

Research Article

Effect of Neocuproine, a Selective Cu(I) Chelator in Isolated Whole-Bladder Preparations From Neonatal and Adult Rats

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Abstract

Background and Objective: The administration of neocuproine (NC), a copper(I)-binding agent, can induce phasic contractions in the neonatal rat bladder, while NC treatment can also promote tonic contractions in the adult rat bladder. These data may indicate that copper is important in regulating developmental contractile responses. Thus, investigating how responses to copper(I)-binding agents change during development could offer valuable insights into the mechanisms underlying overactive bladder. Therefore, this study aimed to induce developmentally different contraction responses in isolated rat whole-bladder tissues using NC. Materials and Methods: Neonatal (1-2 weeks old) and adult (250-300 g) Wistar rats were used in the experiments. Isolated rat bladders were placed in organ baths containing 2 µM atropine and 2 µM guanethidine Krebs solution. The resulting tone changes in the preparation were recorded using a pressure transducer (cmH₂O). Differences between groups were evaluated using one-way analysis of variance (ANOVA). The Student's t-test was used to assess the paired groups (GraphPad Prism); p-values smaller than 0.05 were considered statistically significant. Results: NC administration caused a significant increase in basal spontaneous contraction responses in neonatal rat whole-bladder tissue, whereas in adult whole bladder, NC treatment promoted a significant tonic contraction over basal tonus. In isolated whole-bladder tissue from neonatal rats, the increases in the amplitude and area under the curve (AUC) of spontaneous contraction responses induced by 50 µM NC were significantly reduced by the addition of 1 µM nifedipine, 50 µM adenosine triphosphate (ATP), 100-200 µM suramin, or 30 μM NS1619, a large-conductance Ca²⁺-activated K⁺ [BK] channel opener, to the medium. The number of spontaneous contractions decreased in the presence of NC; however, this finding was reversed in the presence of the aforementioned drugs. Moreover, the tonic contractions observed in the presence of NC in the adult bladder preparations were significantly reduced following the addition of 1 µM nifedipine, 50 μM ATP, 100 μM suramin, or 30 μM NS1619. However, the addition of 100 μM N^w-nitro-L-arginine (LA) to the medium had no significant effect on the amplitude, AUC, and frequency of NC-induced tonic contractions or spontaneous contraction responses. Conclusion: These results suggest that copper may play an important role in the regulation of phasic/tonic contractions in the bladder during postnatal development.

Keywords: copper; urinary bladder; potassium channels; ATP; muscle contraction

1. Introduction

The bladder can produce spontaneous contractions, either small or large. It has been observed in experimental studies that increased spontaneous contractions are similar to contractions that occur without urination due to afferent pathway activation *in vivo* in the overactive bladder. The literature suggests that the phasic contractions occurring in isolated bladder tissue could be linked to activities during the filling phase of the micturition cycle, without leading to urination; these contractions are referred to as non-voiding contractions [1–4]. While the functionally normal bladder contracts only during urination, phasic contractions occur in the detrusor even during the filling phase in the case of an overactive bladder.

The neural control and contractility of the bladder undergo significant changes during postnatal development. In bladder preparations from neonatal rats, it has been observed that the amplitude of spontaneous contractions is low during the first week of postnatal life, but the amplitude increases progressively during the second week [5-7]. Rat bladders in the early postnatal period exhibit symptoms similar to those seen in human overactive bladders. Therefore, the neonatal rat bladder can be used as an experimental model for investigating and solving overactive bladder problems [8]. Some articles suggest that neonatal and adult bladders have different characteristics [7,9–11]. In adult bladders, contractions tend to be of low amplitude and high frequency, whereas neonatal bladders exhibit phasic contractions with higher amplitude and lower frequency. Contraction parameters in the newborn change to adult parameters approximately 4 to 5 weeks after birth. However, neonatal parameters may recur in adults with pathological conditions such as chronic urethral obstruction [9] or spinal cord injury [10]. In a healthy human bladder, atropine

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significantly inhibits parasympathetic nerve-mediated contractions. This inhibition indicates that cholinergic mechanisms are more dominant in parasympathetic-mediated contractions occurring in the healthy bladder. However, there are also publications suggesting that the response is not blocked by atropine (atropine-resistant) and is related to purinergic signaling [12,13]. In an experimental study conducted on adult and neonatal rat bladders, it was observed that purinergic activity was higher in neonates compared to adults [14].

Copper is a trace element that plays an essential role in various cellular functions [15]. Studies have shown that the targets of copper include neuronal postsynaptic neurotransmitter receptors [16–18] and voltage-gated ion channels [19]. In a study conducted by Morera and colleagues [15] on skeletal muscle, it was shown that external Cu^{2+} causes inhibition of BK_{Ca} channel activity. Therefore, in our study, the role of Ca^{2+} -activated K^+ channels, specifically large-conductance Ca^{2+} -activated K^+ channels (BK_{Ca}), in the mechanism of spontaneous contractions observed in the presence of neocuproine has been demonstrated.

Research on copper chelators and rat bladder function has indicated that copper-dependent pathways might influence or contribute to adenosine triphosphate (ATP) or nitric oxide (NO)-driven responses [20]. Moreover, copper ions have been reported to alter the activity of P2X4 purinergic receptors expressed in *Xenopus* oocytes [21,22]. This observation suggests that other P2X receptor subtypes present in the bladder could likewise be susceptible to modulation by copper ions.

A literature review revealed that the effects of copper on neonatal rat bladder tone and its mechanism of action have not been previously investigated. Therefore, this study compared the spontaneous tone changes induced by neocuproine, a copper-binding agent, in the bladders of adult and neonatal rats. While neocuproine (NC) induced spontaneous phasic contractions in the neonatal bladder, it caused an increase in tone in the adult bladder. The role of purinergic pathways, membrane Ca²⁺ channels, and K⁺ channels in this response difference was investigated.

2. Materials and Methods

2.1 Study Area

Preliminary studies were conducted in the Department of Medical Pharmacology at the Faculty of Medicine in 2008. Experiments were initiated in the Department of Pharmacology at the Faculty of Pharmacy between 2022 and 2023, and previously conducted experiments were repeated. Data from these preliminary studies were not used.

2.2 Animals

A total of 56 neonatal (1–2 weeks old) and 42 adult (250–300 g) Wistar rats (Çukurova University Health Sciences Experimental Application and research center-SABIDAM-Adana/Turkiye), both female and male, were

used in the experiments. Animals were maintained under standard laboratory conditions with a 12-hour light/dark cycle. The experimental protocols were approved by the ethics committee of Cukurova University Animal Experiments Local Ethics Committee (03-11-2004/136) and conducted in accordance with the *Principles of Laboratory Animal Care* as outlined by the National Institutes of Health (NIH publication 86–23, revised 1984).

2.3 Drugs

Neocuproine, guanethidine, atropine, diamide, $N^{\rm w}$ -nitro-L-arginine, suramin, and ATP were prepared in distilled water. Nifedipine and NS1619 were dissolved in 75% ethyl alcohol, ensuring that the final concentration in the bath medium did not exceed 0.08%. All compounds were supplied by Sigma-Aldrich (St. Louis, MO, USA).

2.4 In Vitro Whole-bladder Preparation

A total of 98 rats were anesthetized with 4-5% (v/v) halothane and decapitated. Following a midline laparotomy, the urinary bladder, together with the urethra, was excised as a single unit. Whole-bladder preparations were obtained using methods adapted from earlier studies [9,11] A 26-gauge (26G) needle was then inserted into the bladder through the urethra and secured with 5-0 silk sutures. The other end of the needle was connected to a pressure transducer and an infusion pump (Harvard Apparatus) via a polyethylene cannula. Each preparation was placed in a temperature-controlled organ bath filled with Krebs solution (composition in mM: NaCl 113, NaHCO₃ 19.8, glucose 11.1, KH₂PO₄ 1.2, KCl 4.7, MgCl₂ 2.5, CaCl₂ 1.7), maintained at 37 °C and continuously gassed with 95% O₂ and 5% CO₂. The bath fluid also contained atropine (2 μM) to block muscarinic cholinergic receptors and guanethidine (2 μM) to inhibit adrenergic neurotransmission, allowing assessment of non-adrenergic, non-cholinergic (NANC) activity. Preparations were allowed to equilibrate for 30 minutes prior to the initiation of the filling protocol. After this period, the tissues were slowly infused with Krebs solution using an infusion pump at a rate of 50 µL per minute. At the end of each infusion, electrical field stimulation (Grass S88; EFS, 50 volts, 1.5 ms duration, 32 Hz, 15-30 seconds) was applied. The bladder was infused until the contraction induced by EFS reached its maximum. Once the bladder reached a certain level of fullness, the infusion was stopped, and the bladder was washed with Krebs solution. All experiments were carried out with the bladder fully filled. Following a 30-minute equilibration period, drug treatments were applied while maintaining the bladder in this fully filled state. All drugs were administered 20 minutes before neocuproine (NC, 50 µM). Bladder pressure was monitored in cmH2O via a pressure transducer connected to a Data Acquisition System (BIOPAC MP 30). Spontaneous contraction amplitude and frequency were determined during a 5-minute analysis window within



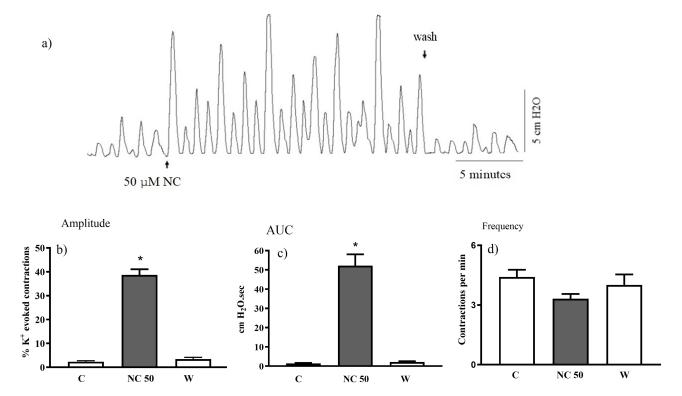


Fig. 1. Effects of 50 μ M NC (NC 50) in neonatal rat bladders. (a) Representative tracing showing the effects of NC, (b) Amplitude, (c) AUC in cmH₂O-seconds, and (d) Frequency in contractions per min. C, Control; W, Washout (n = 8). *: p < 0.05, significant difference from control. NC, neocuproine; AUC, area under the curve.

a 10-minute observation period. Contraction amplitude was expressed relative to the maximal response elicited by 80 mM KCl at the end of the experiment. Frequency was calculated as the number of contractions per 5 minutes. Nifedipine (1 μ M; L-type Ca²+ channel blocker), N ω -nitro-L-arginine (100 μ M; nitric oxide synthase inhibitor), ATP (50 μ M; purinergic receptor agonist), suramin (100–200 μ M; purinergic receptor antagonist), and NS1619 (1,3-dihydro-1-[2-hydroxy-5-(trifluoromethyl) phenyl]-5-(trifluoromethyl)-2H-benzimidazol-2-one) (30 μ M; large-conductance Ca²+-activated K+ channel opener) [23] were studied in the presence of NC.

2.5 Statistical Analysis

In experiments conducted on neonatal and adult rat bladder tissues, the amplitude of the spontaneous or tonic contractions induced by the agents used was measured in cmH₂O. The spontaneous and tonic contractions obtained were expressed as a percentage relative to the contraction induced by 80 mM KCl applied at the end of each experiment. Additionally, the AUC for the spontaneous or tonic contractions in neonatal and adult rat bladders was expressed as cmH₂O·seconds, and the frequency of contractions occurring over 1 minute was expressed as the number of contractions per minute (contractions/min).

In experiments involving more than two groups, differences between groups were evaluated using one-way analysis of variance (one-way ANOVA). These comparisons were performed using the Tukey test (GraphPad Prism; version 5.0; GraphPad Software, San Diego, CA, USA). For the statistical evaluation of paired groups, the Student's t-test was used. The statistical significance of the data was assessed at the level of p < 0.05. All data are expressed as mean \pm standard error.

3. Results

3.1 Neocuproine-Induced Contractions in the Neonatal Rat Bladder

Following basal tone, the administration of 50 μ M NC into the organ bath caused non-reproducible spontaneous contractions in all isolated whole-bladder tissues (Fig. 1a). The amplitudes and AUCs of these spontaneous contractions showed a significant increase compared to the basal spontaneous contractions (Fig. 1b,c, p < 0.05). However, the number of contractions induced by NC resulted in a non-significant decrease compared to the number of basal spontaneous contractions (Fig. 1d). The changes in amplitude, AUC, and frequency of spontaneous contractions induced by NC were reversed by washing the environment with Krebs solution.





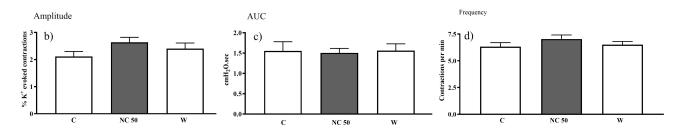


Fig. 2. Effects of 50 μ M NC (NC 50) in adult rat bladders. (a) Representative tracing showing the differential effects of NC in adult bladder, (b) amplitude, (c) AUC in cmH₂O·second, and (d) frequency in contractions per min. C, Control; W, Washout (n = 8).

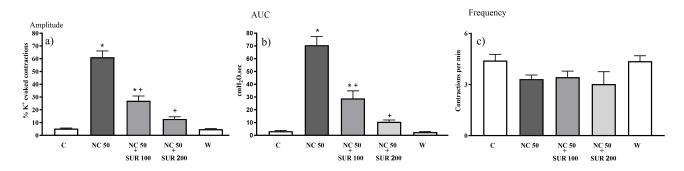


Fig. 3. The inhibitory effect of suramin 100 μ M (SUR100), or 200 μ M (SUR200) on the responses to 50 μ M NC (NC 50) in neonatal rat bladder. (a) Amplitude, (b) AUC in cmH₂O·second, and (c) Frequency in contractions per min. C, Control; W, Washout (n = 8). *: p < 0.05, significant difference from control, +: p < 0.05, significant difference from NC 50.

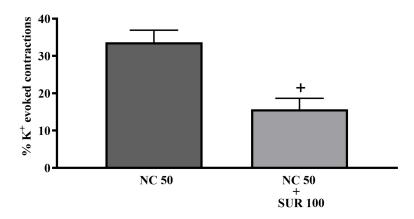


Fig. 4. The effects of suramin 100 μ M (SUR100) on tonic contractions of isolated whole-bladder preparations induced by 50 μ M NC (NC 50) in adult rats (n = 8). +: p < 0.05, significant difference from NC 50.

3.2 Neocuproine-Induced Contractions in the Adult Rat Bladder

In adult whole-bladder tissues, NC application did not cause phasic contractions, unlike in the neonatal blad-

der, but did increase basal tone (Fig. 2a). No significant changes were observed in the amplitude, AUC, and frequency of spontaneous contractions as a result of NC application (Fig. 2b–d).



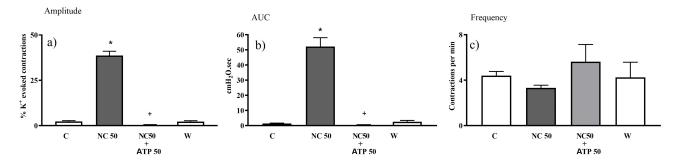


Fig. 5. The inhibitory effect of 50 μ M ATP (ATP 50) on the responses to 50 μ M NC (NC 50) in neonatal rat bladders. (a) Amplitude, (b) AUC in cmH₂O·second, and (c) Frequency in contractions per min. C, Control; W, Washout (n = 8). *: p < 0.05, significant difference from control, +: p < 0.05, significant difference from NC 50. ATP, adenosine triphosphate.

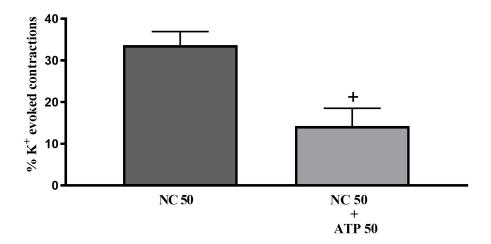


Fig. 6. The effects of 50 μ M ATP (ATP 50) on tonic contractions of isolated whole-bladder preparations induced by 50 μ M NC (NC 50) in adult rats (n = 8). +: p < 0.05, significant difference from NC 50.

3.3 Effect of Suramin on the Neocuproine Response in Neonatal and Adult Bladders

Suramin 100 μ M (SUR100) or 200 μ M (SUR200) significantly reduced the amplitude and AUC of spontaneous contractions induced by NC in neonatal rat bladders in a dose-dependent manner, while no significant change was observed in the frequency (Fig. 3a–c, p < 0.05).

The increase in tone caused by NC in adult bladder tissue was significantly reduced in the presence of 100 μM suramin (Fig. 4, p < 0.05).

3.4 Effects of ATP on the Neocuproine Response in Neonatal and Adult Bladders

In neonatal bladder tissue, ATP (50 μ M) significantly inhibited the amplitude and AUC of spontaneous contractions induced by NC (Fig. 5a,b, p < 0.05). Additionally, ATP reversed the non-significant decrease in the number of contractions caused by NC (Fig. 5c). The increase in tone induced by NC was significantly reduced in the presence of 50 μ M ATP in adult bladder tissue (Fig. 6, p < 0.05).

3.5 Effects of NS1619 on the Neocuproine Response in Neonatal and Adult Bladders

In neonatal rat bladder tissues, NS1619 (30 μ M) significantly inhibited the amplitude and AUC of spontaneous contractions induced by NC (Fig. 7a,b, p < 0.05), while partially reversing the non-significant decrease in the number of contractions (Fig. 7c).

In adult bladder tissue, the increase in tone induced by NC was significantly reduced in the presence of 30 μ M NS1619 (Fig. 8, p < 0.05).

3.6 Effects of Nifedipine on the Neocuproine Response in Neonatal and Adult Bladders

In neonatal rat bladder tissue, 1 μ M nifedipine (NIF 1) significantly inhibited the amplitude and AUC of spontaneous contractions induced by NC (Fig. 9a,b, p < 0.05). Additionally, nifedipine reversed the non-significant decrease in the number of spontaneous contractions induced by NC (Fig. 9c).

In adult rat bladder tissue, the tonic contraction response induced by NC was significantly reduced in the presence of 1 μ M nifedipine (NIF 1; Fig. 10, p < 0.05).



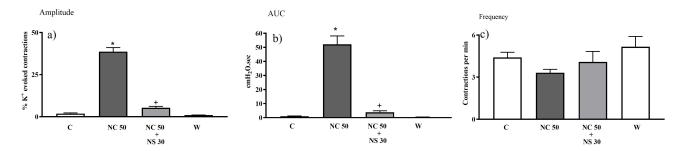


Fig. 7. The inhibitory effect of 30 μ M NS1619 (NS 30) on the responses to 50 μ M NC (NC 50) in neonatal rat bladders. (a) Amplitude, (b) AUC in cmH₂O-second, and (c) Frequency in contractions per min (n = 8), *: p < 0.05, significant difference from control, +: p < 0.05, significant difference from NC 50.

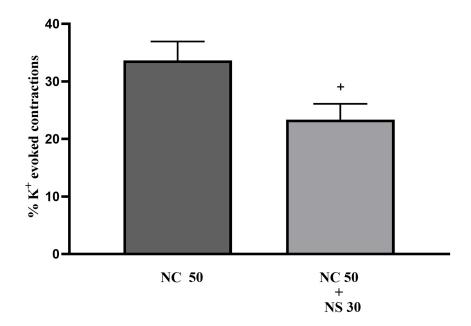


Fig. 8. The effects of 30 μ M NS1619 (NS 30) on tonic contractions of isolated whole-bladder preparation induced by 50 μ M NC in adult rats (n = 6). +: p < 0.05, significant difference from NC 50.

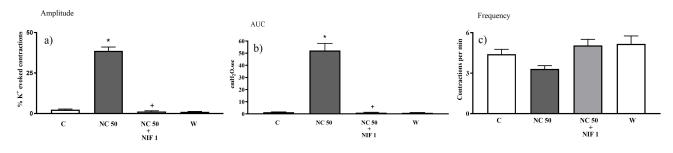


Fig. 9. The inhibitory effect of 1 μ M Nifedipine (NIF 1) on the responses to 50 μ M NC (NC 50) in neonatal rat bladder. (a) Amplitude, (b) AUC as cmH₂O·second, (c) Frequency as contractions per min. C, Control; W, Washout (n = 8). *: p < 0.05, significant difference from control, +: p < 0.05, significant difference from NC 50.

3.7 Effects of N^w-nitro-L-arginine on the Neocuproine Response in Neonatal and Adult Bladders

In neonatal rat bladder tissue, 100 μ M L-NA (L-NA 100) did not cause any significant changes in the amplitude, AUC, or frequency of spontaneous contractions induced by NC, as shown in Fig. 11a–c.

In adult bladder tissue, $100~\mu M$ L-NA also did not have a significant effect on the increase in tone induced by NC (Fig. 12).



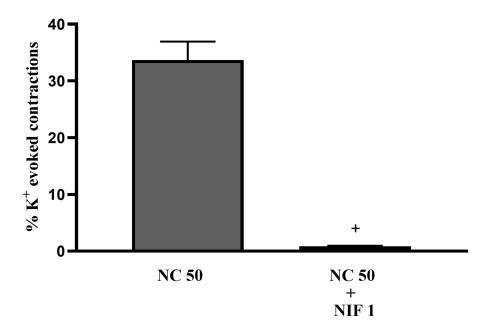


Fig. 10. The effects of 1 μ M nifedipine (NIF 1) on tonic contractions of isolated whole-bladder preparations induced by 50 μ M NC (NC 50) in adult rats (n = 6). +: p < 0.05, significant difference from NC 50.

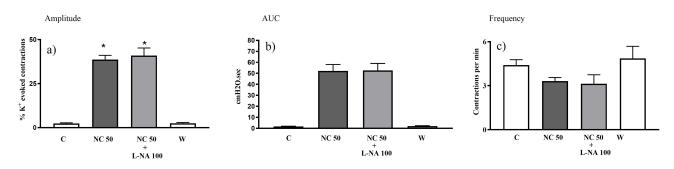


Fig. 11. The effect of 100 μ M L-NA (L-NA 100) on the responses to 50 μ M NC (NC 50) in neonatal rat bladder. (a) Amplitude, (b) AUC in cmH₂O-second, and (c) Frequency in contractions per min. C, Control; W, Washout (n=8). *: p<0.05, significant difference from control.

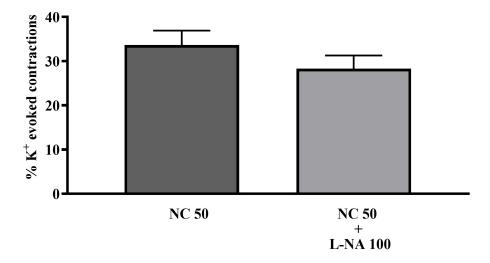


Fig. 12. The effects of 100 μ M L-NA (L-NA 100) on tonic contractions of isolated whole-bladder preparations induced by 50 μ M NC (NC 50) in adult rats (n=6).



4. Discussion

Significant changes occur in the neural control and contractile properties of the bladder during the postnatal period. Studies have shown that neonatal rat bladder preparations exhibit low-amplitude spontaneous contractions during the first week of life, but from the second week onward, these contractions gradually increase in amplitude and frequency, acquiring distinct phasic characteristics [5–7]. In contrast, low-amplitude but high-frequency contractions predominate in adult rat bladders. Therefore, in the early postnatal period, rat bladders exhibit symptoms similar to those seen in human overactive bladders, and the neonatal rat bladder can be considered a suitable experimental model for studying the pathophysiology of overactive bladder [8].

Based on these effects, this study demonstrated that neocuproine, a copper(I)-binding agent, has significantly different effects on the whole-bladder preparations isolated from neonatal and adult rats. While NC induced large amplitude phasic contractions in neonatal bladders, it only caused an increase in tone in adult bladders. Both phasic and tonic responses to NC were attenuated by an L-type Ca^{2+} channel blocker and a K^+ channel activator, aligning with previous evidence that copper can modulate smooth muscle contractility via Ca^{2+} and K^+ channel regulation [15,24].

Calcium channels are essential for normal bladder function, and the detrusor muscles are the smooth muscles in the bladder responsible for bladder contraction and relaxation [25]. Preclinical and clinical studies have demonstrated that calcium influx via L-type calcium channels plays a crucial role in bladder contractions [26,27]. In this study, the complete inhibition of copper-mediated NC effects in neonatal bladders by nifedipine, an L-type Ca²⁺ channel blocker, demonstrates that L-type channels are essential for these excitatory responses.

In the presence of atropine (cholinergic muscarinic receptor antagonist) and guanethidine (adrenergic neuron blocker), NC responses were unchanged in both neonatal and adult bladders, which suggests that this effect is not related to cholinergic or adrenergic mechanisms. However, ATP and suramin, a purinergic receptor antagonist, significantly inhibited NC-induced phasic/tonic contractions in the whole bladders. These findings suggest that NC enhances spontaneous/tonic activity by facilitating ATP release from nerves and/or activating purinergic pathways. Previous studies have shown that NC can modulate postjunctional purinergic signaling in bladder smooth muscle and urothelial cells [20,28,29] and can potentiate purinergic responses in other smooth muscle tissues, such as the rat vas deferens [30]. Research has shown that neuronal-mediated contractions of the detrusor smooth muscle persist in the presence of atropine and guanethidine, and the initial component of these contractions is suppressed by suramin, a P2X purinergic receptor antagonist

[12,31–33]. Husted and Nedergaard [34] also reported that ATP can desensitize contractile responses—a finding that may explain the attenuation of NC-induced contractions following ATP application in the present study. Moreover, ATP released from the urothelium is known to regulate afferent pathways via P2X3 receptors, influencing bladder reflex control [29,35,36]. Therefore, this study hypothesizes that NC may cause bladder overactivity by increasing the release of ATP from the urothelium or the effects of ATP on afferent nerves. The observed reduction in phasic contractions in neonatal rat bladders with suramin also supports this hypothesis. Considering the inhibitory effects of ATP and suramin on the phasic and tonic contractions induced by NC in neonatal and adult bladders, it can be suggested that the purinergic pathway plays a role in this mechanism. Additionally, our study indicates that a copper(I)sensitive mechanism plays a role in the facilitatory effects of neocuproine on bladder activity. Previous studies have suggested that copper can modulate the function of P2X4, another subtype of purinergic receptors, in Xenopus oocytes [21,22]. These results indicated that the purinergic efferent pathway in rat bladder tissue plays an important role in a copper (I)-sensitive mechanism.

This study also examined the role of the nitrergic system. Although nitric oxide (NO) released from the urothelium can influence afferent excitability [37,38], and phasic contractions in neonatal bladders have been reported to be sensitive to NO [39], another study on the adult bladder showed that the bladder is relatively insensitive to nitric oxide [40]. The findings of the present study indicate that NC-induced facilitation persists in the presence of the NO synthase inhibitor L-NA in both age groups. This is consistent with studies on the vas deferens [30] and the mouse bladder [41] showing that NC effects are independent of NO.

Finally, studies have shown that nonselective K⁺ channel blockers and specific Ca²⁺-activated K⁺ channel blockers modulate spontaneous contractions in neonatal rat bladders [11,42]. NS1619 is a potent BK channel activator with the potential to modulate cellular excitability in neurons and smooth muscle [43,44]. BK_{Ca} channels have been shown to be important in the down-regulation of spontaneous bladder contractions postnatally during maturation [13]. Studies have demonstrated that spontaneous contractions in both normal adult and diabetic bladders can be increased by the BK_{Ca} channel blocker iberiotoxin [45–47]. A study using human detrusor smooth muscle showed that the BK channel is an important physiological regulator of smooth muscle excitability and contractility [48]. Research by Soder and Petkov [49] proposed BK channel activation with NS1619 as a therapeutic approach to control myogenic and neurogenic contractions of detrusor smooth muscle (DSM). Therefore, activation of BK channels may be an important step in controlling bladder function in the pathophysiology of detrusor overactivity.



Our results suggest that NC-induced phasic contractions in neonatal bladders depend on both purinergic signaling and BK channel activity, in a manner reminiscent of pathological excitability in adult bladders. Future work should explore the molecular interplay between coppersensitive purinergic mechanisms and K⁺ channel regulation.

5. Conclusion

This study showed that neocuproine (NC), a copper-binding compound, produces different effects in isolated neonatal and adult bladder tissues. In neonatal bladders, NC-induced phasic contractions are mediated by Ca²⁺-activated K⁺ channels and/or purinergic pathways, with no significant involvement of the NO system. The bladder activity observed in neonates parallels changes seen in chronic bladder outlet obstruction and spinal cord injury models, and the early postnatal intrinsic bladder activity reappears in adult pathological states. These findings highlight the potential of copper(I)-binding agents as targets for developing new treatments for overactive and neurogenic bladder conditions.

6. Significance Statement

This study investigated the possible effect of copper(I) in isolated neonatal and adult rat bladder tissue. NC, a copper(I)-binding agent, produced developmentally different responses in neonatal and adult bladders. While NC produced large amplitude low-frequency rhythmic contractions in the neonatal rat bladder, it increased basal tone in the adult rat bladder. Copper may play an important role in the regulation of phasic/tonic contractions in the bladder during postnatal development. Therefore, understanding the effect of copper-binding agents on the neonatal bladder should provide us with information about new treatment approaches in bladder pathologies.

Availability of Data and Materials

The datasets used and analyzed during the current study are available from the corresponding author upon reasonable request.

Author Contributions

HSB designed and performed the experiments, analyzed the results, and drafted the manuscript. CG designed and performed the experiments. FC analyzed the results. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

The study was approved by the Ethics Committee of Cukurova University Animal Experiments Local Ethics

Committee (Decision No. 03-11-2004/136) and was conducted in accordance with the Principles of Laboratory Animal Care as outlined by the National Institutes of Health (NIH publication No. 86–23, revised 1984).

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Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Drake MJ, Harvey IJ, Gillespie JI. Autonomous activity in the isolated guinea pig bladder. Experimental Physiology. 2003; 88: 19–30. https://doi.org/10.1113/eph8802473.
- [2] Gillespie JI, Harvey IJ, Drake MJ. Agonist- and nerve-induced phasic activity in the isolated whole bladder of the guinea pig: evidence for two types of bladder activity. Experimental Physiology. 2003; 88: 343–357. https://doi.org/10.1113/eph8802536.
- [3] Drake MJ, Harvey IJ, Gillespie JI, Van Duyl WA. Localized contractions in the normal human bladder and in urinary urgency. BJU International. 2005; 95: 1002–1005. https://doi.or g/10.1111/j.1464-410X.2005.05455.x.
- [4] Fry CH, McCloskey KD. Spontaneous Activity and the Urinary Bladder. Advances in Experimental Medicine and Biology. 2019; 1124: 121-147. https://doi.org/10.1007/978-981-13-5895-1_5.
- [5] Maggi CA, Santicioli P, Meli A. Postnatal development of myogenic contractile activity and excitatory innervation of rat urinary bladder. The American Journal of Physiology. 1984; 247: R972–R978. https://doi.org/10.1152/ajpregu.1984.247.6.R972.
- [6] Sugaya K, de Groat WC. Effects of MK-801 and CNQX, glutamate receptor antagonists, on bladder activity in neonatal rats. Brain Research. 1994; 640: 1–10. https://doi.org/10.1016/ 0006-8993(94)91850-3.
- [7] Széll EA, Somogyi GT, de Groat WC, Szigeti GP. Developmental changes in spontaneous smooth muscle activity in the neonatal rat urinary bladder. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology. 2003; 285: R809–R816. https://doi.org/10.1152/ajpregu.00641.2002.
- [8] Longhurst P. Developmental aspects of bladder function. Scandinavian Journal of Urology and Nephrology. Supplementum. 2004; 11–19. https://doi.org/10.1080/03008880410015129.
- [9] Sugaya K, de Groat WC. Influence of temperature on activity of the isolated whole bladder preparation of neonatal and adult rats. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology. 2000; 278: R238–R246. https://doi.org/10.1152/ajpregu.2000.278.1.R238.
- [10] Kanai A, Roppolo J, Ikeda Y, Zabbarova I, Tai C, Birder L, et al. Origin of spontaneous activity in neonatal and adult rat bladders and its enhancement by stretch and muscarinic agonists. American Journal of Physiology. Renal Physiology. 2007; 292: F1065–F1072. https://doi.org/10.1152/ajprenal.00229.2006.



- [11] Ng YK, de Groat WC, Wu HY. Smooth muscle and neural mechanisms contributing to the downregulation of neonatal rat spontaneous bladder contractions during postnatal development. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology. 2007; 292: R2100–R2112. https://do i.org/10.1152/ajpregu.00779.2006.
- [12] Burnstock G. Purinergic signalling in lower urinary tract. In Abbracchio MP, Williams M (eds.) Handbook of Experimental Pharmacology, Purinergic and Pyrimidinergic Signalling I: Molecular, Nervous and Urogenitary System Function, Vol151 (pp. 423–515). Springer Berlin: Heidelberg. 2001. https://doi.or g/10.1007/978-3-662-09604-8.
- [13] Büyüknacar HSG, Göçmen C, de Groat WC, Kumcu EK, Wu HY, Onder S. Differential effect of L-cysteine in isolated whole-bladder preparations from neonatal and adult rats. The Journal of Pharmacology and Experimental Therapeutics. 2010; 333: 228–235. https://doi.org/10.1124/jpet.109.161661.
- [14] Oh SJ, Lee KH, Kim SJ, Kim KW, Kim KM, Choi H. Active properties of the urinary bladder: *in vitro* comparative studies between adult and neonatal rats. BJU International. 2000; 85: 1126–1133. https://doi.org/10.1046/j.1464-410x.2000.00621.x.
- [15] Morera FJ, Wolff D, Vergara C. External copper inhibits the activity of the large-conductance calcium- and voltage-sensitive potassium channel from skeletal muscle. The Journal of Membrane Biology. 2003; 192: 65–72. https://doi.org/10.1007/s00232-002-1064-y.
- [16] Trombley PQ, Shepherd GM. Differential modulation by zinc and copper of amino acid receptors from rat olfactory bulb neurons. Journal of Neurophysiology. 1996; 76: 2536–2546. https: //doi.org/10.1152/jn.1996.76.4.2536.
- [17] Sharonova IN, Vorobjev VS, Haas HL. High-affinity copper block of GABA(A) receptor-mediated currents in acutely isolated cerebellar Purkinje cells of the rat. The European Journal of Neuroscience. 1998; 10: 522–528. https://doi.org/10.1046/j. 1460-9568.1998.00057.x.
- [18] Acuña-Castillo C, Morales B, Huidobro-Toro JP. Zinc and copper modulate differentially the P2X4 receptor. Journal of Neurochemistry. 2000; 74: 1529–1537. https://doi.org/10.1046/j. 1471-4159.2000.0741529.x.
- [19] Horning MS, Trombley PQ. Zinc and copper influence excitability of rat olfactory bulb neurons by multiple mechanisms. Journal of Neurophysiology. 2001; 86: 1652–1660. https://doi.org/10.1152/jn.2001.86.4.1652.
- [20] Göçmen C, Giesselman B, de Groat WC. Effect of neocuproine, a copper(i) chelator, on rat bladder function. The Journal of Pharmacology and Experimental Therapeutics. 2005; 312: 1138– 1143. https://doi.org/10.1124/jpet.104.076398.
- [21] Xiong K, Peoples RW, Montgomery JP, Chiang Y, Stewart RR, Weight FF, et al. Differential modulation by copper and zinc of P2X2 and P2X4 receptor function. Journal of Neurophysiology. 1999; 81: 2088–2094. https://doi.org/10.1152/jn.1999.81. 5.2088.
- [22] Coddou C, Lorca RA, Acuña-Castillo C, Grauso M, Rassendren F, Huidobro-Toro JP. Heavy metals modulate the activity of the purinergic P2X4 receptor. Toxicology and Applied Pharmacology. 2005; 202: 121–131. https://doi.org/10.1016/j.taap.2004.06.015.
- [23] Patel HJ, Giembycz MA, Keeling JE, Barnes PJ, Belvisi MG. Inhibition of cholinergic neurotransmission in guinea pig trachea by NS1619, a putative activator of large-conductance, calciumactivated potassium channels. The Journal of Pharmacology and Experimental Therapeutics. 1998; 286: 952–958.
- [24] Delgado R, Vergara C, Wolff D. Divalent cations as modulators of neuronal excitability: emphasis on copper and zinc. Biological Research. 2006; 39: 173–182. https://doi.org/10.4067/s0716-97602006000100019.

- [25] Andersson KE, Arner A. Urinary bladder contraction and relaxation: physiology and pathophysiology. Physiological Reviews. 2004; 84: 935–986. https://doi.org/10.1152/physrev. 00038.2003.
- [26] Rivera L, Brading AF. The role of Ca2+ influx and intracellular Ca2+ release in the muscarinic-mediated contraction of mammalian urinary bladder smooth muscle. BJU International. 2006; 98: 868–875. https://doi.org/10.1111/j.1464-410X.2006.06431.
- [27] Yu W. Reviving Cav1.2 as an attractive drug target to treat bladder dysfunction. FASEB Journal. 2022; 36: e22118. https://doi.org/10.1096/fj.202101475R.
- [28] de Groat WC, Yoshimura N. Pharmacology of the lower urinary tract. Annual Review of Pharmacology and Toxicology. 2001; 41: 691–721. https://doi.org/10.1146/annurev.pharmtox .41.1.691.
- [29] Birder LA, Barrick SR, Roppolo JR, Kanai AJ, de Groat WC, Kiss S, et al. Feline interstitial cystitis results in mechanical hypersensitivity and altered ATP release from bladder urothelium. American Journal of Physiology. Renal Physiology. 2003; 285: F423–F429. https://doi.org/10.1152/ajprenal.00056.2003.
- [30] Göçmen C, Kumcu EK, Büyüknacar HS, Onder S, Singirik E. Neocuproine, a copper (I) chelator, potentiates purinergic component of vas deferens contractions elicited by electrical field stimulation. Pharmacology. 2005; 75: 69–75. https://doi.org/10.1159/000087007.
- [31] Kennedy C, Tasker PN, Gallacher G, Westfall TD. Identification of atropine- and P2X1 receptor antagonist-resistant, neurogenic contractions of the urinary bladder. The Journal of Neuroscience. 2007; 27: 845–851. https://doi.org/10.1523/JNEURO SCI.3115-06.2007.
- [32] Fry CH, McCloskey KD. Purinergic signalling in the urinary bladder - When function becomes dysfunction. Autonomic Neuroscience: Basic & Clinical. 2021; 235: 102852. https://doi.org/ 10.1016/j.autneu.2021.102852.
- [33] Kendig DM, Ets HK, Moreland RS. Effect of type II diabetes on male rat bladder contractility. American Journal of Physiology. Renal Physiology. 2016; 310: F909–F922. https://doi.org/ 10.1152/ajprenal.00511.2015.
- [34] Husted SE, Nedergaard OA. Dual inhibitory action of ATP on adrenergic neuroeffector transmission in rabbit pulmonary artery. Acta Pharmacologica et Toxicologica. 1985; 57: 204– 213. https://doi.org/10.1111/bcpt.1985.57.3.204.
- [35] Andersson KE. Bladder activation: afferent mechanisms. Urology. 2002; 59: 43–50. https://doi.org/10.1016/s0090-4295(01) 01637-5.
- [36] Pandita RK, Andersson KE. Intravesical adenosine triphosphate stimulates the micturition reflex in awake, freely moving rats. The Journal of Urology. 2002; 168: 1230–1234. https://doi.org/ 10.1016/S0022-5347(05)64631-9.
- [37] Mumtaz FH, Khan MA, Thompson CS, Morgan RJ, Mikhailidis DP. Nitric oxide in the lower urinary tract: physiological and pathological implications. BJU International. 2000; 85: 567–578. https://doi.org/10.1046/j.1464-410x.2000.00459.x.
- [38] Sancho M, Ferrero JJ, Triguero D, Torres M, Garcia-Pascual A. Altered neuronal and endothelial nitric oxide synthase expression in the bladder and urethra of cyclophosphamide-treated rats. Nitric Oxide: Biology and Chemistry. 2014; 39: 8–19. https://doi.org/10.1016/j.niox.2014.04.002.
- 39] Artim DE, Kullmann FA, Daugherty SL, Wu HY, de Groat WC. Activation of the nitric oxide-cGMP pathway reduces phasic contractions in neonatal rat bladder strips via protein kinase G. American Journal of Physiology. Renal Physiology. 2009; 297: F333–F340. https://doi.org/10.1152/ajprenal.00207.2009.
- [40] Persson K, Igawa Y, Mattiasson A, Andersson KE. Effects of inhibition of the L-arginine/nitric oxide pathway in the rat lower



- urinary tract *in vivo* and *in vitro*. British Journal of Pharmacology. 1992; 107: 178–184. https://doi.org/10.1111/j.1476-5381. 1992.tb14483.x.
- [41] Eser N, Buyuknacar HS, Kumcu E, Gocmen C. The Copper (I) Chelator Neocuproine Inhibits Mouse Bladder Function, but not the Copper (II) Chelator Cuprizone. Osmangazi Journal of Medicine. 2021; 43: 439–447.
- [42] Ng YK, de Groat WC, Wu HY. Muscarinic regulation of neonatal rat bladder spontaneous contractions. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology. 2006; 291: R1049–R1059. https://doi.org/10.1152/ajpreg u.00236.2006.
- [43] Wrzosek A. The potassium channel opener NS1619 modulates calcium homeostasis in muscle cells by inhibiting SERCA. Cell Calcium. 2014; 56: 14–24. https://doi.org/10.1016/j.ceca.2014.
- [44] Macmillan S, Sheridan RD, Chilvers ER, Patmore L. A comparison of the effects of SCA40, NS 004 and NS 1619 on large conductance Ca(2+)-activated K+ channels in bovine tracheal smooth muscle cells in culture. British Journal of Pharmacology. 1995; 116: 1656–1660. https://doi.org/10.1111/j.1476-5381. 1995.tb16387.x.
- [45] Imai T, Okamoto T, Yamamoto Y, Tanaka H, Koike K, Shigenobu K, et al. Effects of different types of K+ channel modulators on the spontaneous myogenic contraction of guinea-pig

- urinary bladder smooth muscle. Acta Physiologica Scandinavica. 2001; 173: 323–333. https://doi.org/10.1046/j.1365-201X.2001.00908.x.
- [46] Buckner SA, Milicic I, Daza AV, Coghlan MJ, Gopalakrishnan M. Spontaneous phasic activity of the pig urinary bladder smooth muscle: characteristics and sensitivity to potassium channel modulators. British Journal of Pharmacology. 2002; 135: 639–648. https://doi.org/10.1038/sj.bjp.0704499.
- [47] Nakahara T, Mitani A, Kubota Y, Maruko T, Sakamoto K, Tanaka Y, et al. MaxiK channel-triggered negative feedback system is preserved in the urinary bladder smooth muscle from streptozotocin-induced diabetic rats. Journal of Smooth Muscle Research. 2004; 40: 97–109. https://doi.org/10.1540/jsmr.40. 97.
- [48] Hristov KL, Chen M, Kellett WF, Rovner ES, Petkov GV. Large-conductance voltage- and Ca2+-activated K+ channels regulate human detrusor smooth muscle function. American Journal of Physiology. Cell Physiology. 2011; 301: C903–C912. https://doi.org/10.1152/ajpcell.00495.2010.
- [49] Soder RP, Petkov GV. Large conductance Ca2+ -activated K+ channel activation with NS1619 decreases myogenic and neurogenic contractions of rat detrusor smooth muscle. European Journal of Pharmacology. 2011; 670: 252–259. https://doi.org/10.1016/j.ejphar.2011.08.013.

