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Targeting HIF-1 α signaling pathway for gastric cancer treatment

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Gastric cancer is a common malignancy with high mortality and limited therapeutic options. Hypoxia inducible factor-1 α (HIF-1 α) is the key responder adapted to tumor hypoxia. Under hypoxia conditions, HIF-1 α signaling is activated and responsible for cell biology associated with metabolism, inflammation, vascular homeostasis and tumorigenesis. Activation of HIF-1 α signaling has been observed in a series of cancers including gastric cancer. Moreover, a variety of oncogene products may be induced by HIF-1 α expression. Pharmacologic manipulation of HIF-1 α may provide a novel therapeutic approach to gastric cancer. In this review, we summarize the recent information on HIF-1 α in gastric cancer with special focus on the mechanism underlying HIF-1 α signaling effects on proliferation, apoptosis, angiogenesis, EMT and drug-resistance of cancer cells, thereby predicting new therapeutic agents for better management of this malignancy.

1. Introduction

Gastric cancer is one of the leading causes of cancer-related death worldwide (Bertuccio et al. 2009). Despite of advances in the detection and treatment of gastric cancer including surgery, chemotherapy and targeted therapies, many metastatic gastric cancers still persist after primary therapy and eventually become resistant to chemotherapy (Karimi et al. 2014). For these refractory diseases, new drugs specifically inhibiting critical signaling pathways which promote tumorigenesis and tumor development are needed to improve the prognosis of patients.

As a common feature of many cancers, hypoxia contributes to the regulation of cell metabolism and chemoradiotherapy resistance leading to therapeutic failure (Huang et al. 2014; Pucci et al. 2018). In response to tumor hypoxia, upregulation of hypoxia-inducible factor 1 (HIF-1) activates the downstream genes participated in crucial aspects of cancer biology comprising angiogenesis, cell survival, glucose metabolism and invasion (Wang et al. 2014). As a subunit, HIF-1 α is responsible for the function of HIF-1 and its overexpression is observed in a variety of cancer tissues compared with the respective normal tissues, including colorectal, breast, lung and gastric carcinomas (Cai et al. 2016; Kitajima and Miyazaki 2013; Li et al. 2016; Yan et al. 2017). However, the role of HIF-1 α in tumor development as a promoter or inhibitor remains controversial. On one hand, HIF-1 α was verified to be positively correlated with tumor metastasis and negatively with prognosis of patients (Chen et al. 2014; Liu et al. 2018). On the contrary, Carmeliet et al. (1998) showed that HIF-1 α inhibits tumor growth through reducing proliferation and increasing apoptosis of embryonic stem cells due to its association with p53 (An et al. 1998).

Along with the significant progress in our understanding of HIF-1 α , its roles in gastric cancer development have become increasingly clear. At the same time, precise treatment is gradually replacing traditional chemotherapy due to its better therapeutic effects and slighter side effects. Appearance of many HIF-1 α inhibitors significantly improves the prognosis of advanced gastric cancer patients, indicating that HIF-1 α may become a prospective target for gastric cancer treatment.

2. HIF-1 α signaling pathway

HIF-1 consists of alpha (α) and beta (β) subunits which are both helix-loop-helix transcription factors. Among these two subunits, the expression of HIF-1 α

can be high-induced by hypoxia leading to promotion of tumor growth, while HIF-1 α protein levels remain constant and not concerned with oxygen concentrations, implying that HIF-1 α is the decisive factor of HIF-1 transcriptional activity and hypoxia response (Huang et al. 1996). When oxygen is available, HIF-1 α is targeted to be disrupted by hydroxylases, the key oxygen sensor including prolyl-hydroxylases (PHD1-3, especially PHD2) and asparaginyl hydroxylase (such as factor inhibiting HIF-1 (FIH-1)) which mediate ubiquitination and proteasomal degradation of HIF-1 α (Berra et al. 2003; Kaelin and Ratcliffe 2008; Lando et al. 2002; Mahon et al. 2001; Townley-Tilson et al. 2015). Activities of prolyl and asparaginyl hydroxylase can be inhibited by hypoxia, resulting in relief of inhibition on HIF-1 α (Jaakkola et al. 2001; Kaelin and Ratcliffe 2008; Maxwell et al. 1999). Activated HIF-1 α is transferred to the nucleus and acts as a transcription factor with binding to HIF-1 α to form a heterodimer. This heterodimer subsequently binds to hypoxia response elements (HREs) containing the recognizing sequence of 5'-(A/G)CGTG-3', leading to transactivation or inhibition of down-stream target genes associated with metabolism, inflammation, vascular homeostasis and tumorigenesis (Fig. 1). Hundreds of genes are discovered to be regulated by HIF-1 α , of them several genes are found to establish close network with HIF-1 α including NF κ B1, BRCA1, STAT3, STAT1, MMP1, TIMP1, TLR2, FCGR3A, IRF1, FAS, and TFF3 (Wang et al. 2014).

3. The role of HIF-1 α signaling in gastric cancer

3.1. HIF-1 α and proliferation

Accelerated proliferation is an essential character of almost all types of cancer cells leading to a rapid growth of the tumor. Based on recent study results, we identified three major mechanisms by which HIF-1 α induces proliferation of gastric cancer cells: acti-

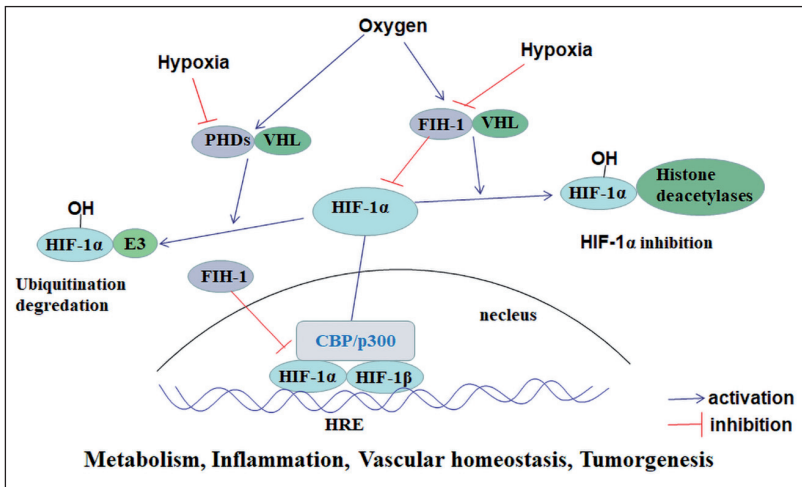


Fig. 1: HIF-1 α signaling activation. When oxygen is available, HIF-1 α is targeted to be disrupted by hydroxylases, the key oxygen sensor including prolyl-hydroxylases (PHDs) and asparaginyl hydroxylase (such as factor inhibiting HIF-1 (FIH-1)). PHDs mediate HIF-1 α ubiquitination degradation through binding to the von Hippel-Lindau (VHL) protein and recruitment of an E3 ubiquitin-protein ligase. FIH-1 inhibits HIF-1 α via the following ways: preventing combination of HIF-1 α to the transcriptional co-activators CBP and p300; binding to HIF-1 α directly and inhibiting its transactivation; binding to the transcriptional corepressor VHL which recruit histone deacetylases to inhibit HIF-1 α . Activities of prolyl and asparaginyl hydroxylase can be inhibited by hypoxia, which resulting in activation of HIF-1 α . Activated HIF-1 α is transferred to nucleus and bind to HIF-1 α to form a heterodimer which binds to hypoxia response elements (HREs) containing the recognizing sequence of 5'-(A/G)CGTG-3', leading to transactivation or inhibition of down-stream target genes associated with metabolism, inflammation, vascular homeostasis and tumorigenesis.

vating PI3K/AKT pathway (Ardyanto et al. 2006; Zhang et al. 2018); upregulating Forkhead box M1 (FoxM1)/PARI pathway (Bai et al. 2018; Zhang et al. 2018), and interaction with long non-coding RNAs (lncRNAs) (Huang et al. 2017; Liu et al. 2016; Zhang et al. 2015). For example, dysregulation of lncRNA related to HIF-1 α has been recognized in many kinds of cancers. In gastric cancer, lncRNAs associated with promoted proliferation induced by HIF-1 α consist of GAPLINC and PVT1. HIF-1 α can increase proliferation activities of gastric cancer cells by inducing high expression of GAPLINC, it can also be a target gene of lncRNA PVT1 which contributes to enhanced cell proliferation (Huang et al. 2017; Liu et al. 2016; Zhang et al. 2015). What's more, several studies suggest that downregulation of HIF-1 α or its activator inhibits proliferative activity of gastric cancer cells (Li et al. 2016; Wang et al. 2017; Zhang et al. 2018). In conclusion, apart from the traditional proliferation-associated pathways, lncRNAs have shown its unique role in HIF-1 α pathway by promoting cell proliferation as well as metastasis of gastric cancer.

3.2. HIF-1 α and apoptosis

Cell apoptosis is the rudimentary life phenomenon initiated with caspase activation which is induced by two main pathways: mitochondria-mediated apoptosis pathway and extracellular signal-mediated apoptosis pathway. Apoptosis-resistance, a common feature of cancer cells, serves as a promoter of gastric cancer malignance and chemo-resistance. A great deal of evidence suggests that HIF-1 α can inhibit apoptosis of gastric cancer cells. In the first place, several studies establishing tumor xenograft models on mice show that HIF-1 α inhibition can reduce the growth speed of gastric tumor by inducing apoptosis (Guan et al. 2017; Shen et al. 2012; Tanaka et al. 2015; Wakiyama and Kitajima 2017; Wang et al. 2017). The agonist of HIF-1 α was discovered to significantly inhibit apoptosis of gastric cancer cells leading to reduced sensitivity to chemotherapeutic agents (Yan et al. 2015). Secondly, substantial evidences suggest that HIF-1 α suppresses mitochondrial apoptosis via interplaying with microRNAs (miR-421, miR-18a) (Ge et al. 2016; Wu et al. 2015) and weakening reactive oxygen species (ROS)/mitochondria pathway which is responsible for the apoptosis of cancer cells (Li et al. 2014; Wakiyama and Kitajima 2017). Moreover, apart from mitochondrial apoptosis, HIF-1 α can also inhibit extracellular signal-mediated apoptosis by activating MEK/ERK pathway *in vivo* and *in vitro* (Liu et al. 2010). Taken together, HIF-1 α contributes to apoptosis-resistance by regulating both mitochondrial apoptosis and extracellular signal-mediated apoptosis. Fortunately, the treatment outcomes for gastric cancer are going to be improved with the development of studies in the apoptosis effect of HIF-1 α inhibition.

3.3 HIF-1 α signaling and angiogenesis

Angiogenesis is known as the sprouting of new capillaries from pre-existing vessels supported by hypoxia cancer cells to provide oxygen for rapid tumor growth (Rey and Semenza 2010). It is known that HIF-1 α signaling is critical for gastric cancer angiogenesis through activating angiogenic growth factors (VEGF, SDF1, ANGPT2, PGF, PDGFB and SCF) (Bosch-Marce et al. 2007; Forsythe et al. 1996; Kelly et al. 2003) and microRNAs (such as miR-382, acting as an angiogenic oncogene) (Seok et al. 2014). A correlation analysis through immunohistochemical staining of resected gastric cancer specimens indeed verified the key role of HIF-1 α in gastric tumor angiogenesis (Zhan et al. 2013). Several animal studies utilizing gastric cancer xenografts show that many HIF-1 α inhibitors suppress tumor angiogenesis by reducing the expression of VEGF under hypoxia conditions (Fang et al. 2017; Kim et al. 2016; Ko et al. 2016; Lee et al. 2014). As the most common metastatic pattern in advanced gastric cancer, peritoneal dissemination is well-recognized to be positively correlated with tumor angiogenesis induced by HIF-1 α (Miyake et al. 2013; Yoshikawa et al. 2006). However, Hiraki et al. (2012) suggested that knockdown of HIF-1 α in gastric cancer cells promotes the progression of peritoneal dissemination by upregulating MMP-1, indicating that the role of HIF-1 α signaling in peritoneal dissemination development is not simply *via* HIF-1 α /VEGF pathway. These data suggest that angiogenesis induced by HIF-1 α is a main pathological characteristic of gastric cancer. HIF-1 α mediates angiogenesis in gastric cancer mainly through two ways: HIF-1 α /VEGF axis and HIF-1 α /microRNAs pathway. The former has been studied extensively, while the relationship between HIF-1 α and microRNAs in tumor angiogenesis is not fully realized. During the development of gastric cancer, HIF-1 α can be an upstream-regulator or a downstream-target gene of microRNAs (Seok et al. 2014; Wu et al. 2015; Zheng et al. 2013), suggesting the microRNA/HIF-1 α /microRNA pathway existing to regulate angiogenesis in gastric cancer.

3.4. HIF-1 α and EMT

Epithelial to mesenchymal transition (EMT) appears as reduced epithelial markers like E-cadherin, desmoplankins, claudins, cytokeratins and the gain of mesenchymal markers such as N-cadherin, vimentin, Snail, etc. Hypoxia is one of the conditions able to trigger EMT which plays a vital role in cancer metastasis. The induction role of HIF-1 α on EMT was assigned to many human cancers (Deng et al. 2018; Kim et al. 2018). In gastric cancer, HIF-1 α is demonstrated to enhance invasion and metastasis of gastric cancer *via* inducing EMT (Liu et al. 2014; Yang et al. 2017; Zhang et al. 2017). Plenty study results confirm that HIF-1 α upregulates EMT

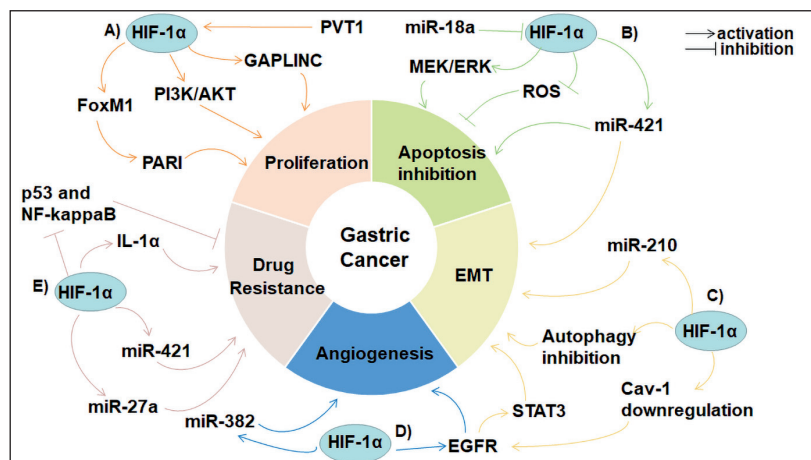


Fig. 2: HIF-1 α signaling pathway in regulating gastric cancer development. A) Activated by lncRNA PVT1, HIF-1 α promotes proliferation of gastric cancer cells through activating PI3K/AKT, FoxM1/PARI pathways and lncRNA GAPLINC. B) Apoptosis of gastric cancer cells can be inhibited by HIF-1 α which activates MEK/ERK pathway and miR-421 as well as controlling production of reactive oxygen species (ROS). MiR-18a was hypothesized to induce apoptosis through antagonism to HIF-1 α . C) HIF-1 α involves in EMT though upregulating miR-210 and inhibiting autophagy. Besides, HIF-1 α /caveolin-1(Cav-1)/EGFR/STAT3 pathway also contributes to EMT initiation. D) Angiogenesis developed in gastric cancer is mainly induced through HIF-1 α /EGFR and HIF-1 α /miR-382 pathway. E) MiR-421, miR-27a and IL-1 α upregulated by HIF-1 α contribute to enhanced chemo-resistance. What's more, HIF-1 α can reduce sensitivity of gastric cancer cells to chemotherapeutic agents by suppressing p53 and NF-kappaB activity.

by mediating EMT-associated transcription factors and signaling pathways in gastric cancer. First, HIF-1 α /microRNAs pathway serves as a promising activator of EMT. Studies *in vivo* and *in vitro* show that miR-421 and miR-210 upregulated by HIF-1 α both promote metastasis or chemo-resistance of gastric cancer by regulating EMT-related proteins (Ge et al. 2016; Yu et al. 2015; Zhang et al. 2016). MiR-224, upregulated by HIF-1 α , is responsible for migration of gastric cancer and is confirmed to induce EMT in other type of cancers including malignant melanoma and colorectal cancer (Amankwatia et al. 2015; He et al. 2017; Knoll et al. 2014). These data indicate that the HIF-1 α /microRNAs/EMT pathway is involved in metastasis of gastric cancer. Second, tumor-associated macrophages (TAMs) and autophagy are associated with EMT induced by HIF-1 α (Qin et al. 2015; Zhang et al. 2017). What's more, many transcription factors targeted by HIF-1 α are suggested to activate EMT covering Snail, Rab11-FIP2, KLF8, caveolin-1 (Cav-1) and RhoE (Dong et al. 2016; Kannan et al. 2014; Liu et al. 2014; Yang et al. 2017; Zhou et al. 2011). Except for microRNAs, lncRNAs also exhibit important roles during the HIF-1 α -induced tumor EMT process (Cai et al. 2017). With more and more studies discovering the function of microRNA or lncRNA in tumor progression, the impact of these non-coding RNAs on invasiveness of gastric cancer will be better known.

4. HIF-1 α and chemo-resistance of gastric cancer

Chemotherapeutic drugs effective for gastric cancer treatment embrace 5-fluorouracil (5-FU), platinum compounds, anthracyclines, taxanes and irinotecan. Although this therapy was observed to be well tolerated and had yielded good responses in clinical trials, acquired resistance is common in clinical practice. Involvement of HIF-1 α in chemo-resistance during gastric cancer treatment has been well recognized. Combined with recent study data, HIF-1 α is suggested to induce anti-cisplatin effects through three major mechanisms: first, dysregulating microRNAs (such as miR-421, miR-27a) and lncRNAs (such as PVT1) (Ge et al. 2016; Zhang et al. 2015; Zhao et al. 2015); second, upregulating inflammatory factors (such as IL-1 α) (Xuan and Wang 2017); third, suppressing activities of p53 and NF-kappaB (Rohwer et al. 2010). Through these three main pathways, cisplatin-induced apoptosis is significantly abrogated. In addition, HIF-1 α is considered as an independent risk factor for gastric cancer relapse after 5-FU chemotherapy because of its function with inhibiting 5-FU-mediated apoptosis (Nakamura et al. 2010). Recent studies reveal that glycolysis inhibitors can reverse HIF-1 α -induced resistance to 5-FU (Chen et al. 2015; Xuan et al. 2014), indicating that a high level of glycolysis under hypoxia is conducive to 5-FU-resistance. Finally, drug sensitivity can be enhanced by HIF-1 α knockdown (Zhao et al. 2016). Except for cisplatin and 5-FU, HIF-1 α also induces resistance to oxaliplatin, vincristine, epirubicin, oxaliplatin and capecitabine during chemotherapy (Liu et al. 2008; Mangia et

al. 2015). All evidence suggests that the efficacy of chemotherapeutic drugs can be impaired by HIF-1 α , which has become a main obstacle during the therapy of gastric cancer.

5. Contribution of HIF-1 α inhibitors to prevent gastric cancer

Due to the pivotal role of HIF-1 α in promoting gastric carcinogenesis and progression as well as inducing resistance to chemotherapy (Fig. 2), the effect of HIF-1 α inhibitors on gastric cancer treatment aroused our concern. There are many kinds of HIF-1 α inhibitors exhibiting anti-gastric cancer effect in recent studies through the following mechanisms. Firstly, with deep understanding of the relationship between HIF-1 α and microRNAs, HIF-1 α inhibitors are discovered to achieve an anticancer function by suppressing the release of microRNAs. The representative drug class of this category is that of proton-pump inhibitors (PPIs), which have been already applied for clinical treatment of peptic ulcer and gastro-esophageal reflux disease (Guan et al. 2017; Shen et al. 2012). Secondly, HIF-1 α inhibitors can prevent progression of gastric cancer by anti-proliferative and pro-apoptotic effects. Low dose YC-1, an inhibitor of HIF-1 α combined with glucose (G) and insulin (I) treatments is established targeting hypoxic gastric cancer cells (Tanaka et al. 2015; Wakiyama and Kitajima 2017). What's more, angiogenesis of gastric cancer can be obstructed by HIF-1 α inhibitors such as celecoxib or berberine (Lin et al. 2004; Wang et al. 2006). Finally, some natural compounds like quercetin and curcumin are discovered to suppress gastric cancer by inhibiting HIF-1 α (Wang et al. 2011, 2017).

These preclinical trials offer promises for the anti-cancer effects of drugs inhibiting HIF-1 α in gastric cancer, in addition to established therapeutic options for gastric cancer.

6. Conclusion and future perspectives

HIF-1 α signaling has been considered as a promoter of gastric cancer progression through mediating enhanced proliferation, angiogenesis, EMT, drug-resistance and inhibited apoptosis of cancer cells. Despite the impressive progress of HIF-1 α signaling in gastric cancer, there are several challenges for a systematic clinical application of HIF-1 α inhibitors. First, although non coding RNAs (microRNAs and lncRNAs) have been confirmed to be a vital factor in HIF-1 α -induced cancer progression, its regulating mode with HIF-1 α is still unclear and worth to be explored more deeply. Second, a phase I clinical trial of drugs targeting HIF-1 α combined with conventional chemotherapy is still underway (ClinicalTrials.gov Identifier: NCT01049620). Further evaluation is necessary for safety and effectiveness of HIF-1 α -targeted drugs. Finally, novel drugs aimed at molecules in HIF-1 α signaling need to be developed due to the limited range of drugs directly targeting for HIF-1 α . New therapeutic strategy with HIF-1 α signaling inhib-

itors is predictable to be developed for gastric cancer clinical trials in the future.

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References

Amankwata EB, Chakravarty P, Carey FA, Weidlich S, Steele RJ, Munro AJ, Wolf CR, Smith G (2015) MicroRNA-224 is associated with colorectal cancer progression and response to 5-fluorouracil-based chemotherapy by KRAS-dependent and -independent mechanisms. *Br J Cancer* 112: 1480-1490.

An WG, Kanekal M, Simon MC, Maltepe E, Blagosklonny MV, Neckers LM (1998) Stabilization of wild-type p53 by hypoxia-inducible factor 1alpha. *Nature* 392: 405-408.

Ardyanto TD, Osaki M, Tokuyasu N, Nagahama Y, Ito H (2006) CoCl2-induced HIF-1alpha expression correlates with proliferation and apoptosis in MKN-1 cells: a possible role for the PI3K/Akt pathway. *Int J Oncol* 29: 549-555.

Bai C, Liu X, Qiu C, Zheng J (2018) FoxM1 is regulated by both HIF-1alpha and HIF-2alpha and contributes to gastrointestinal stromal tumor progression. *Gastric Cancer*: 10.1007/s10120-018-0846-6

Berra E, Benizri E, Ginouves A, Volmat V, Roux D, Pouyssegur J (2003) HIF prolyl-hydroxylase 2 is the key oxygen sensor setting low steady-state levels of HIF-1alpha in normoxia. *EMBO J* 22: 4082-4090.

Bertuccio P, Chatenoud L, Levi F, Praud D, Ferlay J, Negri E, Malvezzi M, La Vecchia C (2009) Recent patterns in gastric cancer: a global overview. *Int J Cancer* 125: 666-673.

Bosch-Marce M, Okuyama H, Wesley JB, Sarkar K, Kimura H, Liu YV, Zhang H, Strazza M, Rey S, Savino L, Zhou YF, McDonald KR, Na Y, Vandiver S, Rabi A, Shaked Y, Kerbel R, Lavallee T, Semenza GL (2007) Effects of aging and hypoxia-inducible factor-1 activity on angiogenic cell mobilization and recovery of perfusion after limb ischemia. *Circ Res* 101: 1310-1318.

Cai FF, Xu C, Pan X, Cai L, Lin XY, Chen S, Biskup E (2016) Prognostic value of plasma levels of HIF-1a and PGC-1a in breast cancer. *Oncotarget* 7: 77793-77806.

Cai Q, Wang Z, Wang S, Weng M, Zhou D, Li C, Wang J, Chen E, Quan Z (2017) Long non-coding RNA LINC00152 promotes gallbladder cancer metastasis and epithelial-mesenchymal transition by regulating HIF-1alpha via miR-138. *Open Biol* 7: pii: 160247.

Carmeliet P, Dor Y, Herbert JM, Fukumura D, Brusselmans K, Dewerchin M, Neeman M, Bono F, Abramovitch R, Maxwell P, Koch CJ, Ratcliffe P, Moons L, Jain RK, Collen D, Keshert E (1998) Role of HIF-1alpha in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature* 394: 485-490.

Chen F, Zhuang M, Zhong C, Peng J, Wang X, Li J, Chen Z, Huang Y (2015) Baicalin reverses hypoxia-induced 5-FU resistance in gastric cancer AGS cells through suppression of glycolysis and the PTEN/Akt/HIF-1alpha signaling pathway. *Oncol Rep* 33: 457-463.

Chen L, Shi Y, Yuan J, Han Y, Qin R, Wu Q, Jia B, Wei B, Wei L, Dai G, Jiao S (2014) HIF-1 alpha overexpression correlates with poor overall survival and disease-free survival in gastric cancer patients post-gastrectomy. *PLoS One* 9: e90678.

Deng SJ, Chen HY, Ye Z, Deng SC, Zhu S, Zeng Z, He C, Liu ML, Huang K, Zhong JX, Xu FY, Li Q, Liu Y, Wang CY, Zhao G (2018) Hypoxia-induced LncRNA-BX111 promotes metastasis and progression of pancreatic cancer through regulating ZEB1 transcription. *Oncogene* 10.1038/s41388-018-0382-1

Dong W, Qin G, Shen R (2016) Rab11-FIP2 promotes the metastasis of gastric cancer cells. *Int J Cancer* 138: 1680-1688.

Fang S, Hong H, Li L, He D, Xu Z, Zuo S, Han J, Wu Q, Dai Z, Cai W, Ma J, Shao C, Gao G, Yang X (2017) Plasminogen kringle 5 suppresses gastric cancer via regulating HIF-1alpha and GRP78. *Cell Death Dis* 8: e3144.

Forsythe JA, Jiang BH, Iyer NV, Agani F, Leung SW, Koos RD, Semenza GL (1996) Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Mol Cell Biol* 16: 4604-13.

Ge X, Liu X, Lin F, Li P, Liu K, Geng R, Dai C, Lin Y, Tang W, Wu Z, Chang J, Lu J, Li J (2016) MicroRNA-421 regulated by HIF-1alpha promotes metastasis, inhibits apoptosis, and induces cisplatin resistance by targeting E-cadherin and caspase-3 in gastric cancer. *Oncotarget* 7: 24466-24482.

Guan XW, Zhao F, Wang JY, Wang HY, Ge SH, Wang X, Zhang L, Liu R, Ba Y, Li HL, Deng T, Zhou LK, Bai M, Ning T, Zhang HY, Huang DZ (2017) Tumor microenvironment interruption: a novel anti-cancer mechanism of Proton-pump inhibitor in gastric cancer by suppressing the release of microRNA-carrying exosomes. *Am J Cancer Res* 7: 1913-1925.

He C, Wang L, Zhang J, Xu H (2017) Hypoxia-inducible microRNA-224 promotes the cell growth, migration and invasion by directly targeting RASSF8 in gastric cancer. *Mol Cancer* 16: 35.

Hiraki M, Kitajima Y, Kai K, Nakamura J, Hashiguchi K, Noshiro H, Miyazaki K (2012) Knockdown of hypoxia-inducible factor-1alpha accelerates peritoneal dissemination via the upregulation of MMP-1 expression in gastric cancer cell lines. *Exp Ther Med* 4: 355-362.

Huang D, Li C, Zhang H (2014) Hypoxia and cancer cell metabolism. *Acta Biochim Biophys Sin (Shanghai)* 46: 214-921.

Huang LE, Arany Z, Livingston DM, Bunn HF (1996) Activation of hypoxia-inducible transcription factor depends primarily upon redox-sensitive stabilization of its alpha subunit. *J Biol Chem* 271: 32253-32259.

Huang T, Liu HW, Chen JQ, Wang SH, Hao LQ, Liu M, Wang B (2017) The long noncoding RNA PVT1 functions as a competing endogenous RNA by sponging miR-186 in gastric cancer. *Biomed Pharmacother* 88: 302-308.

Jaakkola P, Mole DR, Tian YM, Wilson MI, Gaskell SJ, von Kriegsheim A, Hebestreit HF, Mukherji M, Schofield CJ, Maxwell PH, Pugh CW, Ratcliffe PJ (2001) Targeting of HIF-alpha to the von Hippel-Lindau ubiquitylation complex by O2-regulated prolyl hydroxylation. *Science* 292: 468-472.

Kaelin WG, Jr., Ratcliffe PJ (2008) Oxygen sensing by metazoans: the central role of the HIF hydroxylase pathway. *Mol Cell* 30: 393-402.

Kannan A, Krishnan A, Ali M, Subramaniam S, Halagowder D, Sivasithamparam ND (2014) Caveolin-1 promotes gastric cancer progression by up-regulating epithelial to mesenchymal transition by crosstalk of signalling mechanisms under hypoxic condition. *Eur J Cancer* 50: 204-215.

Karimi P, Islami F, Anandasabapathy S, Freedman ND, Kamangar F (2014) Gastric cancer: descriptive epidemiology, risk factors, screening, and prevention. *Cancer Epidemiol Biomarkers Prev* 23: 700-713.

Kelly BD, Hackett SF, Hirota K, Oshima Y, Cai Z, Berg-Dixon S, Rowan A, Yan Z, Campochiaro PA, Semenza GL (2003) Cell type-specific regulation of angiogenic growth factor gene expression and induction of angiogenesis in nonischemic tissue by a constitutively active form of hypoxia-inducible factor 1. *Circ Res* 93: 1074-1081.

Kim EJ, Kwon KA, Lee YE, Kim JH, Kim SH, Kim JH (2018) Korean Red Ginseng extract reduces hypoxia-induced epithelial-mesenchymal transition by repressing NF-kappaB and ERK1/2 pathways in colon cancer. *J Ginseng Res* 42: 288-297.

Kim SY, Ko YS, Park J, Choi Y, Park JW, Kim Y, Pyo JS, Yoo YB, Lee JS, Lee BL (2016) Forkhead Transcription Factor FOXO1 inhibits angiogenesis in gastric cancer in relation to SIRT1. *Cancer Res Treat* 48: 345-354.

Kitajima Y, Miyazaki K (2013) The critical impact of HIF-1a on gastric cancer biology. *Cancers (Basel)* 5: 15-26.

Knoll S, Furst K, Kowtharapu B, Schmitz U, Marquardt S, Wolkenhauer O, Martin H, Putzer BM (2014) E2F1 induces miR-224/452 expression to drive EMT through TXNIP downregulation. *EMBO Rep* 15: 1315-1329.

Ko YS, Cho SJ, Park J, Choi Y, Lee JS, Youn HD, Kim WH, Kim MA, Park JW, Lee BL (2016) Hypoxic inactivation of glycogen synthase kinase-3beta promotes gastric tumor growth and angiogenesis by facilitating hypoxia-inducible factor-1 signaling. *Apmis* 124: 748-756.

Lando D, Peet DJ, Gorman JJ, Whelan DA, Whitelaw ML, Bruick RK (2002) FIH-1 is an asparaginyl hydroxylase enzyme that regulates the transcriptional activity of hypoxia-inducible factor. *Genes Dev* 16: 1466-1471.

Lee SH, Bae SC, Kim KW, Lee YM (2014) RUNX3 inhibits hypoxia-inducible factor-1alpha protein stability by interacting with prolyl hydroxylases in gastric cancer cells. *Oncogene* 33: 1458-1467.

Li J, Zhou X, Lan X, Zeng G, Jiang X, Huang Z (2016) Repression of PES1 expression inhibits growth of gastric cancer. *Tumour Biol* 37: 3043-3049.

Li P, Zhao QL, Wu LH, Jawaaid P, Jiao YF, Kadowaki M, Kondo T (2014) Isofraxidin, a potent reactive oxygen species (ROS) scavenger, protects human leukemia cells from radiation-induced apoptosis via ROS/mitochondria pathway in p53-independent manner. *Apoptosis* 19: 1043-1053.

Li W, Zong S, Shi Q, Li H, Xu J, Hou F (2016) Hypoxia-induced vasculogenic mimicry formation in human colorectal cancer cells: Involvement of HIF-1a, Claudin-4, and E-cadherin and Vimentin. *Sci Rep* 6: 37534.

Lin S, Tsai SC, Lee CC, Wang BW, Liou JY, Shyu KG (2004) Berberine inhibits HIF-1alpha expression via enhanced proteolysis. *Mol Pharmacol* 66: 612-619.

Liu L, Ning X, Sun L, Zhang H, Shi Y, Guo C, Han S, Liu J, Sun S, Han Z, Wu K, Fan D (2008) Hypoxia-inducible factor-1 alpha contributes to hypoxia-induced chemoresistance in gastric cancer. *Cancer Sci* 99: 121-122.

Liu L, Zhang H, Sun L, Gao Y, Jin H, Liang S, Wang Y, Dong M, Shi Y, Li Z, Fan D (2010) ERK/MAPK activation involves hypoxia-induced MGR1-Ag/37LRP expression and contributes to apoptosis resistance in gastric cancer. *Int J Cancer* 127: 820-829.

Liu L, Zhao X, Zou H, Bai R, Yang K, Tian Z (2016) Hypoxia promotes gastric cancer malignancy partly through the HIF-1alpha dependent transcriptional Activation of the Long Non-coding RNA GAPLINC. *Front Physiol* 7: 420.

Liu N, Wang Y, Zhou Y, Pang H, Zhou J, Qian P, Liu L, Zhang H (2014) Kruppel-like factor 8 involved in hypoxia promotes the invasion and metastasis of gastric cancer via epithelial to mesenchymal transition. *Oncol Rep* 32: 2397-2404.

Liu X, Wang Y, Sun L, Min J, Liu J, Chen D, Zhang H, Zhang H, Zhou Y, Liu L (2018) Long noncoding RNA BC005927 upregulates EPHB4 and promotes gastric cancer metastasis under hypoxia. *Cancer Sci* 109: 988-1000.

Mahon PC, Hirota K, Semenza GL (2001) FIH-1: a novel protein that interacts with HIF-1alpha and VHL to mediate repression of HIF-1 transcriptional activity. *Genes Dev* 15: 2675-2686.

Mangia A, Caldarola L, Dell'Endice S, Scarpi E, Saragoni L, Monti M, Santini D, Brunetti O, Simone G, Silvestris N (2015) The potential predictive role of nuclear NHERF1 expression in advanced gastric cancer patients treated with epirubicin/oxaliplatin/capecitabine first line chemotherapy. *Cancer Biol Ther* 16: 1140-1147.

Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, Wykoff CC, Pugh CW, Maher ER, Ratcliffe PJ (1999) The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature* 399: 271-275.

Miyake S, Kitajima Y, Nakamura J, Kai K, Yanagihara K, Tanaka T, Hiraki M, Miyazaki K, Noshiro H (2013) HIF-1alpha is a crucial factor in the development of peritoneal dissemination via natural metastatic routes in scirrhous gastric cancer. *Int J Oncol* 43: 1431-1440.

Nakamura J, Kitajima Y, Kai K, Hashiguchi K, Hiraki M, Noshiro H, Miyazaki K (2010) HIF-1alpha is an unfavorable determinant of relapse in gastric cancer patients who underwent curative surgery followed by adjuvant 5-FU chemotherapy. *Int J Cancer* 127: 1158-1171.

- Pucci P, Rescigno P, Sumanasuriya S, de Bono J, Crea F (2018) Hypoxia and noncoding RNAs in taxane resistance. *Trends Pharmacol Sci* 39: 695-709.
- Qin W, Li C, Zheng W, Guo Q, Zhang Y, Kang M, Zhang B, Yang B, Li B, Yang H, Wu Y (2015) Inhibition of autophagy promotes metastasis and glycolysis by inducing ROS in gastric cancer cells. *Oncotarget* 6: 39839-39854.
- Rey S, Semenza GL (2010) Hypoxia-inducible factor-1-dependent mechanisms of vascularization and vascular remodelling. *Cardiovasc Res* 86: 236-242.
- Rohwer N, Dame C, Haugstetter A, Wiedenmann B, Detjen K, Schmitt CA, Cramer T (2010) Hypoxia-inducible factor 1alpha determines gastric cancer chemosensitivity via modulation of p53 and NF-kappaB. *PLoS One* 5: e12038.
- Seok JK, Lee SH, Kim MJ, Lee YM (2014) MicroRNA-382 induced by HIF-1alpha is an angiogenic miR targeting the tumor suppressor phosphatase and tensin homolog. *Nucleic Acids Res* 42: 8062-8072.
- Shen Y, Wu Y, Chen M, Shen W, Huang S, Zhang L, Zou X (2012) Effects of pantoprazole as a HIF-1alpha inhibitor on human gastric adenocarcinoma sgc-7901 cells. *Neoplasma* 59: 142-149.
- Tanaka T, Kitajima Y, Miyake S, Yanagihara K, Hara H, Nishijima-Matsunobu A, Baba K, Shida M, Wakiyama K, Nakamura J, Noshiro H (2015) The apoptotic effect of HIF-1alpha inhibition combined with glucose plus insulin treatment on gastric cancer under hypoxic conditions. *PLoS One* 10: e0137257.
- Townley-Tilson WH, Pi X, Xie L (2015) The role of oxygen sensors, hydroxylases, and HIF in cardiac function and disease. *Oxid Med Cell Longev* 2015: 676893.
- Wakiyama K, Kitajima Y (2017) Low-dose YC-1 combined with glucose and insulin selectively induces apoptosis in hypoxic gastric carcinoma cells by inhibiting anaerobic glycolysis. *Sci Rep* 7: 12653.
- Wang J, Ni Z, Duan Z, Wang G, Li F (2014) Altered expression of hypoxia-inducible factor-1alpha (HIF-1alpha) and its regulatory genes in gastric cancer tissues. *PLoS One* 9: e99835.
- Wang J, Wu K, Bai F, Zhai H, Xie H, Du Y, Liang J, Han S, Chen Y, Lin T, Fan D (2006) Celecoxib could reverse the hypoxia-induced angiopoietin-2 upregulation in gastric cancer. *Cancer Lett* 242: 20-27.
- Wang K, Liu R, Li J, Mao J, Lei Y, Wu J, Zeng J, Zhang T, Wu H, Chen L, Huang C, Wei Y (2011) Quercetin induces protective autophagy in gastric cancer cells: involvement of Akt-mTOR- and hypoxia-induced factor 1alpha-mediated signaling. *Autophagy* 7: 966-978.
- Wang XP, Wang QX, Lin HP, Chang N (2017) Anti-tumor bioactivities of curcumin on mice loaded with gastric carcinoma. *Food Funct* 8: 3319-3326.
- Wu F, Huang W, Wang X (2015) microRNA-18a regulates gastric carcinoma cell apoptosis and invasion by suppressing hypoxia-inducible factor-1alpha expression. *Exp Ther Med* 10: 717-722.
- Xuan Y, Hur H, Ham IH, Yun J, Lee JY, Shim W, Kim YB, Lee G, Han SU, Cho YK (2014) Dichloroacetate attenuates hypoxia-induced resistance to 5-fluorouracil in gastric cancer through the regulation of glucose metabolism. *Exp Cell Res* 321: 219-30.
- Xuan Y, Wang YN (2017) Hypoxia/IL-1alpha axis promotes gastric cancer progression and drug resistance. *18: 511-520.*
- Yan LH, Wei WY, Cao WL, Zhang XS, Xie YB, Xiao Q (2015) Overexpression of CDX2 in gastric cancer cells promotes the development of multidrug resistance. *Am J Cancer Res* 5: 321-332.
- Yan X, Jiao SC, Zhang GQ, Guan Y, Wang JL (2017) Tumor-associated immune factors are associated with recurrence and metastasis in non-small cell lung cancer. *Cancer Gene Ther* 24: 57-63.
- Yang SW, Zhang ZG, Hao YX, Zhao YL, Qian F, Shi Y, Li PA, Liu CY, Yu PW (2017) HIF-1alpha induces the epithelial-mesenchymal transition in gastric cancer stem cells through the Snail pathway. *Oncotarget* 8: 9535-9545.
- Yoshikawa T, Tsuburaya A, Miyagi Y, Sekiguchi H, Kimura M, Cho H, Kobayashi O (2006) Up-regulation of hypoxia-inducible factor-1 alpha and VEGF mRNAs in peritoneal dissemination of patients with gastric cancer. *Anticancer Res* 26: 3849-3853.
- Yu P, Fan S, Huang L, Yang L, Du Y (2015) MIR210 as a potential molecular target to block invasion and metastasis of gastric cancer. *Med Hypotheses* 84: 209-212.
- Zhan H, Liang H, Liu X, Deng J, Wang B, Hao X (2013) Expression of Rac1, HIF-1alpha, and VEGF in gastric carcinoma: correlation with angiogenesis and prognosis. *Onkologie* 36: 102-107.
- Zhang C, Tian W, Meng L, Qu L, Shou C (2016) PRL-3 promotes gastric cancer migration and invasion through a NF-kappaB-HIF-1alpha-miR-210 axis. *J Mol Med (Berl)* 94: 401-415.
- Zhang J, Xu J, Dong Y, Huang B (2018) Downregulation of HIF-1alpha inhibits the proliferation, migration and invasion of gastric cancer by inhibiting PI3K/AKT pathway and VEGF expression. *Biosci Rep*: 10.1042/bsr20180741
- Zhang WJ, Chen C, Zhou ZH, Gao ST, Tee TJ, Yang LQ, Xu YY, Pang TH, Xu XY, Sun Q, Feng M, Wang H, Lu CL, Wu GZ, Wu S, Guan WX, Xu GF (2017) Hypoxia-inducible factor-1 alpha correlates with tumor-associated macrophages infiltration, influences survival of gastric cancer patients. *J Cancer* 8: 1818-1825.
- Zhang XW, Bu P, Liu L, Zhang XZ, Li J (2015) Overexpression of long non-coding RNA PVT1 in gastric cancer cells promotes the development of multidrug resistance. *Biochem Biophys Res Commun* 462: 227-232.
- Zhang Y, Ye X, Chen L, Wu Q, Gao Y, Li Y (2018) PARI functions as a new transcriptional target of FOXM1 involved in gastric cancer development. *Int J Biol Sci* 14: 531-541.
- Zhao Q, Li Y, Tan BB, Fan LQ, Yang PG, Tian Y (2015) HIF-1alpha induces multidrug resistance in gastric cancer cells by inducing MiR-27a. *PLoS One* 10: e0132746.
- Zhao Q, Tan BB, Li Y, Fan LQ, Yang PG, Tian Y (2016) Enhancement of drug sensitivity by knockdown of HIF-1alpha in gastric carcinoma cells. *Oncol Res* 23: 129-136.
- Zheng Y, Li S, Ding Y, Wang Q, Luo H, Shi Q, Hao Z, Xiao G, Tong S (2013) The role of miR-18a in gastric cancer angiogenesis. *Hepatogastroenterology* 60: 1809-1813.
- Zhou J, Li K, Gu Y, Feng B, Ren G, Zhang L, Wang Y, Nie Y, Fan D (2011) Transcriptional up-regulation of RhoE by hypoxia-inducible factor (HIF)-1 promotes epithelial to mesenchymal transition of gastric cancer cells during hypoxia. *Biochem Biophys Res Commun* 415: 348-354.