDA levels as 9.11, 10.12 and 11.44 nmol/mg, respectively. Fig. 7a,b presents TBARS and hydroperoxides values in the liver of rat models. There were great elevations in tissue TBARS and hydroperoxides for diabetic rats (1.82 and 95.47 mM/100 g) compared to the vehicle group (0.90 and 70.05 mM/100 g), respectively. The administration of natural *Aloe vera* gel, glibenclamide

and sitagliptin as diabetes medications to the diabetic rat models resulted in great reductions as 0.95, 1.51, 1.49, 75.21, 82.04 and 87.92 mM/100 g, respectively.

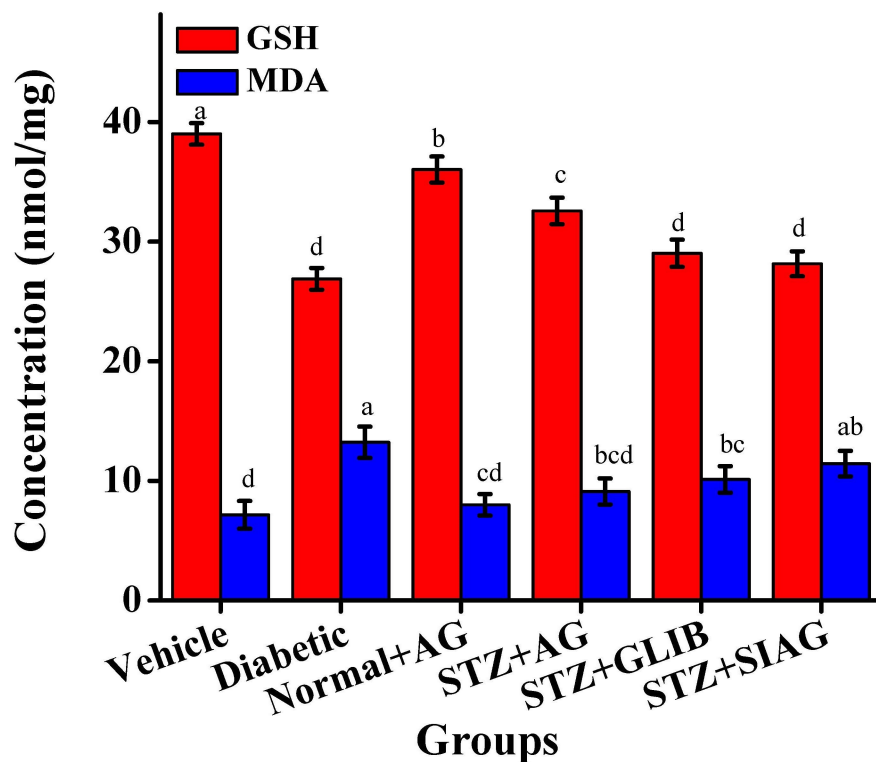
### 3.6 Antioxidant Enzymes in Rat Models

Table 3 presents the enzyme activities of GPx, SOD, CAT and GST in the liver tissues of rat models. The GPx activity recorded a considerable reduction from 1.74 U/mg in the diabetic group to 1.01, 1.17 and 1.20 U/mg for STZ+AG, STZ+GLIB and STZ+SIAG groups, respectively. The effect of *Aloe vera* gel was more pronounced which reached 6.17, 0.67 and 4.04 U/mg compared with the diabetic group that recorded (3.88, 0.48 and 1.23 U/mg) for SOD, CAT and GST activities, respectively.

## 4. Discussion

The diabetic rat models in STZ-induced diabetes may display the majority of problems, including oxidative stress-induced neurological, vas deferens, kidney and urine bladder dysfunctions [6,7].

These positive effects of body weight can be due to the prevention of  $\beta$ -cells death [18]. The greater calorie loss via glucose excretion in the urine is probably the cause of this differential body weight response [25]. Nayak and Pat-tabiraman [26] reported antidiabetic activities of *Aloe vera* helped to gain weight due to normoglycaemia.



**Fig. 6.** Effects of various medications on liver tissue as Glutathione (GSH) and malondialdehyde (MDA) in rat models. Different letters mean significant differences among rat models group, n = 60; each group n = 10.

**Table 3.** Effects of various medications on antioxidant enzymes in rat models.

Group	Gpx activity (U/mg)	SOD activity (U/mg)	CAT activity (U/mg)	GST activity (U/mg)
Vehicle	1.22 ± 0.57 <sup>a</sup>	5.33 ± 0.56 <sup>ab</sup>	0.53 ± 0.12 <sup>a</sup>	2.11 ± 0.33 <sup>b</sup>
Diabetic	1.74 ± 0.63 <sup>a</sup>	3.88 ± 0.76 <sup>b</sup>	0.48 ± 0.13 <sup>a</sup>	1.23 ± 0.29 <sup>c</sup>
Normal+AG	1.03 ± 0.73 <sup>a</sup>	6.12 ± 0.85 <sup>a</sup>	0.61 ± 0.21 <sup>a</sup>	2.23 ± 0.42 <sup>b</sup>
STZ+AG	1.01 ± 0.71 <sup>a</sup>	6.17 ± 0.72 <sup>a</sup>	0.67 ± 0.22 <sup>a</sup>	4.04 ± 0.32 <sup>a</sup>
STZ+GLIB	1.17 ± 0.65 <sup>a</sup>	4.77 ± 0.66 <sup>ab</sup>	0.63 ± 0.24 <sup>a</sup>	3.56 ± 0.31 <sup>a</sup>
STZ+SIAG	1.20 ± 0.69 <sup>a</sup>	5.03 ± 0.86 <sup>a</sup>	0.62 ± 0.23 <sup>a</sup>	3.72 ± 0.29 <sup>a</sup>

Different letters mean significant differences among rat models group. GPx, glutathione peroxidase; SOD, superoxide dismutase; CAT, catalase; GST, glutathione-S-transferase, n = 60; each group n = 10.

*Aloe vera* sterols are plentiful and structurally comparable to cholesterol, including stigmasterol, campesterol and b-sitosterol [25]. Furthermore, it regulates the enzymes that break down carbohydrates to preserve glucose homeostasis [27]. Another study compared oral medication of *Aloe vera* gel (200–400 mg/kg) with metformin (50 mg/kg) in rat models. All therapies decreased the high blood glucose levels in all diabetic-induced rats and there were no significant differences between them [28]. Another study on the alloxan-induced mice that used oral natural *Aloe vera* gel in a dose of 500 mg/kg twice a day reported a reduction in blood glucose levels from 30.7 to 37.7% on the 5th day [29].

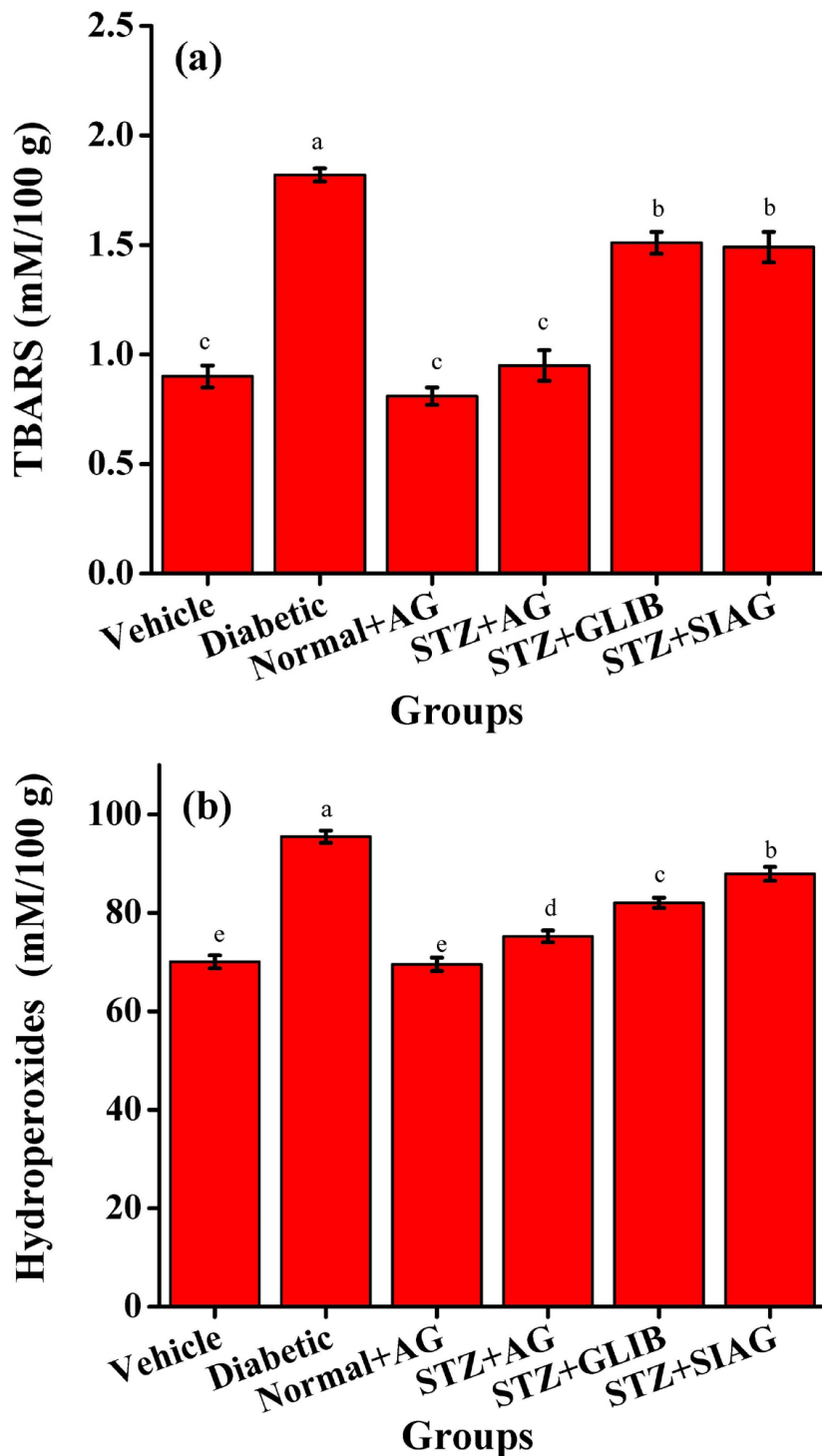
Mohamed [25] isolated phytosterol and fraction components from *Aloe vera* gel for studying the anti-hyperglycemic activities. Glycosylated hemoglobin is cre-

ated when hemoglobin and excess blood glucose combine in diabetes [30]. Glycosylated hemoglobin is increased over a long period in diabetes.

The reduction of glycosylated hemoglobin might be due to the enhancement in the glycemic status. This implies that the natural *Aloe vera* gel treatment will reduce oxidative stress. The DPP IV enzyme activity is known as an attractive anti-diabetic medication target [16]. The current data for the DPP IV enzyme activity was in agreement with Nayak and Pattabiraman [26], who demonstrated that *Aloe vera* gel may inhibit that enzyme and the dipyrrole derivative *in vitro* study. Those results referred to the protective roles of *Aloe vera* gel on the pancreatic IR and  $\beta$  cell functions due to the presence of antioxidant components [27].

As well-known cholesterol, elevated TG levels may contribute to the development of type 2 diabetes by patho-





**Fig. 7. Effects of various medications on liver tissue in rat models.** (a) Thiobarbituric Acid Reactive Substances (TBARS) and (b) Hydroperoxides, n = 60; each group n = 10.

logically impairing  $\beta$ -cell activity [24,31]. A study on *Aloe vera* gel on rat models reported similar values by reducing the TG, total cholesterol and lipase activities with a concomitant decrease in adipose tissues which is considered helpful for insulin [32].

Increased blood activity of certain enzymes, such as ALT and ALP, are common biochemical indicators of liver damage [33]. According to the current results, although ALP is a critical marker for various metabolic functions that might be impaired by diabetes, healthy compounds in *Aloe vera* gel may decrease ALP activity, considered a result of

the toxic metabolites produced by the streptozotocin. The GSH is primarily involved in metabolism and detoxification. As an antioxidant, it shields tissues from oxidative stress and is, therefore, frequently used as a gauge of free radical damage [34]. Reduced GSH levels in the liver with diabetes due to STZ injection indicate tissue damage and the increased oxidative stress-induced GSH consumption [35]. The usage of more “SH” in proteins to protect them from lipid peroxide may be the cause of the decrease in GSH values [36]. This suggests that the natural *Aloe vera* plant may either promote GSH biosynthesis, lessen oxidative stress, which prevents GSH from degrading or have both benefits. The MDA, a biomarker of oxidative stress, is produced when reactive oxygen species break down polyunsaturated fatty acids [19]. The reduction in MDA formation values in the liver was in agreement with Tanaka *et al.* [37], indicating the plant’s function in preventing oxidative damage caused by cellular damage. *Aloe vera* gel can reduce MDA values by raising the production of nitric oxide, which strengthens the antioxidant capacity as a way to regulate blood glucose levels [38]. The TBARS is the most widely used lipid peroxidation indicator [39]. The notable rise in TBARS and hydroperoxide values in the liver of diabetic rat models may be due to the elevated lipid peroxidation in the tissues [40].

Antioxidant enzymes play very essential roles in the defense system, while CAT is a heme protein that catalyzes the reduction of hydrogen peroxide and shields the tissue from extremely dangerous hydroxyl radicals, SOD can scavenge superoxide radicals by converting them to H<sub>2</sub>O<sub>2</sub> and molecular oxygen [20,24]. The GPx and GST enzymes can break down H<sub>2</sub>O<sub>2</sub> and other organic hydroperoxides into non-toxic byproducts at the GSH’s expense, while any inactivation and glycation of the enzyme caused by radicals may lead to decreased GPx activity [40]. These results involved that, when compared to sitagliptin and glibenclamide as diabetes medications, *Aloe vera* gel ingestion significantly enhanced the diabetic rats’ antioxidant defense system.

## 5. Conclusion

The results of the current work indicated that the oral administration of *Aloe vera* gel was sufficient and might have beneficial effects on body weight, blood analyses, fat profile and biochemical and antioxidant enzymes for the liver comparable to sitagliptin and glibenclamide against hepatotoxicity produced by streptozotocin for diabetic rat models. Extra research on the application of *Aloe vera* gel is needed in the treatment and/or prevention of diabetes complications.

## 6. Significance Statement

Glibenclamide, sitagliptin and natural *Aloe vera* gel have been used as diabetes medications in streptozotocin-induced male rat models. The present study concluded

that the oral administration of *Aloe vera* gel was adequate and could potentially have positive effects on body weight, blood analysis, fat profile, liver biochemical and antioxidant enzymes similar to sitagliptin and glibenclamide against streptozotocin-induced hepatotoxicity in diabetic rat models. In order to cure and/or prevent problems from diabetes, more research is required on the use of *Aloe vera* gel.

## Availability of Data and Materials

All relevant data are within the paper and its supporting information files.

## Author Contributions

Conception or design: AAH, RS, RAA, SHQ, AGA, SAA. Acquisition, analysis, or interpretation of data: RMA, HFA, SHA, SBA. Drafting the work or revising: SHQ, AGA, SAA, RMA, HFA, SHA, SBA. Final approval of the manuscript: AAH, RS, RAA. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work in the end.

## Ethics Approval and Consent to Participate

The Gulf Countries Association of Sciences in Animal Experimentation granted Ethics Committee Number (8901/3151) approval for the 30 days animal study. Every animal procedure was carried out in compliance with the 3R principles (replacement, reduction, and refinement) and the ethical standards of the institution’s guidelines. Additionally, the work followed the ARRIVE criteria for reporting research using animals.

## Acknowledgment

The authors extend their appreciation to Taif University, Saudi Arabia, for supporting this work through project number (TU-DSPP-2024-10).

## Funding

This research was funded by Taif University, Saudi Arabia, Project No. (TU-DSPP-2024-10).

## Conflict of Interest

The authors declare no conflict of interest.

## References

- [1] Abdelazez A, Abdelmotaal H, Evvie SE, Bikheet M, Sami R, Mohamed H, *et al.* Verification of *Lactobacillus brevis* tolerance to simulated gastric juice and the potential effects of postbiotic gamma-aminobutyric acid in streptozotocin-induced diabetic mice. *Food Science and Human Wellness*. 2022; 11: 165–176. <https://doi.org/10.1016/j.fshw.2021.07.017>.
- [2] Abdelazez A, Abdelmotaal H, Evvie SE, Melak S, Jia FF, Khoso MH, *et al.* Screening Potential Probiotic Characteristics

- of *Lactobacillus brevis* Strains In Vitro and Intervention Effect on Type I Diabetes In Vivo. *BioMed Research International*. 2018; 2018: 7356173. <https://doi.org/10.1155/2018/7356173>.
- [3] Al-Eisa RA, Helal M, Aljahani AH, Sami R, Banjer HJ, Algehainy NA, *et al.* Ochratoxin A oral mycotoxin and honey dietary intake effects on TNF- $\alpha$  immunology response, lactic acid bacteria microbial loads,  $\beta$ -glucuronidase enzyme activity, some hematological and biochemical parameters on mice. *Materials Express*. 2023; 13: 1203–1211. <https://doi.org/10.1166/mex.2023.2462>.
  - [4] Andallu B, Varadacharyulu NC. Antioxidant role of mulberry (*Morus indica* L. cv. Anantha) leaves in streptozotocin-diabetic rats. *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 2003; 338: 3–10. [https://doi.org/10.1016/s0009-8981\(03\)00322-x](https://doi.org/10.1016/s0009-8981(03)00322-x).
  - [5] Ajabnoor MA. Effect of aloes on blood glucose levels in normal and alloxan diabetic mice. *Journal of Ethnopharmacology*. 1990; 28: 215–220. [https://doi.org/10.1016/0378-8741\(90\)90031-n](https://doi.org/10.1016/0378-8741(90)90031-n).
  - [6] Baazeem A, Helal M, Sami R, Alshehry G, Algarni E, Hilary U, *et al.* Positive effects of dietary honey and aflatoxin B1 on serum enzymes, superoxide dismutase activity,  $\beta$ -glucuronidase enzyme activity, and colonic probiotic bacteria on rats. *Materials Express*. 2024; 14: 60–65. <https://doi.org/10.1166/mex.2024.2584>.
  - [7] Babu SN, Govindarajan S, Vijayalakshmi MA, Noor A. Role of zonulin and GLP-1/DPP-IV in alleviation of diabetes mellitus by peptide/polypeptide fraction of Aloe vera in streptozotocin-induced diabetic wistar rats. *Journal of Ethnopharmacology*. 2021; 272: 113949. <https://doi.org/10.1016/j.jep.2021.113949>.
  - [8] Begum K, Khan I, Wali A, Sami R, Aljumayi H, Abushal SA, *et al.* Buckwheat containing-bread: a scientific inquiry into insulin, polyphenols, antioxidants status, and oxidative stress markers in type-II diabetic individuals. *Frontiers in Sustainable Food Systems*. 2024; 8: 1440053. <https://doi.org/10.3389/fsufs.2024.1440053>.
  - [9] Can A, Akev N, Ozsoy N, Bolkent S, Arda BP, Yanardag R, *et al.* Effect of Aloe vera leaf gel and pulp extracts on the liver in type-II diabetic rat models. *Biological & Pharmaceutical Bulletin*. 2004; 27: 694–698. <https://doi.org/10.1248/bpb.27.694>.
  - [10] Chhabria S, Mathur S, Vadakan S, Sahoo DK, Mishra P, Paital B. A review on phytochemical and pharmacological facets of tropical ethnomedicinal plants as reformed DPP-IV inhibitors to regulate incretin activity. *Frontiers in Endocrinology*. 2022; 13: 1027237. <https://doi.org/10.3389/fendo.2022.1027237>.
  - [11] Deora N, Sunitha MM, Satyavani M, Harishankar N, Vijayalakshmi MA, Venkataraman K, *et al.* Alleviation of diabetes mellitus through the restoration of  $\beta$ -cell function and lipid metabolism by Aloe vera (*L.*) Burm. f. extract in obesogenic WNIN/GR-Ob rats. *Journal of Ethnopharmacology*. 2021; 272: 113921. <https://doi.org/10.1016/j.jep.2021.113921>.
  - [12] Drabkin DL, Austin JH. Spectrophotometric studies: I. Spectrophotometric constants for common hemoglobin derivatives in human, dog, and rabbit blood. *Journal of Biological Chemistry*. 1932; 98: 719–733. [https://doi.org/10.1016/S0021-9258\(18\)76122-X](https://doi.org/10.1016/S0021-9258(18)76122-X).
  - [13] Ghannam N, Kingston M, Al-Meshaal IA, Tariq M, Parman NS, Woodhouse N. The antidiabetic activity of aloes: preliminary clinical and experimental observations. *Hormone Research*. 1986; 24: 288–294. <https://doi.org/10.1159/000180569>.
  - [14] Hammeso WW, Emiru YK, Ayalew Getahun K, Kahaliw W. Antidiabetic and Antihyperlipidemic Activities of the Leaf Latex Extract of *Aloe megalacantha* Baker (Aloaceae) in Streptozotocin-Induced Diabetic Model. Evidence-based Complementary and Alternative Medicine: ECAM. 2019; 2019: 8263786. <https://doi.org/10.1155/2019/8263786>.
  - [15] Noor A, Gunasekaran S, Vijayalakshmi MA. Improvement of Insulin Secretion and Pancreatic  $\beta$ -cell Function in Streptozotocin-induced Diabetic Rats Treated with Aloe vera Extract. *Pharmacognosy Research*. 2017; 9: S99–S104. [https://doi.org/10.4103/pr.pr\\_75\\_17](https://doi.org/10.4103/pr.pr_75_17).
  - [16] Jomova K, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, Valko M. Several lines of antioxidant defense against oxidative stress: antioxidant enzymes, nanomaterials with multiple enzyme-mimicking activities, and low-molecular-weight antioxidants. *Archives of Toxicology*. 2024; 98: 1323–1367. <https://doi.org/10.1007/s00204-024-03696-4>.
  - [17] Kazeem MI, Bankole HA, Fatai AA, Saibu GM, Wusu AD. Genus Aloe as sources of antidiabetic, antihyperglycemic and hypoglycemic agents: A review. *South African Journal of Botany*. 2022; 147: 1070–1077. <https://doi.org/10.1016/j.sajb.2022.05.041>.
  - [18] Kim K, Kim H, Kwon J, Lee S, Kong H, Im SA, *et al.* Hypoglycemic and hypolipidemic effects of processed Aloe vera gel in a mouse model of non-insulin-dependent diabetes mellitus. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2009; 16: 856–863. <https://doi.org/10.1016/j.phymed.2009.02.014>.
  - [19] Kumar R, Sharma B, Tomar NR, Roy P, Gupta AK, Kumar A. *In vivo* evaluation of hypoglycemic activity of Aloe spp. and identification of its mode of action on GLUT-4 gene expression in vitro. *Applied Biochemistry and Biotechnology*. 2011; 164: 1246–1256. <https://doi.org/10.1007/s12010-011-9210-6>.
  - [20] Luo W, Chen X, Ye L, Chen X, Jia W, Zhao Y, *et al.* Kaempferol attenuates streptozotocin-induced diabetic nephropathy by downregulating TRAF6 expression: The role of TRAF6 in diabetic nephropathy. *Journal of Ethnopharmacology*. 2021; 268: 113553. <https://doi.org/10.1016/j.jep.2020.113553>.
  - [21] Lyons TJ. Oxidized low density lipoproteins: a role in the pathogenesis of atherosclerosis in diabetes? *Diabetic Medicine: a Journal of the British Diabetic Association*. 1991; 8: 411–419. <https://doi.org/10.1111/j.1464-5491.1991.tb01624.x>.
  - [22] Stanely Mainzen Prince P, Menon VP. Antioxidant action of *Tinospora cordifolia* root extract in alloxan diabetic rats. *Phytotherapy Research: PTR*. 2001; 15: 213–218. <https://doi.org/10.1002/ptr.707>.
  - [23] Marref SE, Melakhessou MA, Becheker I, Bensaad MS, Marref C, Mekersi N, *et al.* Pharmacological Evaluation of Antidiabetic Capacity of *Moringa oleifera* LAM Leaves. *Journal of Biobased Materials and Bioenergy*. 2024; 18: 404–409. <https://doi.org/10.1166/jbmb.2024.2391>.
  - [24] Melakhessou MA, Becheker I, Marref SE, Bensaad MS, Alyamani A, Sami R, *et al.* A Property-Response Approach to Evaluate Acute Toxicity Profile and Pharmacological Quality of Hydro-Alcoholic Extract of Walnut Root Bark *Juglans Regia* Linn. in Streptozotocin-Induced Diabetic Mode. *Journal of Biobased Materials and Bioenergy*. 2023; 17: 451–459. <https://doi.org/10.1166/jbmb.2023.2288>.
  - [25] Mohamed EAK. Antidiabetic, antihypercholesteremic and antioxidant effect of Aloe vera gel extract in alloxan induced diabetic rats. *Australian Journal of Basic and Applied Sciences*. 2011; 5: 1321–1327.
  - [26] Nayak SS, Pattabiraman TN. A new colorimetric method for the estimation of glycosylated hemoglobin. *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 1981; 109: 267–274. [https://doi.org/10.1016/0009-8981\(81\)90312-0](https://doi.org/10.1016/0009-8981(81)90312-0).
  - [27] Noor A, Gunasekaran S, Vijayalakshmi MA. Improvement of Insulin Secretion and Pancreatic  $\beta$ -cell Function in Streptozotocin-induced Diabetic Rats Treated with Aloe vera Extract. *Pharmacognosy Research*. 2017; 9: S99–S104. [https://doi.org/10.4103/pr.pr\\_75\\_17](https://doi.org/10.4103/pr.pr_75_17).
  - [28] Prasannaraja C, Kamalanathan AS, Vijayalakshmi MA, Venkataraman K. A dipyrrole derivative from *Aloe vera* inhibits

- an anti-diabetic drug target Dipeptidyl Peptidase (DPP)-IV *in vitro*. *Preparative Biochemistry & Biotechnology*. 2020; 50: 511–520. <https://doi.org/10.1080/10826068.2019.1710712>.
- [29] Rahoui W, Merzouk H, El Hachi IA, Bettoui R, Azzi R, Benali M. Beneficial effects of Aloe vera gel on lipid profile, lipase activities and oxidant/antioxidant status in obese rats. *Journal of Functional Foods*. 2018; 48: 525–532. <https://doi.org/10.1016/j.jff.2018.07.050>.
- [30] Raja CP, Venkataraman K. Aloe vera phytochemicals inhibits dipeptidyl peptidase iv (dpp-iv), an anti-diabetic target. *International Journal of Pharma and Bio Sciences*. 2016; 7: 120–128.
- [31] Reusser F. Mode of action of streptozotocin. *Journal of Bacteriology*. 1971; 105: 580–588. <https://doi.org/10.1128/jb.105.2.580-588.1971>.
- [32] Saidi A, Hambaba L, Bensaad MS, Kadi IE, Bensouici C, Sami R, *et al*. Phytochemical analysis using cLC-DAD, nutritional importance and assessment of antioxidant, antidiabetic and anticholinesterase activities of *Ruta tuberculata* Forssk organic extracts. *Sustainability*. 2022; 14: 10451. <https://doi.org/10.3390/su141610451>.
- [33] Kalas MA, Chavez L, Leon M, Taweese PT, Surani S. Abnormal liver enzymes: A review for clinicians. *World Journal of Hepatology*. 2021; 13: 1688–1698. <https://doi.org/10.4254/wjgh.v13.i11.1688>.
- [34] Vázquez-Meza H, Vilchis-Landeros MM, Vázquez-Carrada M, Uribe-Ramírez D, Matuz-Mares D. Cellular Compartmentalization, Glutathione Transport and Its Relevance in Some Pathologies. *Antioxidants (Basel)*. 2023; 12: 834. <https://doi.org/10.3390/antiox12040834>.
- [35] Singh RP, Dhanalakshmi S, Rao AR. Chemomodulatory action of Aloe vera on the profiles of enzymes associated with carcinogen metabolism and antioxidant status regulation in mice. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2000; 7: 209–219. [https://doi.org/10.1016/S0944-7113\(00\)80006-9](https://doi.org/10.1016/S0944-7113(00)80006-9).
- [36] Szkudelski T. The mechanism of alloxan and streptozotocin action in B cells of the rat pancreas. *Physiological Research*. 2001; 50: 537–546.
- [37] Tanaka M, Misawa E, Ito Y, Habara N, Nomaguchi K, Yamada M, *et al*. Identification of five phytosterols from Aloe vera gel as anti-diabetic compounds. *Biological & Pharmaceutical Bulletin*. 2006; 29: 1418–1422. <https://doi.org/10.1248/bpb.29.1418>.
- [38] Mohamed E A K. Antidiabetic, antihypercholestermic and antioxidative effect of Aloe vera gel extract in alloxan induced diabetic rats. *Australian Journal of Basic and Applied Sciences*. 2011; 5: 1321–1327.
- [39] Venu L, Harishankar N, Prasanna Krishna T, Raghunath M. Maternal dietary vitamin restriction increases body fat content but not insulin resistance in WNIN rat offspring up to 6 months of age. *Diabetologia*. 2004; 47: 1493–1501. <https://doi.org/10.1007/s00125-004-1506-4>.
- [40] Zagórska-Dziok M, Furman-Toczek D, Dudra-Jastrzębska M, Zygo K, Stanisławek A, Kapka-Skrzypczak L. Evaluation of clinical effectiveness of Aloe vera-a review. *Journal of Pre-Clinical and Clinical Research*. 2017; 11: 86–93.