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A composite of graphene oxide and iron oxide nanoparticles for targeted drug delivery of temozolomide

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Received October 21, 2019, accepted January 17, 2020

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Pharmazie 75: 313-317 (2020)

doi: 10.1691/ph.2020.9170

A magnetic targeting nanoparticle based on graphene oxide-ferroferric oxide (GO-Fe₃O₄) was investigated as a potential drug delivery vehicle. The formation of GO/Fe₃O₄ hybrid material was confirmed by Fourier transform infrared spectroscopy, X-ray diffraction, scanning electron microscopy and transmission electron microscopy. The GO/Fe₃O₄ hybrid still shows a higher saturation magnetization of 58.42 emu/g after coating with graphene oxide. Drug loading and releasing experiments demonstrate the GO-Fe₃O₄ hybrid has a good loading capacity of (6.47±0.08) mg/mg for temozolomide and a satisfactory release under slightly acidic condition. The MTT assays of glioma C6 cells exhibits the GO-Fe₃O₄ hybrid does not display toxicity with the concentration ranged from 40 to 120 µg/mL *in vitro*, while the complex of temozolomide loaded on GO/Fe₃O₄ has a better inhibitory effect on the proliferation of rat glioma C6 cells. All results suggest the prepared GO/Fe₃O₄ has potential applications in targeted anticancer drug delivery.

1. Introduction

With the development of modern industrialization, nanoscale drug carriers have emerged as a bridge linking nanotechnology and drug delivery, involving liposomes, microspheres (Deng et al. 2005), nanoparticles (Pankhurst et al. 2003) and polymeric shells (Noble et al. 2004; Bédard et al. 2008). The medicine is loaded on these nanoscale carriers, which have the characteristics of targeting, improve solubility and bioavailability. However, the loading capacity of the current nanoscale drug carrier is still low. Therefore, developing an efficient drug carrier is critical in drug carrier research.

Recently, graphene oxide (GO) has attracted great attention due to its large specific surface area for efficient drug binding and abundant functional groups (such as -OH, -COOH, -O-, -C=O, etc.) for improving solubility (Yang et al. 2016; Wang et al. 2011). As a drug carrier, GO and drugs could form complexes through π - π conjugated, hydrogen bonding or an ionic reaction. Moreover, GO derivatives are reported to have significantly improved tumor passive targeting effect and higher tumor uptake *via* the enhanced permeability and retention effect (Pramanik et al. 2019). Magnetic nanoparticles can be manipulated by an external magnetic field and play a targeting role (Guo et al. 2016; Li et al. 2018). As one of the magnetic substances, Fe₃O₄ is widely used in biomedical applications such as drug delivery, magnetic resonance imaging, magnetic hyperthermia, and cell labeling and separation (Cole et al. 2011; Schleich et al. 2014). Combining the characteristics of GO and Fe₃O₄, the magnetic GO/Fe₃O₄ carrier has the ability of passive targeting, and can be located at the designated site and release the drugs under the magnetic field *in vitro*. Furthermore, cell viability and metabolic activity decrease significantly when cells are exposed to high iron oxide nanoparticle concentrations.

Temozolomide (TMZ), a novel small molecule alkylating agent with activity in the central nervous system, is able to pass the blood-brain barrier (Prabhu et al. 2017; Nordling-David et al. 2017). Despite its important role in chemotherapy for glioblastoma treatment, the cytotoxicity and bioavailability of TMZ to normal cells cannot be ignored. Besides, due to its short half-life, the concentration of TMZ around the brain rarely reaches therapeutic

levels. To improve its application, a lot of experimental work has been reported in the literature. However, low drug loading capacity still limited the use of TMZ.

In the context of the current studies for magnetic nanoparticles, the aim of this work is to synthesize a GO-Fe₃O₄ nanoparticles hybrid by chemical deposition method and investigate the binding of TMZ on the hybrid. The morphology, microstructure and magnetic properties were examined by transmission electron microscopy (TEM), scanning electron microscopy (SEM), Fourier transform infrared spectrometer (FT-IR) and X-ray diffractometer (XRD), respectively. The loading and release behavior of TMZ was also evaluated. The results demonstrated that the Fe₃O₄/GO composites have high loading capacity, favorable magnetism and low cytotoxicity, which is helpful to conduct the further study of loading and releasing drugs.

2. Investigations, results and discussion

2.1. Synthesis and characterization of samples

The SEM and TEM images of GO and GO-Fe₃O₄ at different magnifications were shown in Figs. 1 and 2, respectively. It can be seen that GO showed two-dimensional structure, similar to graphene, which was consistent with previous reports (Li et al. 2012). Different from GO, the structure of GO in GO-Fe₃O₄ was distorted (Figs. 1C and 1D), the uniform Fe₃O₄ particles were distributed randomly on the surface or edges of GO (Figs. 2C and 2D). The particle diameters of GO and GO-Fe₃O₄ measured by dynamic light scattering were 202.4±15.1 and 239.8±17.4 nm, respectively. And the nanocomposites dispersed in water showed negative zeta potential values of 34.2±3.2 mV (GO) and 38.7±4.1 mV (GO-Fe₃O₄). The enlarged particle size and zeta potential could be attributed to chemical impurities retained in GO (Yue et al. 2012). Some scholars suggested that it may be related to higher drug loading capacity (Chai et al. 2017).

The crystal phases of GO and GO-Fe₃O₄ were analyzed by X-Ray Diffraction (XRD). As presented in Fig. 3(a), the diffraction peak of GO appeared at 11.8° (001). Six diffraction peaks of GO-Fe₃O₄ appeared at 31.5° (220), 36.47° (311), 44.09° (400), 53.87° (422),

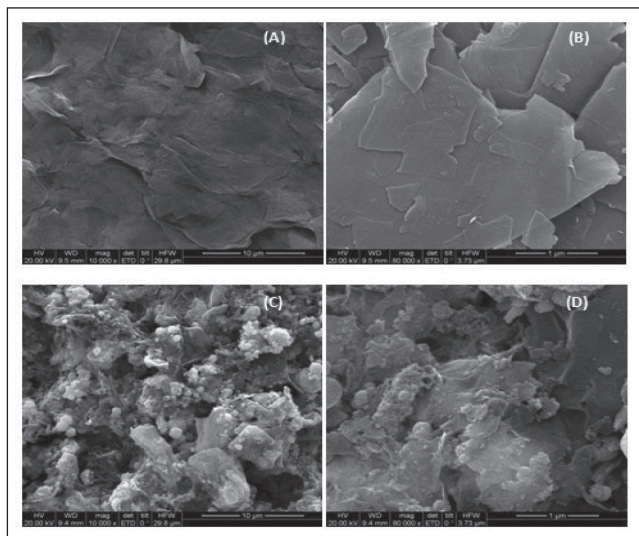


Fig. 1: SEM images of GO (A, B) and GO-Fe₃O₄ (C, D)

58.62° (511), and 63.75° (440), respectively (see Fig. 3b), which are well-matched with standard Fe₃O₄ (JCPDS. Card NO.19-0629). It can be found out that the diffraction peak of GO at 11.80° (001) did not appeared in the peaks of GO-Fe₃O₄, which may be attributed to the structural distortion of GO in GO-Fe₃O₄ system (Madadrang et al. 2012).

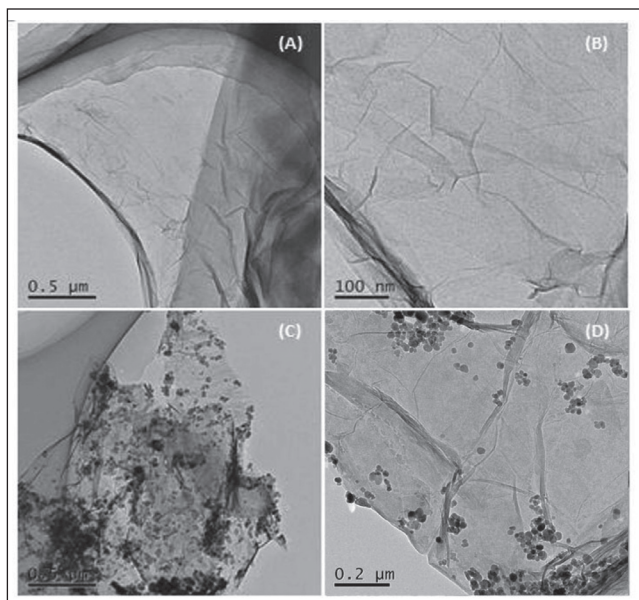


Fig. 2: TEM images of GO (A, B) and GO-Fe₃O₄ (C, D)

FT-IR analysis was carried out in the 400–4000 cm⁻¹ range to investigate the chemical bonding of GO, Fe₃O₄ and GO-Fe₃O₄ hybrid, and the findings are presented in Fig. 3(c). From this Fig., the strong absorption peak of GO-Fe₃O₄ at 3425 cm⁻¹ was the stretching vibration peak of –OH, while the peak at 1632 cm⁻¹ could be attributed to aromatic C=C vibrations of the skeletal vibrations for unoxidized graphitic domains (Ouyang et al. 2015). The new peak at 563 cm⁻¹ corresponded to the stretching vibration of the Fe–O bond, indicating the formation of Fe–O bonds between GO and Fe₃O₄. The formation of GO-Fe₃O₄ was confirmed by FT-IR spectra.

2.2. Magnetism of GO-Fe₃O₄

A vibrating sample magnetometer (VSM) was used for characterizing the magnetic properties of samples. From Fig. 3(d), the

measured saturation magnetization value of Fe₃O₄ and GO-Fe₃O₄ was 75.17 and 58.42 emu/g respectively. The result showed that GO-Fe₃O₄ was still superparamagnetic, and the reduction of the saturation magnetization value of 6.75 emu/g for GO-Fe₃O₄ attributed to the existence of the GO. Furthermore, the insert image exhibited a dispersion and quick separation process of the GO-Fe₃O₄. It is easy to see that the GO-Fe₃O₄ is well dispersed in water with the absence of an external magnetic field and can be easily separated in the presence of an external magnetic field.

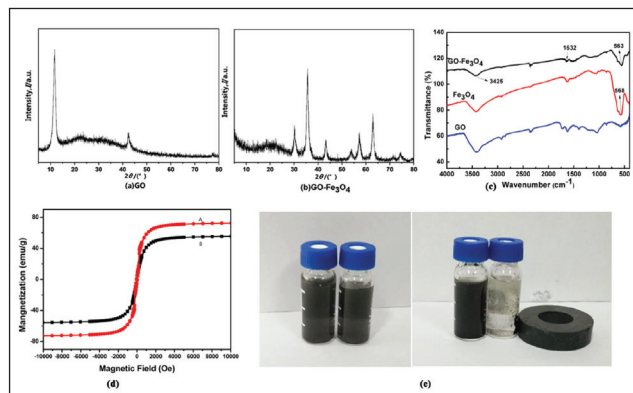


Fig. 3: XRD patterns of GO (a), GO-Fe₃O₄ (b), FT-IR spectra of GO, Fe₃O₄ and GO-Fe₃O₄ (c), hysteresis curve (M-H curve) of GO (A) and GO-Fe₃O₄ (B) at room temperature (d), changes in the distribution of GO-Fe₃O₄ under the external magnetic field (e).

2.3. TMZ loaded on GO-Fe₃O₄

The maximum drug loading capacity is an important factor for evaluating the therapeutic potential of a carrier. In this study, TMZ was loaded on GO-Fe₃O₄ surface through hydrogen bonding and intermolecular adsorption between GO and TMZ. As shown in Fig. 4(a), initially the drug loading capacity of GO-Fe₃O₄ increased sharply with the concentration of TMZ increasing till the TMZ concentration reached to 0.7 mg/ml; further increase the TMZ concentration reached to 1.5 mg/ml, drug loading capacity increased slightly; loading capacity dropped when TMZ concentration was greater than 1.5 mg/ml. The maximum drug loading was 6.47±0.08 mg/mg at the concentration of TMZ at 1.5 mg/ml. The drug loading efficiency of GO-Fe₃O₄ was calculated according to TMZ standard curve, and the loading efficiency at peak drug loading was 43.10±0.11 %. The result showed the drug loading efficiency of GO-Fe₃O₄ (ranged from 40% to 90%) was higher than that of other drug carriers, for example, the loading capacity of liposomes was usually below 40%. Therefore, GO-Fe₃O₄ is an excellent nanocarrier for delivery of TMZ-targeted formulations.

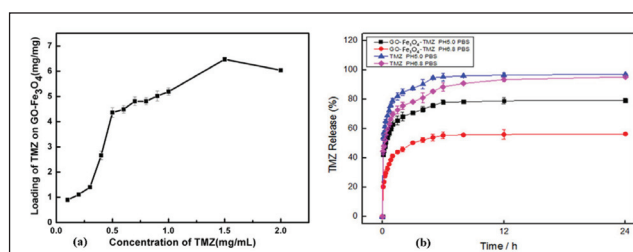


Fig. 4: Drug loading capacity of GO-Fe₃O₄ carrier at different concentrations of TMZ (n=6) (a), *In vitro* release profiles of TMZ at different pH (5.0 and 6.8) at 37 °C (b).

2.4. TMZ released from GO-Fe₃O₄-TMZ

Given the high loading capacity of GO-Fe₃O₄ composite toward TMZ, it may be used as a potential drug carrier. However, drug release behavior from a carrier are affected by many external

Table: In vitro release model fitting of TMZ and GO-FA-TMZ

	Models	Regression equations	R ²
TMZ	zero-order kinetics	$Q=3.3977t + 69.151$	0.6433
	first-order kinetics	$Q=85.8058*(1-\exp(-5.93086*t))$	0.5272
	Higuchi	$Q=14.0077*(t^{1/2})+59.02836$	0.8373
	Ritger-Peppas	$Q=75.5*(t^{0.12})$	0.9658
	Weibull	$Q=99.006*(1-\exp(-2.5033*(t+0.1296)^{0.4065}))$	0.9948
GO/Fe ₃ O ₄ -TMZ	zero-order kinetics	$Q=2.9868t + 54.602$	0.6410
	first-order kinetics	$Q=69.57*(1-\exp(-5.189*t))$	0.5375
	Higuchi	$Q=12.2331*(t^{1/2})+45.8177$	0.8555
	Ritger-Peppas	$Q=60.1889*(t^{0.13362})$	0.9714
	Weibull	$Q=80.3386-49.2799*\exp(-0.9231*t)^{0.5631}$	0.9956

factors, such as pH values, the interactions between the drug and carrier, and their physical and chemical properties. To clarify the release behavior of TMZ from GO-Fe₃O₄-TMZ composites, the release rates of TMZ under different conditions of 5.0 and 6.8 at 37 °C were investigated in the following experiments. As shown in Fig. 4(b), the TMZ released slowly from GO-Fe₃O₄-TMZ composites and a slightly acidic condition could be suitable for TMZ releases. The cumulative amount of TMZ from GO-Fe₃O₄-TMZ composites reached 74.86% at 8 h under the weak acid condition of pH 5.0. However, a fewer cumulative amount of 57.24% at 8 h was observed in a neutral condition of pH 6.8. As described in the literature, the hydrogen-bonding interaction between TMZ and GO-Fe₃O₄ is more firmly under neutral than under acidic conditions, resulting in an inefficient release (Pourjavadi et al. 2015; Kassaei et al. 2011). As the extracellular tissues of tumors, intracellular lysosomes and endosomes are acidic, this sensitive property of slightly acid for GO-Fe₃O₄-TMZ can be beneficial for the drug release in the acidic tumor microenvironment.

In addition, the release behavior of TMZ from GO-Fe₃O₄-TMZ composites was studied by fitting experimental data to five different kinetic models (zero-order kinetics, first-order kinetics, Higuchi, Ritger-Peppas, and Weibull models) (Maity and Sa 2016; Sang et al. 2018). The fitting equations are displayed in the Table, which shows the relation coefficient (R²) of five kinds of release curves. Compared with other equations, the Weibull model has the largest R² value, thus the best-fit equation was the Weibull equation. The result of this study was consistent with the theory.

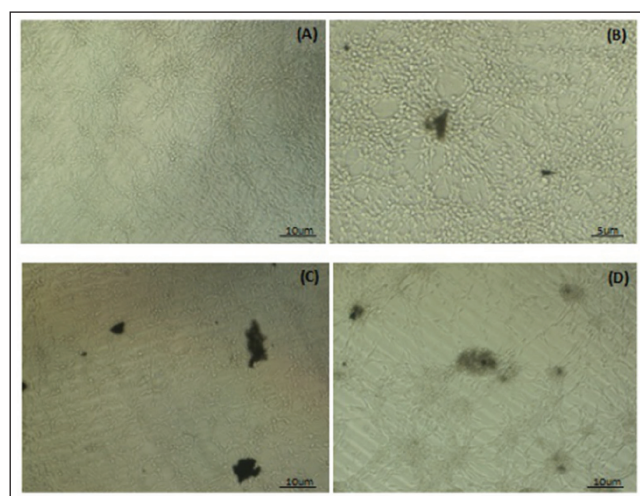


Fig. 5: Morphology of C6 cells cultured with GO-Fe₃O₄ after different times (A: 0 h, B: 24 h, C: 48 h, D: 72 h).

2.5. Cytotoxicity experiments

To evaluate the cytotoxicity of GO-Fe₃O₄-TMZ to tumor cells, MTT assays were performed and rat glioma C6 cells were selected. First, the cytotoxicity of GO-Fe₃O₄ was tested and results were shown in Fig. 5. The morphology of C6 cell cultured with GO-Fe₃O₄ for different periods (24 h, 48 h, and 72 h) was pretty much the same as initial morphology of C6 cell, no obvious changes were identified even at a higher concentration of 80 µg/ml. Furthermore, results of cell viability experiments showed that the cell viability of C6 cells was more than 91% after cultured with GO-Fe₃O₄ at the concentration of 120 µg/ml for 72 h (see Fig. 6(a)). All above results indicated the non-cytotoxic property of GO-Fe₃O₄. The cytotoxicity test result of GO-Fe₃O₄-TMZ is shown in Fig. 6(b). Clearly, the inhibition rate increased with the increase in concentration of GO-Fe₃O₄-TMZ and culture time, which was consistent with the change rule of TMZ inhibition rate (Fig. 6(c)). After C6 cells were cultured using GO-Fe₃O₄-TMZ under the concentration of 600 µg/ml for 72 h, the cell inhibition rate of GO-Fe₃O₄-TMZ reached to 89.37%. That is to say, GO-Fe₃O₄-TMZ had a better inhibitory effect on the proliferation of rat glioma C6 cells, and presented the time- and dose-dependent characteristic. In addition, it can be found that inhibition rate of GO-Fe₃O₄-TMZ was higher than that of TMZ in the same condition after 48 h (Fig. 6(d)), which may be attributed to the sustained-release property of GO-Fe₃O₄-TMZ.

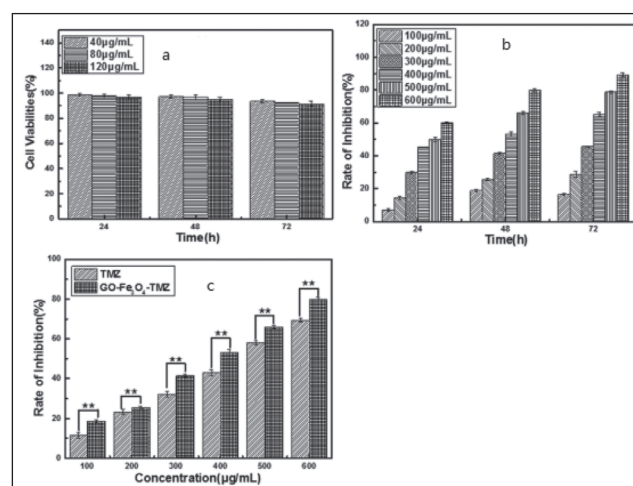


Fig. 6: MTT assay for the GO/Fe₃O₄ (a), GO/Fe₃O₄-TMZ (b) and comparison of TMZ and GO-Fe₃O₄-TMZ (c) The results in Fig. 6(c) has subtracted the effect of cells by themselves or cells with GO-Fe₃O₄. Statistical testing was performed using two-way ANOVA. (**P < 0.01)

In summary, the objective of this study was accomplished by designing GO-Fe₃O₄ nanoparticles of TMZ. With combining of GO sheets with Fe₃O₄ nanoparticles, the GO/Fe₃O₄ showed multi-

functional characterization of GO sheets and magnetic properties of Fe_3O_4 . The $\text{GO-Fe}_3\text{O}_4$ displayed a strong magnetic property and could be controlled by an external magnetic field. Moreover, the $\text{GO-Fe}_3\text{O}_4$ has a high drug loading capacity for TMZ. According to the experimental results, $\text{GO-Fe}_3\text{O}_4$ -TMZ showed good biocompatibility and pH-dependent release behavior. Thus, the developed $\text{GO/Fe}_3\text{O}_4$ -TMZ nanoparticles can be used for the potential treatment of brain tumors.

3. Experimental

3.1. Materials

TMZ was purchased from Biolang Biotechnology Co., Ltd. (Shanghai, China). GO was obtained from Fenghua Material Development Co., Ltd. (Hubei, China). Ham's F12K medium was obtained from GIBCO (Grand Island, NY, USA). Ferric chloride hexahydrate ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$), ferrous chloride tetrahydrate ($\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$), dimethyl sulfoxide (DMSO) and ammonia ($\text{NH}_3 \cdot \text{H}_2\text{O}$) used in this study were of analytical grade.

3.2. Synthesis of $\text{GO-Fe}_3\text{O}_4$

$\text{GO-Fe}_3\text{O}_4$ nanocomposites were synthesized by the co-precipitation method according to the literature with slight modification (Huang et al. 2017). In brief, FeCl_3 solution and FeCl_2 solution (molar ratio of $\text{Fe}^{2+}:\text{Fe}^{3+}=1:2$) were slowly added into graphene oxide dispersion (1 mg/mL) under nitrogen for 1 h. Then, an ammonia solution was injected into the solution and the reaction was continued for 4 h at 80 °C. Afterwards, the obtained black precipitate was collected and washed with distilled water three times to remove reactants. The $\text{GO-Fe}_3\text{O}_4$ nanocomposites were dried in a vacuum oven at 40 °C before use.

3.3. Characterization of $\text{GO-Fe}_3\text{O}_4$

The morphology and microstructure of $\text{GO-Fe}_3\text{O}_4$ were observed through a S-4800 scanning electron microscope (SEM, Hitachi, Japan) operating at 20.00 kV and Tecnai G220 transmission electron microscope (TEM, FEI, USA). FT-IR spectra were recorded on a 650-Fourier Transform Infrared Spectrometer (Tianjin GangDong Sci. & Tech. Development Co., Ltd., China) with the wavenumber ranged from 400 cm^{-1} to 4000 cm^{-1} . X-ray diffraction (XRD) was performed by the 7000-X-ray diffractometer (Shimadzu, Japan) in a scanning range of 5 ° to 80 ° with the tube voltage of 40 kV and current of 30 mA. The hysteresis loop of $\text{GO-Fe}_3\text{O}_4$ was measured by the MPMS DynaCool hysteresis loop instrument (Quantum Design, USA) with the scanning range from -20kOe to 20kOe.

3.4. TMZ loading and release of $\text{GO-Fe}_3\text{O}_4$ -TMZ

TMZ at different concentrations were mixed with $\text{GO-Fe}_3\text{O}_4$ (the concentration of 0.1 mg/mL) (1:1) in a shaker at room temperature for 24 h. The magnetic complex of $\text{GO-Fe}_3\text{O}_4$ loaded with TMZ ($\text{GO-Fe}_3\text{O}_4$ -TMZ) was separated by centrifugation at 10,000 rpm for 0.5 h. The clear supernatant at the top was collected, and the concentration of TMZ in the supernatant was measured by HPLC. The loading capacity of TMZ in $\text{GO-Fe}_3\text{O}_4$ was calculated based on the following formula:

$$\text{Drug loading capacity} = (W_{\text{initial dose}} - W_{\text{residual dose in solution}}) / W_{\text{carrier}} \quad (1)$$

where $W_{\text{initial dose}}$ is the initial weight of TMZ before loading, $W_{\text{residual dose in solution}}$ is the weight of residual TMZ in solution after loading, and W_{carrier} is the weight of $\text{GO-Fe}_3\text{O}_4$.

The release behavior of TMZ from $\text{GO-Fe}_3\text{O}_4$ was investigated by dynamic membrane dialysis. The drug-loaded $\text{GO-Fe}_3\text{O}_4$ was dispersed in PBS with pH of 5.0 and 6.8, respectively. Samples of 3 ml of eluate were taken after certain time interval (0.083, 0.167, 0.25, 0.33, 0.5, 0.67, 0.83, 1, 1.5, 2, 3, 4, 5, 6, 8, 12, 24 h) for the determination of dissolution rate, then the same volume of PBS buffer was added. The concentration of TMZ was measured by HPLC, and the cumulative release rate was calculated based on the following formula:

$$W = (C_t V + \sum C_i V_i) / (C_0 V_0 - C_1)$$

where W is the cumulative release rate, V is the volume of phosphate buffer, C_0 is the initial TMZ concentration, V_0 is the initial TMZ volume, C_1 is the TMZ concentration in supernatant after centrifugation, V_i is the volume of supernatant by centrifugation after loading, C_i is the concentration of TMZ in buffer solution, and V_i represents the volume of the buffer solution taken out at this time.

In this experiment, five different kinetic models (zero-order kinetics, first-order kinetics, Higuchi, Ritger-Peppas and Weibull models) were adopted to explain the release behavior of TMZ in $\text{GO-Fe}_3\text{O}_4$ -TMZ system, where the optimum kinetic model was selected based on the values of R^2 .

3.5. Cell culture and cytotoxicity experiments

In order to investigate the targeted and therapeutic effects of $\text{GO-Fe}_3\text{O}_4$ -TMZ to tumor cells, rat glioma C6 cells were selected. The rat glioma C6 cells were seeded in 96-well plates and cultured with $\text{GO-Fe}_3\text{O}_4$ -TMZ in the dark at 37 °C under the humidified atmosphere (5 % CO_2) for different time intervals (24 h, 48 h and 72 h). Meanwhile, cells also cultured with $\text{GO-Fe}_3\text{O}_4$ as control group. The morphology of rat C6 glioma cells was observed by microscope.

In a cytotoxicity experiment, rat glioma C6 cells were seeded in a 96-well plate with the density of 4×10^4 cells per well. After 24 h, the medium was discarded, and

the cells were cultured with different concentrated $\text{GO-Fe}_3\text{O}_4$ -TMZ and $\text{GO-Fe}_3\text{O}_4$ (0 mg/l, 100 mg/l, 200 mg/l, 300 mg/l, 400 mg/l, 600 mg/l, 800 mg/l), respectively. After reacted for 4 h, MTT solution (0.5 %, 10 μl) was added into each well, and the cells was further cultured for 4 h. At the end of the incubation, DMSO (100 μl) was added into each well and dissolved for 10 min. The optical density (OD) was measured by using the micro-plate reader (Multiskan FC, Thermo Electron Co., America) at the wavelength of 490 nm. The rat glioma C6 cell line was used to evaluate the drug delivery and imaging capability of $\text{GO-Fe}_3\text{O}_4$ -TMZ.

3.6. Magnetic targeting of $\text{GO-Fe}_3\text{O}_4$ -TMZ *in vitro*

In this section, the C6 cells were cultured in the 96-well culture plate for 48 h with $\text{GO-Fe}_3\text{O}_4$ -TMZ as the medium. Meanwhile, a magnetic field was set on one side. The distribution of $\text{GO-Fe}_3\text{O}_4$ -TMZ and the growth status of the cells were observed.

3.7. Statistical analysis

All the experiments were performed in duplicates and the results are expressed as mean \pm standard deviation (SD). Repeated measured two-way ANOVA followed by the Bonferroni post-hoc test was used to statistically compare data.

Acknowledgements: This study was supported by Educational Department of Hebei Provincial (No. QN2015221), Science and Technology and Earthquake Bureau of Zhangjiakou (No. 1811019A), and Foundation of Hebei North University (No. YB2018019).

Conflicts of interest: None declared.

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