

Opinion

One Size Does Not Fit All: Precision Combinations for FGFR4-driven Cancers

Emmy D.G. Fleuren^{1,2,3,*} 

¹Children's Cancer Institute, Lowy Cancer Research Centre, UNSW Sydney, Sydney, NSW 2031, Australia

²School of Clinical Medicine, UNSW Medicine & Health, UNSW Sydney, Sydney, NSW 2052, Australia

³UNSW Centre for Childhood Cancer Research, UNSW Sydney, Sydney, NSW 2031, Australia

*Correspondence: efleuren@ccia.org.au (Emmy D.G. Fleuren)

Academic Editor: Amancio Carnero Moya

Submitted: 18 June 2025 Revised: 12 August 2025 Accepted: 18 August 2025 Published: 24 September 2025

Abstract

Oncogenic FGFR4 signalling represents an attractive therapeutic target across multiple cancers, yet treatment resistance almost uniformly occurs. A critical mechanism steering resistance is a rapid and complex reprogramming of kinase signalling networks, called the adaptive bypass response. Capturing this dynamic rewiring to pinpoint, on a molecular level, the right combinatorial drug for the right FGFR4-driven cancer patient at the right time, will be key to achieving sustained tumour responses. But how can one accurately capture this process across different cancer types exhibiting contrasting levels of FGFR4 signalling pathway components and network behaviours? A recent study by Shin *et al.* delivers a technically elegant and biologically grounded exploration of the adaptive signalling landscape to tackle this, revealing cell context-dependent combinatorial strategies.

Keywords: cancer; FGFR4; molecular targeted therapy; signal transduction; precision medicine; drug resistance; drug combination

1. Introduction

Aberrant signalling of the Receptor Tyrosine Kinase (RTK) FGFR4 is an emerging therapeutic target across various cancers. Mechanisms driving pathway dysregulation are diverse, and include FGFR4 activating mutations, amplification and overexpression in breast cancer (BC), and ligand/receptor co-expression, such as FGF19/FGFR4 in hepatocellular carcinoma (HCC) and FGF8/FGFR4 in a subset of rhabdomyosarcoma (RMS) [1–4]. FGFR4-specific kinase inhibitors have shown encouraging efficacy in preclinical and early clinical settings, yet, as with any kinase inhibitor, intrinsic and acquired resistance remain a major barrier to durable outcomes. An important mechanism steering resistance is the dynamic adaptive bypass response, which involves rapid reprogramming of kinase signalling networks upon drug exposure that enable tumour cells to circumvent drug target inhibition. Understanding the mechanism, molecules and biology driving this initial resistance and identifying matched rational drug combinations to overcome it, will be key to achieving durable responses. Capturing this process is however complicated, not in the least due to the complexity of positive and negative feedback loops, and the vast molecular heterogeneity across the different FGFR4-driven cancers. Thus, for FGFR4-targeted therapies, how can one accurately capture and understand these complex resistance dynamics across different FGFR4-driven cancers, to ultimately determine which targeted drug combination is expected to work best for which patient? A recent study from Shin *et al.* [5] elegantly addresses this problem by integrating computational

network modelling with experimental validation, offering a framework to predict resistance dynamics and tailor combinatorial strategies accordingly. This Opinion Article aims to both discuss the scientific and methodological achievements of this study, and place their findings into the broader context of FGFR4 resistance mechanisms.

2. Computational Modelling of FGFR4 Network Behaviour

To elucidate mechanisms of rapid adaptive resistance to FGFR4-inhibitors across cancer types, Shin *et al.* [5] first developed a mechanistic mathematical model of the FGFR4 signalling network. At its core, the model centres on FGFR4 and its downstream signalling cascades via FRS2-, PI3K/AKT/mTOR- and RAS/RAF/MEK/ERK-pathways. To account for pathway convergence from other RTKs, it also incorporates IGF1R/IR and ErbB family receptors. Key negative feedback regulators, such as CBL, SPRY2, and protein tyrosine phosphatase (PTP) activity, are also integrated in the model (Fig. 1A, Ref. [5]). This model was iteratively refined using experimental data from MDA-MB-453 triple-negative breast cancer (TNBC) cells harbouring an activating FGFR4 mutation. Model calibration was guided by cell viability assays and Western blot data capturing key kinases' temporal responses to FGFR4 inhibition alone and in combination with predicted synergistic or non-synergistic agents. Upon successful prediction and experimental validation in this model, the authors expanded their framework using protein expression profiles from 350 cancer cell lines in the Cancer Cell Line Encyclopedia



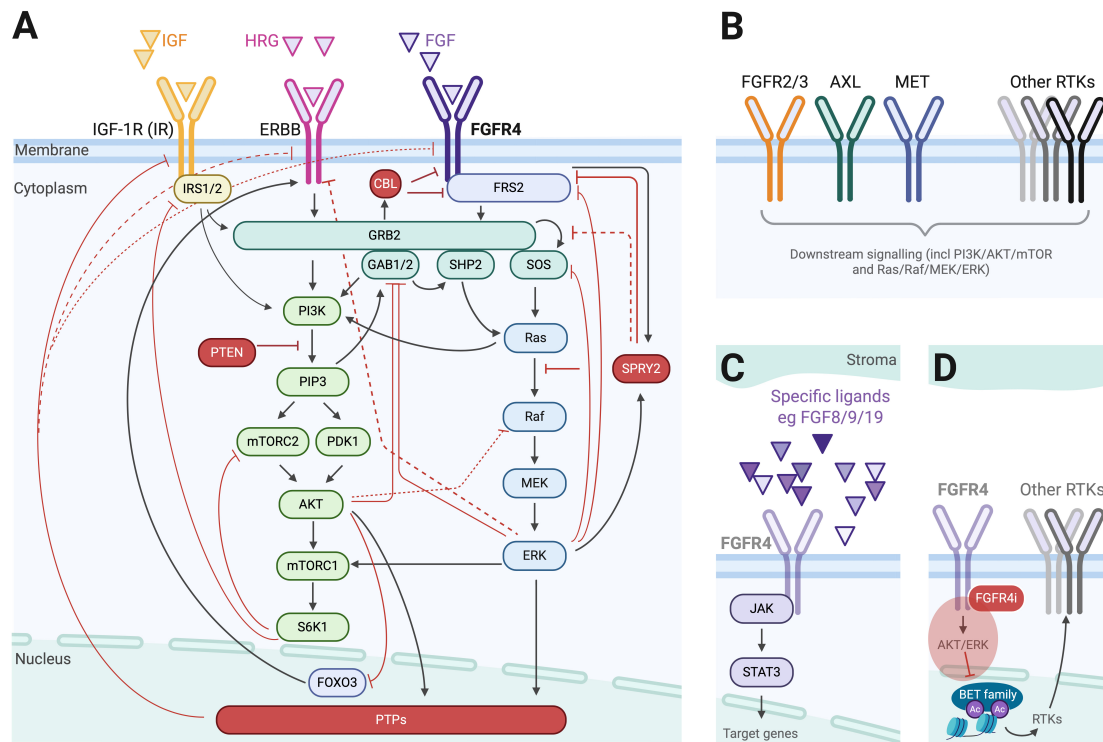


Fig. 1. The FGFR4 signalling network and molecular mediators of resistance to FGFR4 inhibition. (A) The FGFR4 signalling network utilized by Shin *et al.* [5] as the basis for their mathematical model. Red molecules and lines represent negative pathway modulators; black arrows represent pathway stimulation. PTPs, protein tyrosine phosphatases. (B) Illustration of alternate molecular mediators driving FGFR4-inhibitor resistance not captured in the published model, focussing on receptor tyrosine kinases (RTKs). (C) Illustration of alternate molecular mediators driving FGFR4-inhibitor resistance not captured in the published model, illustrating extracellular ligand- and intracellular STAT3-signalling. (D) Illustration of how inhibition of FGFR4-signalling with an FGFR4-inhibitor (FGFR4i, shown in red) is hypothesized to relieve transcriptional repression of other RTKs via nuclear BET/epigenetic-signalling. Created in BioRender. Fleuren, E. (2025) <https://BioRender.com/vwaokpc>.

(CCLE) [6]. This enabled simulation of context-specific adaptive responses to FGFR4 inhibition across diverse tumour types. Model outputs stratified cancer cell lines into four distinct categories based on AKT and/or ERK pathway reactivation (“rebound”) dynamics following FGFR4 inhibition. Functional validation confirmed AKT-mediated resistance in the FGFR4 mutationally-driven TNBC model, and ERK-mediated resistance in an FGF19-amplified HCC cell line, Hep3B. These findings support rational combination strategies, pairing FGFR4-inhibitors with AKT- or ERK-inhibitors, tailored to the network dynamics of each tumour.

This study represents a novel integration of mechanistic modelling, *in silico* drug combination screening, and experimental validation to comprehensively interrogate adaptive resistance mechanisms within the FGFR4 signalling network. Its iterative model-experiment cycle enables unbiased predictions of non-obvious drug combinations: a substantial advantage over more intuitive approaches. A key study implication is that FGFR4-targeted drug combinations that may work for one patient or one cancer type, may not work for another, based on the dynamic behaviour

of the associated FGFR4 network. What would be worthy of further exploration, is examine how many CCLE cell lines could be considered FGFR4-driven (e.g., by harbouring FGFR4 activating mutations or FGFR4 and/or ligand overexpression), and how these would map specifically onto the model’s rebound categories. For instance, when considering FGFR4 protein expression alone, only 12 lines exhibit a normalized score >1, including some BC, HCC and RMS cell lines [6]. Ligands also play a role in some of these, and potentially other, cell models. In this context, do all FGF19/FGFR4-driven HCC models display ERK rebound, or do individual variations exist? Similarly, what characterises FGF8/FGFR4-driven rhabdomyosarcoma rebound? Application of the novel model can undoubtedly answer these questions to further increase our understanding of precision-guided combinatorial drug strategies for FGFR4-driven cancers.

Another intriguing finding emerged from TNBC cells, where co-inhibition of FGFR4- with AKT-inhibitors proved more effective than co-treatment with PI3K-inhibitors, despite both targeting the same signalling axis. This divergence is particularly noteworthy given AKT-, PI3K-, or

mTOR-inhibitors are often used interchangeably, in clinical practice often guided by drug accessibility or anticipated toxicity rather than a mechanistic rationale, and often favouring mTOR-inhibitors in that respect [7]. By leveraging a systems-level approach, the authors further elucidated that the observed differential efficacy stemmed from the network's negative feedback architecture. Specifically, negative regulators such as CBL, SPRY2, and PTPs played a pivotal role in modulating AKT reactivation dynamics following FGFR4 inhibition. These insights underscore the importance of interrogating full network behaviours, involving both positive and negative regulators, rather than solely relying on static analysis of canonical pathway nodes.

3. Proteomics-informed FGFR4-based Drug Combinations

A notable strength of the study is its integration of large-scale proteomics data to inform combinatorial drug decision-making, for which biomarkers are notoriously difficult to pinpoint. While genomics has traditionally dominated biomarker discovery, genomics-based predictive biomarkers for FGFR4-inhibitor efficacy such as FGFR4 mutations or FGF-ligand expression do not fully capture the molecular context of tumours, nor account for dynamic changes in protein expression. Proteomics offers a more comprehensive view of the proteome, complementary to genomics, enabling the identification of additional or more nuanced markers of response and resistance. As precision medicine platforms increasingly incorporate proteomic data, including tumour microenvironmental components, the integration of such profiles into predictive models is expected to enhance the identification of optimal combinatorial therapies and improve patient outcomes [8].

4. Future Directions: Expanding and Refining the FGFR4 Signalling Model

Shin *et al.*'s framework [5] distinguishes itself from earlier models through its breadth and depth, incorporating both canonical effectors and less commonly modelled components such as tumour suppressor genes and phosphatases. But does it capture all potential molecular effect modulators? While the model is comprehensive and offers valuable insights, it is inherently shaped by its scope. If model complexity would allow, and appreciating the trade-offs with identifiability and predictive power, the model could benefit from some refinements and expansions, as detailed below.

4.1 Incorporating Additional RTKs and Pathways

At its core, the model centres on FGFR4 and its major downstream signalling cascades, supported by both literature and iterative experimental validation in a TNBC cell line (Fig. 1A). While the authors included two additional RTK families, namely IGF1R/IR and ErbB, this largely

reflects relevance to TNBC. Given the diversity of RTK expression across tumour types, incorporating additional RTKs where feasible might improve applicability across cancers, and reveal alternate resistance mediators to further guide combination strategies (Fig. 1B). FGFR3 can for example compensate for FGFR4 inhibition in HCC, and the RTKs MET and AXL confer resistance to various (receptor tyrosine) kinase inhibitors across different cancer types, including those sensitive to FGFR4 inhibition [9–14]. As the model, in addition to AKT-targeting, also predicted and validated ErbB-targeting to be of synergistic relevance in TNBC, a similar rebound might exist with other RTKs in other cancer types. Another pathway worthy of exploration is the JAK/STAT-pathway, as recent studies highlighted STAT3 activation in response to FGFR4 blockade (Fig. 1C) [15].

4.2 Modelling the Tumour Microenvironment

Additionally, in an ideal scenario, this and many other predictive models would benefit from capturing extracellular features, such as ligand-level dynamics and other tumour microenvironment (TME) features. The TME is however notoriously difficult to model and challenging to experimentally validate. Although ligands for IGF1R, FGFR4 and ErbB are for example broadly depicted in the model figures, it is unclear how differential protein expression of such ligands will affect the system. While Hep3B cells used in this study overexpress FGF19, and two rhabdomyosarcoma cell lines (Rh41 and Rh30) included in CCLE are known to overexpress FGF8, neither FGF19, nor FGF8, is listed in the CCLE protein dataset [6]. Furthermore, in HCC models, including Hep3B, stroma-derived FGF9 promotes tumorigenicity and resistance to the multi-kinase inhibitor sorafenib, and pro-tumorigenic effects of FGF9 could only be abrogated by an FGFR1/2/3-inhibitor, but not through FGFR4-inhibition. FGFR2 was suggested as a potential mediator in this response (Fig. 1B,C) [16]. Another study in HCC cells illustrates how exogenous EGF supplementation resulted in FGFR4-inhibitor resistance, which was effectively reverted by EGFR- (and SHP2)-inhibitors [17].

4.3 Accounting for Dynamic Treatment Adaptation

By examining kinase network remodelling within hours after drug exposure, Shin *et al.* [5] demonstrated that cancer context-dependent co-targets exist for FGFR4-mediated therapies. But will these novel combinations be the key to eliciting durable tumour responses in patients with FGFR4-driven cancers? As the authors acknowledge, it remains critical to evaluate the long-term efficacy and durability of such combinations in models that better recapitulate the clinical setting. Will the proposed combinatorial strategies elicit sustained tumour control, or will resistance eventually re-emerge via alternative pathways? Unless the combination effectively eradicates all cancer cells, surviving cell populations, even under com-

combination treatment pressure, may undergo further adaptive rewiring. Taking TNBC with AKT rebound and accordingly co-targeted with AKT- and FGFR4-inhibitors as an example, activation of alternative RTKs or bypassing the PI3K–AKT axis through reactivation of MAPK-signalling might occur in the longer term [18]. A particularly compelling feature of the novel framework is that it could, in principle, capture this next layer of network dynamics and work beyond predicting synergistic drug pairs. Re-deploying the model to map adaptive reprogramming under combination therapy might reveal additional targets for triplet or higher-order drug combinations. The prospect of transforming the model to a dynamic platform, with scope to deliver adaptable, evolution-informed sequential therapeutic strategies over time, significantly extends its future impact. One point of caution however is that not all high(er)-level combinations might be feasible. An interesting alternate approach suggested in the literature to bypass this, involves blocking the adaptive response itself via transcriptional repression at the epigenetic level. In HER2-amplified cell lines treated with lapatinib for example, targeting BET-family bromodomain mediators inhibited lapatinib-induced kinome reprogramming [19,20], and a similar effect was reported in melanoma, where targeting BRD/BET-proteins inhibited the adaptive kinome up-regulation and enhanced effects of BRAF/MEK-inhibitors [21]. While a similar approach might enhance efficacy of FGFR4-inhibitors (Fig. 1D), an important point of caution is that BRD/BET-inhibitors can, particularly when used in drug combinations, result in significant toxicity [22]. Thus, while rationally-selected kinase inhibitor combinations might still provoke adaptive signalling responses, selectively targeting critical nodes, even if this necessitates a triplet regimen, remains the favourable approach.

5. Clinical Translation

While tumour heterogeneity and the TME are hard to capture in current systems, these will be essential for predicting clinical efficacy. Future work incorporating longitudinal sampling, functional profiling of resistant clones, and more clinically-relevant models will be important in truly determining the therapeutic promise of these combinations. It will also be of interest to explore how the new modelling approach could be applied to new FGFR4-based therapy combinations, such as the combination of FGFR4-inhibitors with the immune checkpoint inhibitors pembrolizumab (NCT04699643) and spartalizumab (NCT02325739), since these approaches have shown promising efficacy in FGFR4-driven cancers, including HCC [23,24].

6. Concluding Remarks

Shin *et al.*'s study [5] exemplifies how coupling computational modelling with experimental validation can decode the complexity of FGFR4 signalling, reveal context-

specific resistance mechanisms and identify rational, personalised combination strategies to overcome FGFR4-inhibitor resistance. The ability to predict and validate synergistic drug combinations based on computational modelling of early signalling network dynamics, incorporating non-intuitive behaviour, represents a major advantage over earlier studies. While the model omits elements like the TME and broad RTK coverage, it lays a strong foundation for future explorations. As clinical proteomics data become increasingly available, this framework offers an exciting opportunity to tailor therapies across heterogeneous and adaptive tumour landscapes. It highlights the potential of signalling network modelling as a powerful addition to the precision oncology toolbox; one that anticipates tumour evolution and informs rational and adaptable therapeutic strategies accordingly.

Author Contributions

EDGF confirms responsibility for all aspects of this manuscript; from its conception, literature review, manuscript writing, results presentation, figure construction, and revising the final version. EDGF contributed to editorial changes in the manuscript. EDGF read and approved the final manuscript. EDGF participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

EDGF is supported through a Col Reynolds Mid-Career Discover Fellowship from The Kids' Cancer Project.

Conflict of Interest

The author declares no conflict of interest.

References

- [1] Brown LM, Ekert PG, Fleuren EDG. Biological and clinical implications of FGFR aberrations in paediatric and young adult cancers. *Oncogene*. 2023; 42: 1875–1888. <https://doi.org/10.1038/s41388-023-02705-7>.
- [2] Chew NJ, Lim Kam Sian TCC, Nguyen EV, Shin SY, Yang J, Hui MN, *et al.* Evaluation of FGFR targeting in breast cancer through interrogation of patient-derived models. *Breast Cancer Research: BCR*. 2021; 23: 82. <https://doi.org/10.1186/s13058-021-01461-4>.
- [3] Wu X, Ge H, Lemon B, Vonderfecht S, Weiszmann J, Hecht R, *et al.* FGF19-induced hepatocyte proliferation is mediated through FGFR4 activation. *The Journal of Biological Chemistry*. 2010; 285: 5165–5170. <https://doi.org/10.1074/jbc.M109.068783>.
- [4] Fordham AM, Brown LM, Mayoh C, Salib A, Barger ZA, Wong

- M, *et al.* Comprehensive multi-platform tyrosine kinase profiling reveals novel actionable FGFR aberrations across pediatric and AYA sarcomas. *bioRxiv*. 2023; 2023.07.19.548825. <https://doi.org/10.1101/2023.07.19.548825>. (preprint)
- [5] Shin SY, Chew NJ, Ghomlaghi M, Chüeh AC, Jeong Y, Nguyen LK, *et al.* Integrative Modeling of Signaling Network Dynamics Identifies Cell Type-Selective Therapeutic Strategies for FGFR4-Driven Cancers. *Cancer Research*. 2024; 84: 3296–3309. <https://doi.org/10.1158/0008-5472.CAN-23-3409>.
 - [6] Nusinow DP, Szpyt J, Ghandi M, Rose CM, McDonald ER, 3rd, Kalocsay M, *et al.* Quantitative Proteomics of the Cancer Cell Line Encyclopedia. *Cell*. 2020; 180: 387–402.e16. <https://doi.org/10.1016/j.cell.2019.12.023>.
 - [7] Dienstmann R, Rodon J, Serra V, Tabernero J. Picking the point of inhibition: a comparative review of PI3K/AKT/mTOR pathway inhibitors. *Molecular Cancer Therapeutics*. 2014; 13: 1021–1031. <https://doi.org/10.1158/1535-7163.MCT-13-0639>.
 - [8] Xie Y, Chen X, Xu M, Zheng X. Application of the Human Proteome in Disease, Diagnosis, and Translation into Precision Medicine: Current Status and Future Prospects. *Biomedicines*. 2025; 13: 681. <https://doi.org/10.3390/biomedicines13030681>.
 - [9] Fernandes M, Jamme P, Cortot AB, Kherrouche Z, Tulasne D. When the MET receptor kicks in to resist targeted therapies. *Oncogene*. 2021; 40: 4061–4078. <https://doi.org/10.1038/s41388-021-01835-0>.
 - [10] Hsu CH, Huang YH, Lin SM, Hsu C. AXL and MET in Hepatocellular Carcinoma: A Systematic Literature Review. *Liver Cancer*. 2022; 11: 94–112. <https://doi.org/10.1159/000520501>.
 - [11] Scaltriti M, Elkabets M, Baselga J. Molecular Pathways: AXL, a Membrane Receptor Mediator of Resistance to Therapy. *Clinical Cancer Research: an Official Journal of the American Association for Cancer Research*. 2016; 22: 1313–1317. <https://doi.org/10.1158/1078-0432.CCR-15-1458>.
 - [12] Tao Z, Cui Y, Xu X, Han T. FGFR redundancy limits the efficacy of FGFR4-selective inhibitors in hepatocellular carcinoma. *Proceedings of the National Academy of Sciences of the United States of America*. 2022; 119: e2208844119. <https://doi.org/10.1073/pnas.2208844119>.
 - [13] van Erp AEM, Versleijen-Jonkers YMH, van der Graaf WTA, Fleuren EDG. Targeted Therapy-based Combination Treatment in Rhabdomyosarcoma. *Molecular Cancer Therapeutics*. 2018; 17: 1365–1380. <https://doi.org/10.1158/1535-7163.MCT-17-1131>.
 - [14] Wood GE, Hockings H, Hilton DM, Kermorgant S. The role of MET in chemotherapy resistance. *Oncogene*. 2021; 40: 1927–1941. <https://doi.org/10.1038/s41388-020-01577-5>.
 - [15] Narisawa T, Naito S, Ito H, Ichiyanagi O, Sakurai T, Kato T, *et al.* Fibroblast growth factor receptor type 4 as a potential therapeutic target in clear cell renal cell carcinoma. *BMC Cancer*. 2023; 23: 170. <https://doi.org/10.1186/s12885-023-10638-3>.
 - [16] Seitz T, Freese K, Dietrich P, Thasler WE, Bosserhoff A, Hellerbrand C. Fibroblast Growth Factor 9 is expressed by activated hepatic stellate cells and promotes progression of hepatocellular carcinoma. *Scientific Reports*. 2020; 10: 4546. <https://doi.org/10.1038/s41598-020-61510-4>.
 - [17] Shen B, Shi JP, Zhu ZX, He ZD, Liu SY, Shi W, *et al.* EGFR Inhibition Overcomes Resistance to FGFR4 Inhibition and Potentiates FGFR4 Inhibitor Therapy in Hepatocellular Carcinoma. *Molecular Cancer Therapeutics*. 2023; 22: 1479–1492. <https://doi.org/10.1158/1535-7163.MCT-23-0096>.
 - [18] Szymczyk J, Sluzalska KD, Materla I, Opalinski L, Otlewski J, Zakrzewska M. FGF/FGFR-Dependent Molecular Mechanisms Underlying Anti-Cancer Drug Resistance. *Cancers*. 2021; 13: 5796. <https://doi.org/10.3390/cancers13225796>.
 - [19] Stuhlmiller TJ, Miller SM, Zawistowski JS, Nakamura K, Beltran AS, Duncan JS, *et al.* Inhibition of Lapatinib-Induced Kinome Reprogramming in ERBB2-Positive Breast Cancer by Targeting BET Family Bromodomains. *Cell Reports*. 2015; 11: 390–404. <https://doi.org/10.1016/j.celrep.2015.03.037>.
 - [20] Stuhlmiller TJ, Miller SM, Johnson GL. Epigenetic inhibition of adaptive bypass responses to lapatinib by targeting BET Bromodomains. *Molecular & Cellular Oncology*. 2015; 3: e1052182. <https://doi.org/10.1080/23723556.2015.1052182>.
 - [21] Tiago M, Capparelli C, Erkes DA, Purwin TJ, Heilman SA, Berger AC, *et al.* Targeting BRD/BET proteins inhibits adaptive kinome upregulation and enhances the effects of BRAF/MEK inhibitors in melanoma. *British Journal of Cancer*. 2020; 122: 789–800. <https://doi.org/10.1038/s41416-019-0724-y>.
 - [22] Doroshow DB, Eder JP, LoRusso PM. BET inhibitors: a novel epigenetic approach. *Annals of Oncology: Official Journal of the European Society for Medical Oncology*. 2017; 28: 1776–1787. <https://doi.org/10.1093/annonc/mdx157>.
 - [23] Chan SL, Schuler M, Kang YK, Yen CJ, Edeline J, Choo SP, *et al.* A first-in-human phase 1/2 study of FGF401 and combination of FGF401 with spartalizumab in patients with hepatocellular carcinoma or biomarker-selected solid tumors. *Journal of Experimental & Clinical Cancer Research: CR*. 2022; 41: 189. <https://doi.org/10.1186/s13046-022-02383-5>.
 - [24] Xu J, Cui J, Jiang H, Zeng Y, Cong X. Phase 1 dose escalation study of FGFR4 inhibitor in combination with pembrolizumab in advanced solid tumors patients. *Cancer Medicine*. 2023; 12: 7762–7771. <https://doi.org/10.1002/cam4.5532>.