

Department of Cardiology<sup>1</sup>, the Second Xiangya Hospital of Central South University, Hunan; Pharmaceutical Informatics Institute<sup>2</sup>, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou, Zhejiang, China

## Intracellular heat shock protein 70: a possible therapeutic target for preventing postoperative atrial fibrillation

FENG HUANG<sup>1</sup>, JIAN-PING HUANG<sup>2</sup>, JIA-YI PAN<sup>1</sup>, ZHONG-LE BAI<sup>1</sup>, LIANG TANG<sup>1</sup>, SHENG-HUA ZHOU<sup>1</sup>

Received December 14, 2011, accepted January 6, 2012

Sheng-Hua Zhou, Department of Cardiology, the Second Xiangya Hospital, Central South University, Changsha 410011, Hunan, China  
zhougqin@21cn.com

Pharmazie 67: 747–755 (2012)

doi: 10.1691/ph.2012.1844

Postoperative atrial fibrillation is a frequent complication after cardiac surgery, associated with an increased risk of mortality and morbidity; thus, additional treatment and increasing costs of postoperative care are required. Inflammation and oxidative stress caused by ischemia-reperfusion during cardiac surgery may play an important role in the pathogenesis of postoperative atrial fibrillation. Stress-inducible heat shock proteins act as molecular chaperones that maintain cell homeostasis against stress in these events. Heat shock protein 70, the 70-kDa family of heat shock proteins, has been shown to closely associate with the incidence of postoperative atrial fibrillation in patients undergoing cardiac surgery. Extracellular heat shock protein 70 may be pro-inflammatory in the myocardial innate immune response caused by ischemia-reperfusion. In contrast, intracellular heat shock protein 70 exerts primarily anti-inflammatory and anti-apoptotic effects by preventing response to inflammatory cytokines, and inhibiting the nuclear factor Kappa B signaling pathway and different stages of mitochondrial-dependent pathways. Furthermore, the intracellular molecule can inhibit the induction and the maintenance of atrial fibrillation by attenuating Ca<sup>2+</sup> overload in injured myocardial cells. It is the intracellular heat shock protein 70, but not the extracellular molecule that holds as a therapeutic strategy for preventing postoperative atrial fibrillation.

### 1. Introduction

Open heart surgery is the most frequent surgical procedure performed with the use of cardiopulmonary bypass (CPB) and cardioplegic arrest. Though technical and technological improvements have made the operation safer, postoperative atrial fibrillation (POAF) remains the most common complication after cardiac surgery. The incidence of POAF after coronary artery bypass graft (CABG) ranges from 15% to 40% (Aranki et al. 1996; El-Chami et al. 2010; Elahi et al. 2003; Nazeri et al. 2010). These events usually occur within 2 to 5 days and often resolved within several weeks. Though generally well tolerated and seen as a temporary problem related to surgery, POAF increases the risk of stroke and is associated with significant morbidity and long-term mortality. In the meantime, POAF is invariably associated with prolonged hospitalization and significant additional costs to patient care. It was reported that patients developing POAF after CABG were hospitalized about 4.9 days longer than patients remaining in sinus rhythm, with \$10,000 to \$11,500 of additional hospital charges for CABG in the USA (Aranki et al. 1996).

### 2. Pathophysiology and mechanisms of postoperative atrial fibrillation (POAF)

The pathophysiology of POAF after cardiac surgery is not precisely known and the mechanisms are thought to be multifactorial and are influenced by preoperative, intraoperative and

postoperative factors (Fig. 1). Different risk factors have been reported, included increasing age, a previous history of atrial fibrillation (AF), male gender, hypertension, obesity, decreased left-ventricular ejection fraction, left atrial enlargement, chronic obstructive pulmonary disease, prolonged ventilation, chronic renal failure, diabetes mellitus, and rheumatic heart disease (Girerd et al. 2009; Magee et al. 2007; Mathew et al. 2004).

Inflammation and oxidative stress caused by ischemia-reperfusion during cardiac surgery may play an important role in the pathogenesis of POAF (Fig. 1). Although the relative strength of the association of inflammation and oxidative stress markers with POAF remains unclear, the role of inflammation and oxidative stress on structural and electrical remodeling is under investigation. Previous studies showed that POAF is a probable consequence of the electrophysiological disturbances associated with inflammation and reperfusion injury in cardiac surgery patients (Ascione et al. 2000; Wu et al. 2003). Inflammatory cytokines, such as interleukin (IL)-1, IL-6, IL-8, IL-10, tumor necrosis factor (TNF)-alpha, monocyte chemoattractant protein-1 (MCP-1), N-terminal pro-brain B-type natriuretic peptide (NT-proBNP) and the renin-angiotensin system have all been investigated for an association with AF (Boos et al. 2006; Li et al. 2010). Inflammation can alter atrial conduction, facilitating reentry and then predisposing to the development of POAF (Beere et al. 2000; Ishii et al. 2005).

Oxidative stress is a major contributory factor representing the unavoidable consequences of ischemia-reperfusion cycle

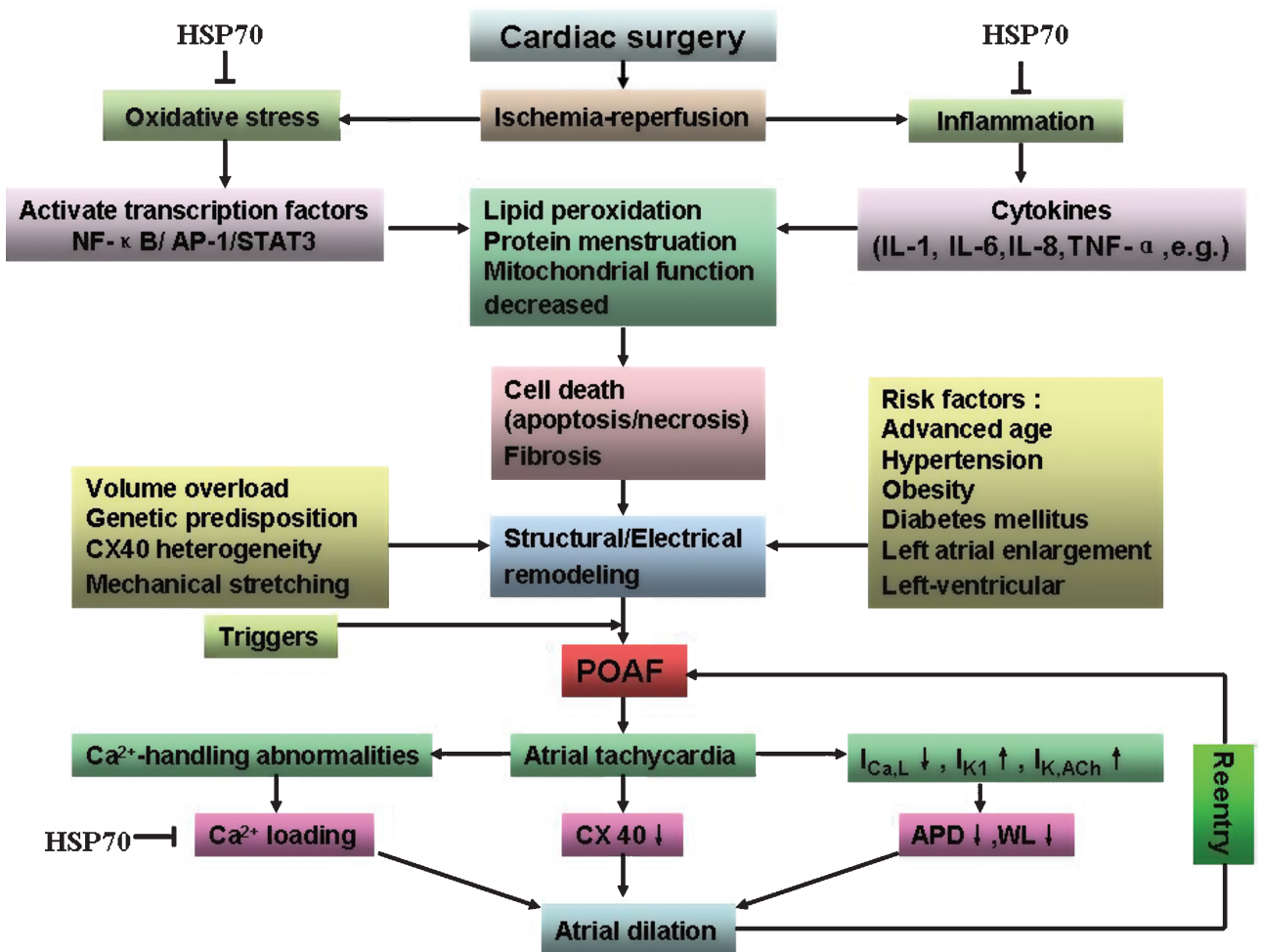


Fig. 1: Proposed hypothesize the pathogenesis of postoperative atrial fibrillation in patients undergoing cardiac surgery. NF-κB = nuclear factor Kappa B; AP-1 = transcriptional factor activating protein-1; STAT3 = signal transducer and activator of transcription 3; IL-1 = interleukin-1; IL-6 = interleukin-6; IL-8 = interleukin-8; TNF-α = tumor necrosis factor-α; POAF = postoperative atrial fibrillation;  $I_{Ca,L}$  = inward L-type  $Ca^{2+}$  channel;  $I_{K1}$  = inward rectifier  $K^+$  current;  $I_{K,ACh}$  = acetylcholine-dependent  $K^+$  current; CX40 = connexin 40; APD = action potential duration; WL = wavelength

occurring in cardiac surgery. Many studies have strongly suggested a link between oxidative stress and cardiac arrhythmias, especially AF (Li et al. 2010; Neuman et al. 2007). Oxidative stress triggers inflammation by activating nuclear factor Kappa B (NF-κB) signaling pathways and activator protein-1 (AP-1) transcription factors (Bowie and O'Neill 2000). Signal transducer and activator of transcription-3 (STAT-3) has also been reported to participate in the regulation of this signaling pathway (Hilfiker-Kleiner et al. 2005). Ischemia-reperfusion injury during cardiac surgery leads to the formation of reactive oxygen species (ROS), causing oxidative stress and a systemic inflammatory response (Matata et al. 2000). The increased ROS may lead to lipid peroxidation, breakdown of cell membranes, mitochondrial function decrease, calcium overload, cells death (apoptosis/necrosis), or fibrosis in the myocyte. Thus atrial oxidative injury can lead to the development of POAF by impairing atrial contraction, altering myofibrillar energetics and reducing atrial effective refractory period after cardiac surgery (Kim et al. 2008). Although both oxidative stress and inflammation could be separately involved in the mechanism of POAF, there is a close relation between the two processes. Nicotinamide adenine dinucleotide phosphate (NADPH) oxidase activity is potentially stimulated by cytokines, suggesting that this oxidase system may be an important link between the systemic inflammatory response to cardiac surgery and myocardial oxidative stress (Kim et al. 2008). Other pathophysiological mechanisms might also intervene as contributed factors in the development of POAF, including volume overload (Kalus et al.

2004), genetic predisposition assessed by the IL-6 promoter gene variant (Gaudino et al. 2003), and increased expression of the gap-junctional protein connexin 40 (CX40) (Dupont et al. 2001). In addition, mechanical stretching of the atrium can alter cellular electrophysiologic properties suggesting that increased intravascular volume due to postoperative mobilization of interstitial fluid could contribute to POAF (Bruins et al. 1997).

Atrial tachycardia promotes the induction and maintenance of AF, a phenomenon called atrial tachycardia remodeling. Rapid electrical activation reduces atrial refractoriness and decreases the wavelength (WL), primarily due to inward L-type  $Ca^{2+}$  channel ( $I_{Ca,L}$ ) down-regulation (Yue et al. 1997) but also via increasing inward-rectifier  $K^+$  currents such as the background current  $I_{K1}$  and a constitutively active form of acetylcholine-dependent  $K^+$  current ( $I_{K,ACh}$ ). The reduced WL caused by action potential duration (APD) abbreviation decreases the size of functional reentry circuits and promotes the induction and maintenance of AF by multiple circuit reentry (Nattel 2002; Nattel et al. 2007). Furthermore, atrial tachycardia appears likely to suppress expression of the atrial-selective CX40 (van der Velden et al. 2000), which associated with AF. In addition, atrial tachycardia remodeling also leads to atrial contractile dysfunction, mainly through  $Ca^{2+}$ -handling abnormalities, including increased cellular  $Ca^{2+}$  loading and reduced systolic  $Ca^{2+}$  transients (Sun et al. 2001; Sun et al. 1998), which causes atrial dilation that further promotes reentry (Fig. 1).

### 3. Intracellular HSP70, but not extracellular molecule prevents POAF

Heat shock proteins (HSPs) are characterized as molecular chaperones, playing a dominant role in the preservation and protection of cells and organs from oxidative stress and ischemia-reperfusion injury. The heat shock protein 70 (HSP70) family includes a number of proteins, with slightly different molecular masses and cellular localization. Two members of this family, HSP78 (glucoseregulated protein (Grp)78 or immunoglobulin-binding protein and HSP75 (mitochondrial (mt)HSP70), perform chaperone functions in the endoplasmic reticulum and mitochondria. The remaining member proteins, HSC70 (sometimes called HSP73), and its inducible isoform, HSP70 (often referred to as HSP72), are usually distributed in the cytoplasm and nucleus. Intracellular HSP70 and HSC70 function as molecular chaperones, playing important roles in protein folding and transport.

The expression of HSP70 is very low under normal physiological conditions. HSP70 may be released into the blood stream under a number of pathological conditions that lead to widespread cell death. Cardiac surgery is not only associated with an up-regulation of intracellular HSP70 in leukocytes but also may result in increasing liberation of HSP70. Many studies have reported that inducible HSP70 is released into the circulation by the myocardium due to myocardial ischemia-reperfusion after cardiac surgery (Dybdahl et al. 2004, 2002; Szerafin et al. 2008). CPB machine used in cardiac surgery may cause physical damage to the cells in circulation, leading to the induction and release of HSP70. A significant increase of serum levels of HSP70 concentration was observed in the on-pump CABG patients shortly after surgery (Lin et al. 2010). Significantly more HSP70 is released into the circulation after on-pump CABG than after off-pump CABG (Dybdahl et al. 2004, 2002; Lin et al. 2010). This difference may be explained by increased leukocyte and tissue damage or increased complexity of surgical procedures in the on-pump group. Leukocytes, including monocytes, granulocytes, and lymphocytes, are sources of elevated serum HSP70 in the inflammatory response of CABG patients (Fehrenbach et al. 2000; Lin et al. 2010).

Intracellular function of HSPs is molecular chaperone, activity counteracting the unfolding, misfolding and pathological modification of critical proteins in ischemic injury (Williams and Benjamin 2000). Glutamine has been shown to induce HSP70 in the heart, lung and liver, leading to increased nitric oxide (NO) production in an attempt to reduce the CPB-induced inflammatory response (Hayashi et al. 2002). Myocardial biopsies showed up-regulation of HSP70 transcripts and protein at the end of CABG (Schmitt et al. 2002; Szerafin et al. 2008). It appears more likely that most of HSP70 induced in the heart are not released into the circulation, but localized in the heart, which may be due to the surgical trauma, or the local ischemia caused by the clamping of coronary arteries (Lin et al. 2010). Moreover, myocardial stretch or prevention of systolic shortening may lead to increase of myocardial HSP70 in rabbits (Knowlton et al. 1991), indicating that manipulation and stretching of the heart during surgery may cause an increase of HSP70 in the heart. Intracellular HSP70 not only prevents the adverse atrial remodeling leading to AF, but also inhibits its progression from paroxysmal to persistent AF. Both Mandal et al. (2005) and Rammos et al. (2002) studied HSP70 expression levels in atrial tissues of patients in sinus rhythm undergoing cardiac surgery. They consistently showed a lower incidence of POAF in patients with higher atrial intracellular HSP70 expression levels, but not serum HSP70. HSP70 has protective abilities only when it localizes intracellularly and loses its protective role when it is released into the blood (Oc et al. 2008). Higher serum HSP70 levels

may be correlated with worsened clinical outcome in various critical conditions (Ganter et al. 2006). Extracellular HSC70 induces myocardial expression of pro-inflammatory cytokines, such as TNF- $\alpha$ , IL-1 $\beta$ , IL-6, MCP-1 and keratinocyte-derived chemokine (KC, a human IL-8 analog) (Ao et al. 2009; Zou et al. 2008), which may lead to the development of POAF in patients undergoing cardiac surgery. The extracellular molecule can lead to inflammatory cytokine production and has a pro-inflammatory effect in the myocardial innate immune response caused by ischemia-reperfusion, and these effects are interacted with multiple transmembrane immune receptors, such as Toll-like receptor (TLR) 2, TLR4, lectin-like oxidized low-density lipoprotein receptor-1 (LOX-1), CD91, CD94, CD40 and CC-chemokine receptor 5 (CCR5) (de Jong et al. 2009). The mechanisms of intracellular HSP70 preventing POAF may be due to its preservation of myocardial tissue and cells from stress and ischemia-reperfusion injury.

### 4. Intracellular HSP70 anti-oxidative stress

Previous studies strongly suggested a link between myocardial oxidative stress and AF (Li et al. 2010; Neuman et al. 2007). Products of oxidative stress increased in the plasma and myocardium of patients undergoing cardiac surgery, and before and for 5 days following surgery, while supplementing patients with ascorbate, a known antioxidant, could reduce the incidence of POAF more than 2-fold (Carnes et al. 2001). Right atrial appendages of patients with AF express higher levels of the oxidative markers 3-nitrotyrosine and protein carbonyls compared to the patients with sinus rhythm (Mihm et al. 2001). Left atria of patients with AF exhibit up-regulation of Ras related C3 botulinum toxin substrate-1 (Rac-1) correlating with increased NADPH oxidase activity (Adam et al. 2007). NADPH oxidases are major sources of ROS in myocytes and vascular cells, and involved in hypertension and inflammation states such as atherosclerosis (Griendling et al. 2000), and heart failure (Heymes et al. 2003). Atrial NADPH oxidase activity was shown to be independently associated with an increased risk of AF in patients undergoing conventional CABG (Kim et al. 2008; Oral 2008). ROS produced by NADPH oxidases can promote ROS generation by other sources thereby amplifying total levels of ROS (Murdoch et al. 2006). ROS-mediated activation of p38MAPK which in turn activates Akt, results in over-expression of HSP70 in heat-stressed V79 fibroblasts (Banerjee Mustafi et al. 2009). Previous studies showed that HSP70 inhibits NADPH oxidase activity by regulating superoxide production (Maridonneau-Parini et al. 1988; Polla et al. 1995). HSP70 over-expressing astrocyte cultures down-regulated matrix metalloproteinase-9 after oxygen glucose deprivation, compared to wild-type cell cultures (Lee et al. 2004); in the meanwhile, via activation of the janus tyrosine kinase (JAK)/STAT pathway that may help vascular smooth muscle cells adapt to oxidative stress (Gaudino et al. 2003).

### 5. Intracellular HSP70 has anti-inflammatory effect

#### 5.1. Intracellular HSP70 regulates inflammatory response

In recent years, mechanisms of POAF have been proposed. Increased IL-6 and CRP levels in patients with AF suggest a role of inflammation in the pathogenesis and recurrence of AF (Hatzinikolaou-Kotsakou et al. 2006; Madamanchi et al. 2001; Ucar et al. 2007). Interestingly, there is also increasing evidence to support an association between inflammatory response and POAF after cardiac surgery (Amar et al. 2006; Fontes et al. 2009). Pro-inflammatory cytokines, such as TNF- $\alpha$ ,

IL-1, IL-6, and IL-8, play a dominant role in the upstream of the inflammatory cascade. TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 contributed to myocardial injury after ischemia-reperfusion. Systemic inflammatory response after on-pump CABG is involved in the pathogenesis of POAF (Ishida et al. 2006). Long half-life for monocytes in circulation and their ability to transform into long-lived tissue macrophages may also participate in the inflammatory response leading to POAF. In addition, monocyte CD11b up-regulation perioperatively was significantly associated with POAF (Fontes et al. 2005). The molecular is the  $\beta_2$ -integrin that mediates leukocyte adhesion to vascular endothelial cells and leukocyte migration from the vasculature into tissues (Smith et al. 1989).

Acting as a molecular chaperone in cardiac tissues, intracellular HSP70 is also associated with the orchestration of inflammatory responses after cardiac surgery and plays a role in modulating inflammation caused by ischemia-reperfusion. It was demonstrated that preoperative induction of HSP70 may attenuate CPB-induced inflammatory response by regulating NO synthase (NOS) activity (Hayashi et al. 2002). Intracellular HSP70 can inhibit responses to inflammatory cytokines, such as TNF- $\alpha$ , IL-1, IL-6 and IL-10. The experiment of Van Molle et al. showed that heat shock treatment coincides with a strong induction of HSP70, which protects against TNF-induced toxicity in the whole-body mice, yet mice missing the HSP70.1 gene are no longer protected (Van Molle et al. 2002). Liposomally delivered HSP70 protected against IL-1 $\beta$ -induced impaired pancreatic beta-cell function in a rat's diabetes model (Margulis et al. 1991). In addition to regulating the response to inflammatory cytokines, intracellular HSP70 also down-modulates their production. Inflammatory cytokines such as TNF- $\alpha$  and IL-1, are potent, multifunctional cytokine mediators of inflammation and immune responses that are produced primarily by activated monocytes and macrophages. LPS-induced increases in the production of TNF- $\alpha$ , IL-1 $\beta$ , IL-10 and IL-12 are significantly inhibited by the over-expression of HSP70 in human peripheral blood monocyte-derived macrophages (Ding et al. 2001). The study of Su et al. (2010) suggested that HSC70 preserves cardiac function involves the suppression of TLR4 signaling and production of intercellular adhesion molecule-1 (ICAM-1), which has a central role in endotoxemic myocardial dysfunction.

### 5.2. Intracellular HSP70 disrupts NF- $\kappa$ B signaling pathway

Activation of the NF- $\kappa$ B pathway acts as a functional switch and is critical to the initiation of inflammatory responses by innate cells. Intracellular HSP70 anti-inflammatory activity could account for ischemia-reperfusion injury, for its function of regulating the NF- $\kappa$ B signaling pathway (Voegeli et al. 2008). This may be due to direct interaction of HSP70 with NF- $\kappa$ B proteins, or due to interactions with other proteins in the NF- $\kappa$ B regulatory pathway (Fig. 2). Three NF- $\kappa$ B/Rel family members p50, c-Rel, p50 have been shown to co-precipitate with HSP70 (Guzhova et al. 1997). The p50/p65 heterodimer that is most well studied, is normally sequestered in the cytoplasm by its interaction with inhibitor of  $\kappa$ B (I- $\kappa$ B). Phosphorylation of I- $\kappa$ B by the I- $\kappa$ B kinase (IKK) leads to ubiquitination and degradation of I- $\kappa$ B (Gilmore 2006; Matthews and Hay 1995; Tang et al. 2007), which ultimately results in the translocation active NF- $\kappa$ B p50/p65 heterodimers from the cytosol to the nucleus (de Jong et al. 2009). The p65 subunits bind to promoter regions on the chromosome, where it induces expression of a multitude of pro-inflammatory cytokines and enzymes, such as TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and inducible NOS (iNOS) (de Jong et al. 2009). Previous studies suggested that anti-inflammatory

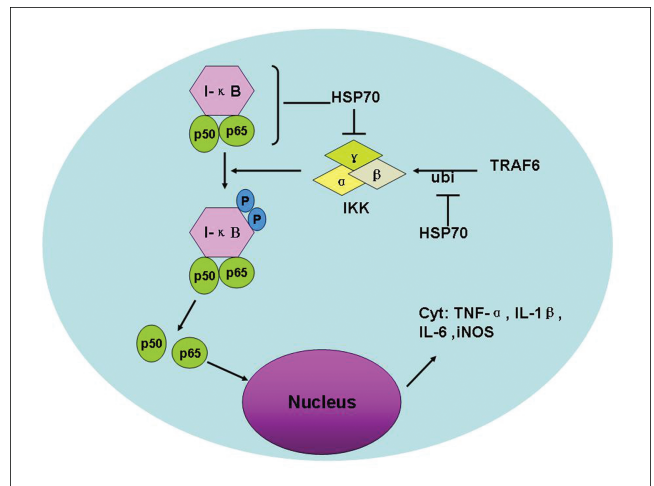


Fig. 2: Intracellular HSP70 disrupts the activation of the transcription factor NF- $\kappa$ B reducing inflammatory cytokine production. It can inhibit the activation of IKK and the ubiquitination of TRAF6, keep the stabilization of the inhibitory complex with I- $\kappa$ B. NF- $\kappa$ B = nuclear factor Kappa B; I- $\kappa$ B = inhibitor of NF- $\kappa$ B; TNF = tumor necrosis factor- $\alpha$ ; TRAF6 = TNF associated factor 6; ubi = ubiquitination; IKK = I- $\kappa$ B kinase; IKK complex is composed of three subunits including IKK $\alpha$ , IKK $\beta$  and IKK $\gamma$ . IKK $\gamma$  is an essential regulatory component of the IKK complex that is necessary for NF- $\kappa$ B activation. p50 and p65 are two of the NF- $\kappa$ B subunits which after release move to the nucleus to act as a transcription factor resulting in activation of inflammatory genes. Cyt = cytokines; IL-1 $\beta$  = interleukin-1 $\beta$ ; IL-6 = interleukin-6; iNOS = inducible nitric oxide synthase

effects of intracellular HSP70 may also involve in preserving I- $\kappa$ B complex by interacting with IKK, thereby inhibiting the degradation of I- $\kappa$ B and the subsequent activation of the NF- $\kappa$ B pathway (Chen et al. 2004a; Chen and Currie 2006; Chen et al. 2004b). IKK is composed of two catalytic subunits including IKK $\alpha$  and IKK $\beta$ , which could phosphorylate I- $\kappa$ B $\alpha$  and cause its rapid degradation by the ubiquitin-proteasome pathway and thereby mediates NF- $\kappa$ B activation. IKK $\gamma$  is an essential regulatory component of the IKK complex that is necessary for NF- $\kappa$ B activation. It was reported that the ability of HSP70 to promote TNF-mediated apoptosis was attributed to binding with IKK $\gamma$  and impairing NF- $\kappa$ B survival signaling (Ran et al. 2004). In addition, NF- $\kappa$ B activation induced by various stimuli is mediated by members of the TNF receptor-associated factor (TRAF) adapter family. TRAF6 is essential for the activation of NF- $\kappa$ B (Kobayashi et al. 2001). Intracellular HSP70 was demonstrated to inhibit LPS-induced NF- $\kappa$ B activation by binding TRAF6 via the TRAF-C domain and preventing its ubiquitination, thus resulting in inhibition of inflammatory mediator production (Cao et al. 1996). The ubiquitination of TRAF6 is a key step in the activation of the NF- $\kappa$ B pathway via activating downstream kinase IKK (Baud et al. 1999; Cao et al. 1996; Kobayashi et al. 2001). TRAF6 may also play an important role in TLR signaling, which can activate NF- $\kappa$ B by inducing I- $\kappa$ B kinase activity and degradation of I- $\kappa$ B $\alpha$  (Lee and Kim 2007). It appears likely that intracellular HSP70 involves in many levels of the NF- $\kappa$ B pathway to regulate its activation. Apparently independent of its effects on inflammatory responses, the NF- $\kappa$ B pathway has been proved to be involved in cell survival. But the mechanism of intracellular HSP70 modulation on NF- $\kappa$ B activity is still in its infancy and needs further research.

### 6. Intracellular HSP70 inhibits cell death signaling pathways

Ischemia-reperfusion injury during cardiac surgery may lead to cells death (apoptosis/necrosis) or fibrosis in the myocyte, which could associate with the development of POAF. Several

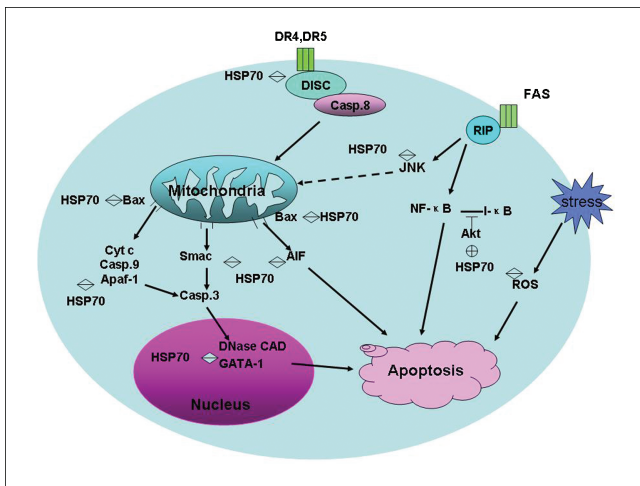


Fig. 3: HSP70 is an important regulator of apoptosis by inhibiting the mitochondrial-dependent pathway at different levels. At the extrinsic cell death receptors level, HSP70 binds to DR4 and DR5, thereby inhibiting TNF-related apoptosis-inducing ligand (TRAIL)-induced assembly and activity of death inducing signaling complex (DISC). At the pre-mitochondrial level, HSP70 inhibits stress-activated kinases. At the mitochondrial stage, HSP70 prevents mitochondrial membrane permeabilization through the blockage of Bax translocation. At the post-mitochondrial level, HSP70 interacts with AIF and Apaf-1 or protects essential nuclear proteins from caspase-3 cleavage. FAS, DR4 and DR5 all belong to the transmembrane receptors of TNF family, which can activate caspase-8 and can proceed independently of the intrinsic pathway, and they also can lead to activation of the intrinsic pathway. JNK is implicated in apoptosis triggered by FAS. Cyt c = cytochrome c; casp = caspase; AIF = apoptosis inducing factor; RIP = receptor interacting protein; JNK = Jun N terminal Kinase; Akt = PKB = protein kinase B; Apaf-1 = apoptosis protease activating factor 1; DNase CAD = caspase activated DNase; ROS = reactive oxygen species; Smac/DIABLO is a mitochondrial protein that potentiates some forms of apoptosis; Bax is a member of Bcl-2 family that acts as an anti-apoptotic protein; GATA-1 is a member of the GATA transcription factor family, it plays a role in erythroid development by regulating the switch of fetal hemoglobin to adult hemoglobin

studies have shown that intracellular HSP70 may directly interfere with cell death pathways (Lanneau et al. 2008; Mokhtari et al. 2009; Zhao et al. 2007), which are described for mammals as the extrinsic and the intrinsic pathways. The extrinsic or death receptor-mediated pathway plays a major role in, for example, the immune system, while the intrinsic pathway, also called the mitochondrial-dependent pathway, is activated in response to extracellular or intracellular injury such as DNA damage. Mitochondria are known to have a great impact on the regulation of apoptosis; it relies on the release of mitochondrial pro-apoptotic molecules, opening of the mitochondrial permeability transition pore, and activation of caspases (Gogvadze and Orrenius 2006). Intracellular HSP70 acts as an important regulator of apoptosis, by inhibiting the apoptotic pathways at the extrinsic receptor pathway and at different levels of the mitochondrial-dependent pathway (Fig. 3).

### 6.1. HSP70 targets at the extrinsic cell death receptor pathway

The extrinsic cell death pathway is initiated upon ligand-receptor interactions at the cell surface including FAS ligand-FAS/APO1 (CD95), TNF-TNF receptors, and TNF-related apoptosis-inducing ligand (TRAIL)-TRAIL receptors (Ozoren and El-Deiry 2003). They can activate caspase-8 and can proceed independently of the intrinsic pathway, and also can lead to activation of the intrinsic pathway (Thorburn 2004). Overall eight extrinsic death receptors have been identified and all belong to the receptors of TNF family, such as TNF-R1, FAS, DR3, TRAIL-R1 (DR4), TRAIL-R2 (KILLER/DR5) and

DR6 (Locksley et al. 2001). TRAIL-death receptors can activate both the extrinsic and intrinsic death signaling cascades (Ozoren and El-Deiry 2003). At the extrinsic cell death receptors level, HSP70 binds to DR4 and DR5, thereby inhibiting TRAIL-induced assembly and activity of death inducing signaling complex (DISC) (Guo et al. 2005).

### 6.2. HSP70 targets at the pre-mitochondrial level

At the pre-mitochondrial level, intracellular HSP70 blocks the activation of c-Jun N-terminal kinase (JNK), which plays a critical role in mitochondrial intrinsic apoptotic pathways by directly modulating the activities of mitochondrial pro- and anti-apoptotic proteins through distinct phosphorylation events (Dhanasekaran and Reddy 2008). HSP70 inhibits JNK dephosphorylation thereby blocking its activation (Lee et al. 2005; Park et al. 2001; Yaglom et al. 1999). The ATPase domain of HSP70 is dispensable for this binding. The deficiency of HSP70 induces JNK and caspase-3 activation in hyperosmolarity-induced apoptosis (Lee et al. 2005). In the meantime, JNK is implicated in apoptosis triggered by FAS (Aggarwal 2003). HSP70 is capable of protecting the cardiomyocyte from stress-induced injury by inhibiting FAS-mediated apoptosis (Zhao et al. 2007). In addition, HSP70 binds to nonphosphorylated protein kinase C (PKC) and Akt, stabilizing both proteins and allowing rephosphorylation of PKC (Gao and Newton 2002), both of which are well known for protecting from damage-induced cells death. It has been discussed above that HSP70 acts as a regulator for the important anti-apoptotic prosurvival kinase Akt/PKB (Barati et al. 2006; Rafiee et al. 2006), which is involved in inhibiting the activation of NF- $\kappa$ B signaling pathway in the upstream.

### 6.3. HSP70 targets at the mitochondrial level

At the mitochondrial level, intracellular HSP70 primarily helps to maintain mitochondrial membrane potential. This effect may contribute to the preservation of mitochondrial function and mitochondrial protein import, by preventing mitochondrial membrane permeabilization through the blockage of Bax translocation. Bax belongs to the Bcl-2 family, which comprise both anti-apoptotic members, for example, Bcl-2, Bcl-XL, and Mcl-1, and pro-apoptotic molecules, such as Bax, Bak, and BH3 domain only molecules (Adams and Cory 2007). The balances between pro- and anti-apoptotic members of the Bcl-2 family determine whether cells undergo apoptosis by interacting with intrinsic mitochondrial proteins or regulating the mitochondrial membrane permeability transition. Bcl-2 acts as a key anti-apoptotic protein by its increased expression blocks the release of cytochrome c and apoptosis inducing factor (AIF), and reduces caspase activation. Cytochrome c translocates from the mitochondria to the cytosol, thereby interacts with the CED-4 homologue, apoptosis protease activating factor-1 (Apaf-1), and dATP, to form the apoptosome and activate caspase-9 (Gogvadze and Orrenius 2006; Leist and Jaattela 2001). Caspase-9 activates caspase-3, thereby activates caspase-activated DNase. With cleavage of multiple targets within the cell and DNA fragmentation, apoptotic cell death results. HSP70 can inhibit Bax translocation and insertion into the outer mitochondrial membrane, thereby preventing mitochondrial membrane permeabilization and release of cytochrome c and AIF (Stankiewicz et al. 2005).

### 6.4. HSP70 targets at the post-mitochondrial level

At the post-mitochondrial level, intracellular HSP70 interacts with AIF and Apaf-1 or protects essential nuclear proteins from caspase-3 cleavage. It interferes with the activity of AIF,

inhibits AIF nuclear translocation and chromatin condensation; the interaction involves a domain of AIF between aminoacids 150 and 228 (Gurbuxani et al. 2003). Furthermore, HSP70 has been shown to interfere with recruitment of procaspase-9 into the apoptosome (Beere et al. 2000; Saleh et al. 2000), and to associate with EndoG and to prevent DNA fragmentation (Kalinowska et al. 2005). Since EndoG can form complexes with AIF, its association with HSP70 could involve AIF as a molecular bridge. It also inhibits release of the pro-apoptotic protein Smac/DIABLO from myocyte mitochondria and apoptosis (Jiang et al. 2005). HSP70 also binds directly to Apaf-1, thereby inhibiting the recruitment of procaspase-9 to the apoptosome and preventing activation of the caspase-dependent apoptosis cascade (Beere et al. 2000; Saleh et al. 2000). In addition, HSP70 rescues cells from a later phase of apoptosis than any known survival protein, downstream caspase-3 activation (Jaattela et al. 1998). During the final phases of apoptosis, chromosomal DNA is digested by the DNase CAD (caspase activated DNase), following activation by caspase-3. The enzymatic activity and proper folding of CAD has been reported to be regulated by HSP70 (Sakahira and Nagata 2002). Another final target of caspase-3 is the transcription factor GATA-1. HSP70 can protect the transcription factor from caspase-3 cleavage in the nucleus (Ribeil et al. 2007).

A growing number of data show now that ROS, mainly produced by the mitochondria (Ali et al. 2010; Criscuolo et al. 2010; Pouvreau 2010), can also be involved in cell death. On the one hand, they can produce oxidative stress leading to cell destruction, as observed during necrosis or the post-mitochondrial phase of apoptosis. On the other hand, ROS derived from mitochondria are also involved in the initiation phase of apoptosis contributing to cell death signaling. Over-expression of HSP72 was shown to enhance the activity of the mitochondrial antioxidant enzyme manganese superoxide dismutase in myocardial cells (Suzuki et al. 2002). HSP70 inhibits NADPH oxidase activity by regulating superoxide production (Maridonneau-Parini et al. 1988; Polla et al. 1995).

## 7. Intracellular HSP70 attenuates Ca<sup>2+</sup> overload

Under normal conditions, Ca<sup>2+</sup> enters cardiomyocyte primarily through I<sub>Ca,L</sub>. Ca<sup>2+</sup> can also enter via the reverse-mode Na<sup>+</sup>/Ca<sup>2+</sup> exchanger (NCX). This Ca<sup>2+</sup> influx triggers much larger Ca<sup>2+</sup> release via ryanodine-receptor channels (RyR2) in the sarcoplasmic reticulum through Ca<sup>2+</sup>-induced Ca<sup>2+</sup> release. Abnormalities in intracellular Ca<sup>2+</sup> handling may constitute key missing links in AF-initiating focal activity and AF perpetuation by rapidly firing foci and reentry. AF is associated with phosphorylation of phospholamban and RyR2, which might endorse cellular Ca<sup>2+</sup> overload (El-Armouche et al. 2006; Vest et al. 2005). Atrial tachycardia promotes the induction and maintenance of AF. When AF persists, tissue adaptive responses in the form of structural remodeling will occur. Previous experiments suggested that pre-induction of HSPs by a mild non-toxic heat shock or by the drug geranylgeranylacetone (GGA) prevented atrial remodeling processes induced by tachypacing (Brundel et al. 2006a, b). Activation of intracellular HSP70 attenuated Ca<sup>2+</sup> overload by increasing the release of Ca<sup>2+</sup> via RyR2, sped up the uptake of Ca<sup>2+</sup> via sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase, and removal of Ca<sup>2+</sup> via NCX in the rat cardiomyocytes (Liu et al. 2006). But Brundel et al.'s study suggested that HSP27, not HSP70 induction by GGA protected HL-1 myocytes against suppression of cellular Ca<sup>2+</sup> release and contractility resulting from tachypacing (Brundel et al. 2006b). The precise mechanism of intracellular HSP70 attenuates Ca<sup>2+</sup> overload is still unclear and needs further research.

## 8. Summary

POAF is associated with significant morbidity and increased health care costs. It is a probable consequence of the electrophysiological disturbances that are mainly associated with reperfusion injury and inflammation in patients undergoing cardiac surgery patients. HSP70 is a group of proteins which facilitate the folding of newly, synthesized polypeptides in an ATP dependent manner and play an important role in maintaining the dynamic stability of the intracellular proteome. Besides its function as a molecular chaperone in cardiac tissues, the intracellular molecule is also involved in the orchestration of inflammatory responses after myocardial ischemia reperfusion and can inhibit the pathogenesis of POAF. HSPs could be induced by a mild non-toxic heat shock and the drug GGA. And there is an experiment support that oral GGA is cardioprotective against ischemia reperfusion injury insult through its induction of HSP72 (Ooie et al. 2001). Since intramyocardial HSP70 acts as an anti-arrhythmic and a protective function in cardiac surgery, preoperative induction of HSP70 by myocardial precondition or any drug could be a new and useful strategy for decreasing the incidence of POAF and its associated complications. But more clinical proofs are needed to support that HSP70 holds as a novel target for the prevention and treatment of POAF. In addition, for its extracellular action as pro-inflammatory, the use of plasma HSP70 as a marker in the development of POAF and its possible use in prognosis are being investigated.

## References

- Adam O, Frost G, Custodis F, Sussman MA, Schafers HJ, Bohm M, Laufs U (2007) Role of Rac1 GTPase activation in atrial fibrillation. *J Am Coll Cardiol* 50: 359–367.
- Adams J, Cory S (2007) The Bcl-2 apoptotic switch in cancer development and therapy. *Oncogene* 26: 1324–1337.
- Aggarwal B (2003) Signalling pathways of the TNF superfamily: a double-edged sword. *Nat Rev Immunol* 3: 745–756.
- Ali SS, Marcondes MC, Bajova H, Dugan LL, Conti B (2010) Metabolic depression and increased reactive oxygen species production by isolated mitochondria at moderately lower temperatures. *J Biol Chem* 285: 32522–32528.
- Amar D, Goenka A, Zhang H, Park B, Thaler HT (2006). Leukocytosis and increased risk of atrial fibrillation after general thoracic surgery. *Ann Thorac Surg* 82: 1057–1061.
- Ao L, Zou N, Cleveland JC Jr, Fullerton DA, Meng X (2009) Myocardial TLR4 is a determinant of neutrophil infiltration after global myocardial ischemia: mediating KC and MCP-1 expression induced by extracellular HSC70. *Am J Physiol Heart Circ Physiol* 297: H21–28.
- Aranki SF, Shaw DP, Adams DH, Rizzo RJ, Couper GS, VanderVliet M, Collins JJ Jr, Cohn LH, Burstin HR (1996) Predictors of atrial fibrillation after coronary artery surgery. Current trends and impact on hospital resources. *Circulation* 94: 390–397.
- Ascione R, Caputo M, Calori G, Lloyd CT, Underwood MJ, Angelini GD (2000) Predictors of atrial fibrillation after conventional and beating heart coronary surgery: A prospective, randomized study. *Circulation* 102: 1530–1535.
- Banerjee Mustafi S, Chakraborty PK, Dey RS, Raha S (2009) Heat stress upregulates chaperone heat shock protein 70 and antioxidant manganese superoxide dismutase through reactive oxygen species (ROS), p38MAPK, and Akt. *Cell Stress and Chaperones* 14: 579–589.
- Barati MT, Rane MJ, Klein JB, McLeish KR (2006) A proteomic screen identified stress-induced chaperone proteins as targets of Akt phosphorylation in mesangial cells. *J Proteome Res* 5: 1636–1646.
- Baud V, Liu ZG, Bennett B, Suzuki N, Xia Y, Karin M (1999). Signaling by proinflammatory cytokines: oligomerization of TRAF2 and TRAF6 is sufficient for JNK and IKK activation and target gene induction via an amino-terminal effector domain. *Genes Dev* 13: 1297–1308.
- Beere HM, Wolf BB, Cain K, Mosser DD, Mahboubi A, Kuwana T, Tailor P, Morimoto RI, Cohen GM, Green DR (2000) Heat-shock protein 70 inhibits apoptosis by preventing recruitment of procaspase-9 to the Apaf-1 apoptosome. *Nat Cell Biol* 2: 469–475.

- Boos CJ, Anderson RA, Lip GY (2006) Is atrial fibrillation an inflammatory disorder? *Eur Heart J* 27: 136–149.
- Bowie A, O'Neill LA (2000) Oxidative stress and nuclear factor-kappaB activation: a reassessment of the evidence in the light of recent discoveries. *Biochem Pharmacol* 59: 13–23.
- Bruins P, te Velthuis H, Yazdanbakhsh AP, Jansen PG, van Hardevelt FW, de Beaumont EM, Wildevuort CR, Eijssman L, Trouwborst A, Hack CE (1997) Activation of the complement system during and after cardiopulmonary bypass surgery: postsurgery activation involves C-reactive protein and is associated with postoperative arrhythmia. *Circulation* 96: 3542–3548.
- Brundel BJ, Henning RH, Ke L, van Gelder IC, Crijns HJ, Kampinga HH (2006a) Heat shock protein upregulation protects against pacing-induced myolysis in HL-1 atrial myocytes and in human atrial fibrillation. *J Mol Cell Cardiol* 41: 555–562.
- Brundel BJ, Shiroshita-Takeshita A, Qi X, Yeh YH, Chartier D, van Gelder IC, Henning RH, Kampinga HH, Nattel S (2006b) Induction of heat shock response protects the heart against atrial fibrillation. *Circ Res* 99: 1394–1402.
- Cao Z, Xiong J, Takeuchi M, Kurama T, Goeddel DV (1996) TRAF6 is a signal transducer for interleukin-1. *Nature* 383: 443–446.
- Carnes CA, Chung MK, Nakayama T, Nakayama H, Baliga RS, Piao S, Kanderian A, Pavia S, Hamlin RL, McCarthy PM, Bauer JA, Van Wagener DR (2001) Ascorbate attenuates atrial pacing-induced peroxynitrite formation and electrical remodeling and decreases the incidence of postoperative atrial fibrillation. *Circ Res* 89: E32–38.
- Chen Y, Arrigo AP, Currie RW (2004a) Heat shock treatment suppresses angiotensin II-induced activation of NF-kappaB pathway and heart inflammation: a role for IKK depletion by heat shock? *Am J Physiol Heart Circ Physiol* 287: H1104–1114.
- Chen Y, Currie RW (2006) Small interfering RNA knocks down heat shock factor-1 (HSF-1) and exacerbates pro-inflammatory activation of NF-kappaB and AP-1 in vascular smooth muscle cells. *Cardiovasc Res* 69: 66–75.
- Chen Y, Ross BM, Currie RW (2004b) Heat shock treatment protects against angiotensin II-induced hypertension and inflammation in aorta. *Cell Stress Chaperones* 9: 99–107.
- Criscuolo F, Font-Sala C, Bouillaud F, Poulin N, Trabalon M (2010) Increased ROS production: a component of the longevity equation in the male mygalomorph, *Brachypelma albopilosa*. *PLoS One* 5.
- de Jong PR, Schadenberg AW, Jansen NJ, Prakken BJ (2009) Hsp70 and cardiac surgery: molecular chaperone and inflammatory regulator with compartmentalized effects. *Cell Stress Chaperones* 14: 117–131.
- Dhanasekaran DN, Reddy EP (2008) JNK signaling in apoptosis. *Oncogene* 27: 6245–6251.
- Ding XZ, Fernandez-Prada CM, Bhattacharjee AK, Hoover DL (2001) Over-expression of hsp-70 inhibits bacterial lipopolysaccharide-induced production of cytokines in human monocyte-derived macrophages. *Cytokine* 16: 210–219.
- Dupont E, Ko Y, Rothery S, Coppen SR, Baghai M, Haw M, Severs NJ (2001) The gap-junctional protein connexin40 is elevated in patients susceptible to postoperative atrial fibrillation. *Circulation* 103: 842–849.
- Dybdahl B, Wahba A, Haaverstad R, Kirkeby-Garstad I, Kierulf P, Espevik T, Sundan A (2004) On-pump versus off-pump coronary artery bypass grafting: more heat-shock protein 70 is released after on-pump surgery. *European Journal of Cardio-Thoracic Surgery* 25: 985–992.
- Dybdahl B, Wahba A, Lien E, Flo TH, Waage A, Qureshi N, Sellevold OF, Espevik T, Sundan A (2002). Inflammatory response after open heart surgery: release of heat-shock protein 70 and signaling through toll-like receptor-4. *Circulation* 105: 685–690.
- El-Armouche A, Boknik P, Eschenhagen T, Carrier L, Knaut M, Ravens U, Dobrev D (2006) Molecular determinants of altered Ca<sup>2+</sup> handling in human chronic atrial fibrillation. *Circulation* 114: 670–680.
- El-Chami MF, Kilgo P, Thourani V, Lattouf OM, Delurgio DB, Guyton RA, Leon AR, Puskas JD (2010) New-onset atrial fibrillation predicts long-term mortality after coronary artery bypass graft. *J Am Coll Cardiol* 55: 1370–1376.
- Elahi M, Hadjiminikolaou L, Galinanes M (2003) Incidence and clinical consequences of atrial fibrillation within 1 year of first-time isolated coronary bypass surgery. *Circulation* 108 Suppl 1: II207–212.
- Fehrenbach E, Niess AM, Schlotz E, Passet F, Dickhuth HH, Northoff H (2000) Transcriptional and translational regulation of heat shock proteins in leukocytes of endurance runners. *J Appl Physiol* 89: 704–710.
- Fontes ML, Amar D, Kulak A, Koval K, Zhang H, Shi W, Thaler H (2009) Increased preoperative white blood cell count predicts postoperative atrial fibrillation after coronary artery bypass surgery. *J Cardiothorac Vasc Anesth* 23: 484–487.
- Fontes ML, Mathew JP, Rinder HM, Zelterman D, Smith BR, Rinder CS (2005) Atrial fibrillation after cardiac surgery/cardiopulmonary bypass is associated with monocyte activation. *Anesth Analg* 101: 17–23.
- Ganter MT, Ware LB, Howard M, Roux J, Gartland B, Matthay MA, Flesher M, Pitte, JF (2006) Extracellular heat shock protein 72 is a marker of the stress protein response in acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 291: L354–361.
- Gao T, Newton, AC (2002) The turn motif is a phosphorylation switch that regulates the binding of Hsp70 to protein kinase C. *J Biol Chem* 277: 31585–31592.
- Gaudio M, Andreotti F, Zamparelli R, Di Castelnuovo A, Nasso G, Burzotta F, Iacoviello L, Donati MB, Schiavello R, Maseri A, Possati G (2003). The –174G/C interleukin-6 polymorphism influences postoperative interleukin-6 levels and postoperative atrial fibrillation. Is atrial fibrillation an inflammatory complication? *Circulation* 108 Suppl 1: II195–199.
- Gilmore TD (2006) Introduction to NF-kappaB: players, pathways, perspectives. *Oncogene* 25: 6680–6684.
- Girerd N, Pibarot P, Fournier D, Daleau P, Voisine P, O'Hara G, Despres JP, Mathieu P (2009) Middle-aged men with increased waist circumference and elevated C-reactive protein level are at higher risk for postoperative atrial fibrillation following coronary artery bypass grafting surgery. *Eur Heart J* 30: 1270–1278.
- Gogvadze V, Orrenius S (2006) Mitochondrial regulation of apoptotic cell death. *Chem Biol Interact* 163: 4–14.
- Griendling KK, Sorescu D, Ushio-Fukai M (2000) NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ Res* 86: 494–501.
- Guo F, Sigua C, Bali P, George P, Fiskus W, Scuto A, Annavarapu S, Mouttaki A, Sondarva G, Wei S, Wu J, Djeu J, Bhalla K (2005). Mechanistic role of heat shock protein 70 in Bcr-Abl-mediated resistance to apoptosis in human acute leukemia cells. *Blood* 105: 1246–1255.
- Gurbuxani S, Schmitt E, Cande C, Parcellier A, Hammann A, Daugas E, Kouranti I, Spahr C, Pance A, Kroemer G, Garrido C (2003) Heat shock protein 70 binding inhibits the nuclear import of apoptosis-inducing factor. *Oncogene* 22: 6669–6678.
- Guzhova IV, Darieva ZA, Melo AR, Margulis BA (1997) Major stress protein Hsp70 interacts with NF-kB regulatory complex in human T-lymphoma cells. *Cell Stress Chaperones* 2: 132–139.
- Hatzinikolaou-Kotsakou E, Tziakas D, Hotidis A, Stakos D, Floros D, Papanas N, Chalikias G, Maltezos E, Hatseras DI (2006) Relation of C-reactive protein to the first onset and the recurrence rate in lone atrial fibrillation. *Am J Cardiol* 97: 659–661.
- Hayashi Y, Sawa Y, Fukuyama N, Nakazawa H, Matsuda H (2002) Preoperative glutamine administration induces heat-shock protein 70 expression and attenuates cardiopulmonary bypass-induced inflammatory response by regulating nitric oxide synthase activity. *Circulation* 106: 2601–2607.
- Heymes C, Bendall JK, Ratajczak P, Cave AC, Samuel JL, Hasenfuss G, Shah AM (2003) Increased myocardial NADPH oxidase activity in human heart failure. *J Am Coll Cardiol* 41: 2164–2171.
- Hilfiker-Kleiner D, Hilfiker A, Drexler H (2005). Many good reasons to have STAT3 in the heart. *Pharmacol Therap* 107: 131–137.
- Ishida K, Kimura F, Imamaki M, Ishida A, Shimura H, Kohno H, Sakurai M, Miyazaki M (2006) Relation of inflammatory cytokines to atrial fibrillation after off-pump coronary artery bypass grafting. *Eur J Cardiothorac Surg* 29: 501–505.
- Ishii Y, Schuessler RB, Gaynor SL, Yamada K, Fu AS, Boineau JP, Damiano RJ Jr (2005) Inflammation of atrium after cardiac surgery is associated with inhomogeneity of atrial conduction and atrial fibrillation. *Circulation* 111: 2881–2888.
- Jaattela M, Wissing D, Kokholm K, Kallunki T, Egeblad M (1998) Hsp70 exerts its anti-apoptotic function downstream of caspase-3-like proteases. *EMBO J* 17: 6124–6134.
- Jiang B, Xiao W, Shi Y, Liu M, Xiao X (2005) Heat shock pretreatment inhibited the release of Smac/DIABLO from mitochondria and apoptosis induced by hydrogen peroxide in cardiomyocytes and C2C12 myogenic cells. *Cell Stress Chaperones* 10: 252–262.
- Kalinowska M, Garncarz W, Pietrowska M, Garrard WT, Widlak P (2005) Regulation of the human apoptotic DNase/RNase endonuclease G: involvement of Hsp70 and ATP. *Apoptosis* 10: 821–830.
- Kalus JS, Caron MF, White CM, Mather JF, Gallagher R, Boden WE, Kluger J (2004) Impact of fluid balance on incidence of atrial fibrillation after cardiothoracic surgery. *Am J Cardiol* 94: 1423–1425.
- Kim YM, Kattach H, Ratnatunga C, Pillai R, Channon KM, Casadei B (2008) Association of atrial nicotinamide adenine dinucleotide phosphate oxidase activity with the development of atrial fibrillation after cardiac surgery. *J Am Coll Cardiol* 51: 68–74.

- Knowlton AA, Eberli FR, Brecher P, Romo GM, Owen A, Apstein CS (1991) A single myocardial stretch or decreased systolic fiber shortening stimulates the expression of heat shock protein 70 in the isolated, erythrocyte-perfused rabbit heart. *J Clin Invest* 88: 2018–2025.
- Kobayashi N, Kadono Y, Naito A, Matsumoto K, Yamamoto T, Tanaka S, Inoue J (2001) Segregation of TRAF6-mediated signaling pathways clarifies its role in osteoclastogenesis. *EMBO J* 20: 1271–1280.
- Lanneau D, Brunet M, Frisan E, Solary E, Fontenay M, Garrido C (2008) Heat shock proteins: essential proteins for apoptosis regulation. *J Cell Mol Med* 12: 743–761.
- Lee JE, Kim YJ, Kim JY, Lee WT, Yenari MA, Giffard RG (2004) The 70 kDa heat shock protein suppresses matrix metalloproteinases in astrocytes. *Neuroreport* 15: 499–502.
- Lee JS, Lee JJ, Seo JS (2005) HSP70 deficiency results in activation of c-Jun N-terminal Kinase, extracellular signal-regulated kinase, and caspase-3 in hyperosmolarity-induced apoptosis. *J Biol Chem* 280: 6634–6641.
- Lee MS, Kim YJ (2007) Signaling pathways downstream of pattern-recognition receptors and their cross talk. *Annu Rev Biochem* 76: 447–480.
- Leist M, Jaattela M (2001) Four deaths and a funeral: from caspases to alternative mechanisms. *Nat Rev Mol Cell Biol* 2: 589–598.
- Li J, Solus J, Chen Q, Rho YH, Milne G, Stein CM, Darbar D (2010) Role of inflammation and oxidative stress in atrial fibrillation. *Heart Rhythm* 7: 438–444.
- Lin CY, Yang TL, Hong GJ, Li CY, Lin FY, Tsai CS (2010) Enhanced intracellular heat shock protein 70 expression of leukocytes and serum interleukins release: comparison of on-pump and off-pump coronary artery surgery. *World J Surg* 34: 675–681.
- Liu J, Kam KW, Borchert GH, Kravtsov GM, Ballard HJ, Wong TM (2006) Further study on the role of HSP70 on Ca<sup>2+</sup> homeostasis in rat ventricular myocytes subjected to simulated ischemia. *Am J Physiol Cell Physiol* 290: C583–591.
- Locksley RM, Killeen N, Lenardo MJ (2001) The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell* 104: 487–501.
- Madamanchi NR, Li S, Patterson C, Runge MS (2001) Reactive oxygen species regulate heat-shock protein 70 via the JAK/STAT pathway. *Arterioscler Thromb Vasc Biol* 21: 321–326.
- Magee MJ, Herbert MA, Dewey TM, Edgerton JR, Ryan WH, Prince S, Mack MJ (2007) Atrial fibrillation after coronary artery bypass grafting surgery: development of a predictive risk algorithm. *Ann Thorac Surg* 83: 1707–1712; discussion 1712.
- Mandal K, Torsney E, Poloniecki J, Camm AJ, Xu Q, Jahangiri M (2005) Association of high intracellular, but not serum, heat shock protein 70 with postoperative atrial fibrillation. *Ann Thorac Surg* 79: 865–871.
- Margulis BA, Sandler S, Eizirik DL, Welsh N, Welsh M (1991) Liposomal delivery of purified heat shock protein hsp70 into rat pancreatic islets as protection against interleukin 1 beta-induced impaired beta-cell function. *Diabetes* 40: 1418–1422.
- Maridonneau-Parini I, Clerc J, Polla BS (1988) Heat shock inhibits NADPH oxidase in human neutrophils. *Biochem Biophys Res Commun* 154: 179–186.
- Matata BM, Sosnowski AW, Galinanes M (2000) Off-pump bypass graft operation significantly reduces oxidative stress and inflammation. *Ann Thorac Surg* 69: 785–791.
- Mathew JP, Fontes ML, Tudor IC, Ramsay J, Duke P, Mazer CD, Barash PG, Hsu PH, Mangano DT (2004) A multicenter risk index for atrial fibrillation after cardiac surgery. *JAMA* 291: 1720–1729.
- Matthews JR, Hay RT (1995) Regulation of the DNA binding activity of NF-kappa B. *Int J Biochem Cell Biol* 27: 865–879.
- Mihm MJ, Yu F, Carnes CA, Reiser PJ, McCarthy PM, Van Wagoner DR, Bauer JA (2001) Impaired myofibrillar energetics and oxidative injury during human atrial fibrillation. *Circulation* 104: 174–180.
- Mokhtari D, Kerblom B, Mehmeti I, Wang X, Funa NS, Olerud J, Lenzen S, Welsh N, Welsh M (2009) Increased Hsp70 expression attenuates cytokine-induced cell death in islets of Langerhans from Shb knockout mice. *Biochem Biophys Res Commun* 387: 553–557.
- Murdoch CE, Zhang M, Cave AC, Shah AM (2006) NADPH oxidase-dependent redox signalling in cardiac hypertrophy, remodelling and failure. *Cardiovasc Res* 71: 208–215.
- Nattel S (2002) New ideas about atrial fibrillation 50 years on. *Nature* 415: 219–226.
- Nattel S, Maguy A, Le Bouter S, Yeh YH (2007) Arrhythmogenic ion-channel remodeling in the heart: heart failure, myocardial infarction, and atrial fibrillation. *Physiol Rev* 87: 425–456.
- Nazeri A, Razavi M, Elayda MA, Lee VV, Massumi A, Wilson JM (2010) Race/ethnicity and the incidence of new-onset atrial fibrillation after isolated coronary artery bypass surgery. *Heart Rhythm* 7: 1458–1463.
- Neuman RB, Bloom HL, Shukrullah I, Darrow LA, Kleinbaum D, Jones DP, Dudley SC Jr (2007) Oxidative stress markers are associated with persistent atrial fibrillation. *Clin Chem* 53: 1652–1657.
- Oc M, Ucar HI, Pinar A, Akbulut B, Oc B, Akyon Y, Kanbak M, Dogan R (2008) Heat shock protein70: a new marker for subsequent atrial fibrillation development? *Artif Organs* 32: 846–850.
- Ooie T, Takahashi N, Saikawa T, Nawata T, Arikawa M, Yamanaka K, Hara M, Shimada T, Sakata T (2001) Single oral dose of geranylgeranylacetone induces heat-shock protein 72 and renders protection against ischemia/reperfusion injury in rat heart. *Circulation* 104: 1837–1843.
- Oral H (2008) Post-operative atrial fibrillation and oxidative stress: a novel causal mechanism or another biochemical epiphenomenon? *J Am Coll Cardiol* 51: 75–76.
- Ozoren N, El-Deiry WS (2003) Cell surface Death Receptor signaling in normal and cancer cells. *Semin Cancer Biol* 13: 135–147.
- Park HS, Lee JS, Huh SH, Seo JS, Choi EJ (2001) Hsp72 functions as a natural inhibitory protein of c-Jun N-terminal kinase. *EMBO J* 20: 446–456.
- Polla BS, Stubbe H, Kantengwa S, Maridonneau-Parini I, Jacquier-Sarlin MR (1995) Differential induction of stress proteins and functional effects of heat shock in human phagocytes. *Inflammation* 19: 363–378.
- Pouvreau S (2010) Superoxide flashes in mouse skeletal muscle are produced by discrete arrays of active mitochondria operating coherently. *PLoS One* 5.
- Rafiee P, Theriot ME, Nelson VM, Heidemann J, Kanaa Y, Horowitz SA, Rogaczewski A, Johnson CP, Ali I, Shaker R, Bibion DG (2006) Human esophageal microvascular endothelial cells respond to acidic pH stress by PI3K/AKT and p38 MAPK-regulated induction of Hsp70 and Hsp27. *Am J Physiol Cell Physiol* 291: C931–945.
- Ramos KS, Koullias GJ, Hassan MO, Argyrakis NP, Voucharas CG, Scarupa SJ, Cowte TG (2002) Low preoperative HSP70 atrial myocardial levels correlate significantly with high incidence of postoperative atrial fibrillation after cardiac surgery. *Cardiovasc Surg* 10: 228–232.
- Ran R, Lu A, Zhang L, Tang Y, Zhu H, Xu H, Feng Y, Han C, Zhou G, Rigby AC, Sharp FR (2004) Hsp70 promotes TNF-mediated apoptosis by binding IKK gamma and impairing NF-kappa B survival signaling. *Genes Dev* 18: 1466–1481.
- Ribeil JA, Zermati Y, Vandekerckhove J, Cathelin S, Kersual J, Dussiot M, Coulon S, Moura IC, Zeuner A, Kirkegaard-Sorensen T, Varet B, Solary E, Garrido C, Hermine O (2007) Hsp70 regulates erythropoiesis by preventing caspase-3-mediated cleavage of GATA-1. *Nature* 445: 102–105.
- Sakahira H, Nagata S (2002) Co-translational folding of caspase-activated DNase with Hsp70, Hsp40, and inhibitor of caspase-activated DNase. *J Biol Chem* 277: 3364–3370.
- Saleh A, Srinivasula SM, Balkir L, Robbins PD, Alnemri ES (2000) Negative regulation of the Apaf-1 apoptosome by Hsp70. *Nat Cell Biol* 2: 476–483.
- Schmitt JP, Schunkert H, Birnbaum DE, Aebert H (2002) Kinetics of heat shock protein 70 synthesis in the human heart after cold cardioplegic arrest. *Eur J Cardiothorac Surg* 22: 415–420.
- Smith CW, Marlin SD, Rothlein R, Toman C, Anderson DC (1989) Cooperative interactions of LFA-1 and Mac-1 with intercellular adhesion molecule-1 in facilitating adherence and transendothelial migration of human neutrophils *in vitro*. *J Clin Invest* 83: 2008–2017.
- Stankiewicz AR, Lachapelle G, Foo CP, Radicioni SM, Mosser DD (2005) Hsp70 inhibits heat-induced apoptosis upstream of mitochondria by preventing Bax translocation. *J Biol Chem* 280: 38729–38739.
- Su X, Sykes JB, Ao L, Raeburn CD, Fullerton DA, Meng X (2010) Extracellular heat shock cognate protein 70 induces cardiac functional tolerance to endotoxin: differential effect on TNF-alpha and ICAM-1 levels in heart tissue. *Cytokine* 51: 60–66.
- Sun H, Chartier D, Leblanc N, Nattel S (2001) Intracellular calcium changes and tachycardia-induced contractile dysfunction in canine atrial myocytes. *Cardiovasc Res* 49: 751–761.
- Sun H, Gaspo R, Leblanc N, Nattel S (1998) Cellular mechanisms of atrial contractile dysfunction caused by sustained atrial tachycardia. *Circulation* 98: 719–727.
- Suzuki K, Murtuza B, Sammut IA, Latif N, Jayakumar J, Smolenski RT, Kaneda Y, Sawa Y, Matsuda H, Yacoub MH (2002) Heat shock protein 72 enhances manganese superoxide dismutase activity during myocardial ischemia-reperfusion injury, associated with mitochondrial protection and apoptosis reduction. *Circulation* 106: I270–276.
- Szerafin T, Hoetzenecker K, Hacker S, Horvath A, Polleis A, Arpad P, Mangold A, Wiszczak T, Dworschak M, Seitelberger R (2008) Heat Shock Proteins 27, 60, 70, 90 [alpha], and 20S proteasome in on-pump

- versus off-pump coronary artery bypass graft patients. *The Annals Thorac Surg* 85: 80–87.
- Tang D, Kang R, Xiao W, Wang H, Calderwood SK, Xiao X (2007) The anti-inflammatory effects of heat shock protein 72 involve inhibition of high-mobility-group box 1 release and proinflammatory function in macrophages. *J Immunol* 179: 1236–1244.
- Thorburn A (2004) Death receptor-induced cell killing. *Cell Signal* 16: 139–144.
- Ucar HI, Tok M, Atalar E, Dogan OF, Oc M, Farsak B, Guvener M, Yilmaz M, Dogan R, Demircin M, Pasaoglu I (2007) Predictive significance of plasma levels of interleukin-6 and high-sensitivity C-reactive protein in atrial fibrillation after coronary artery bypass surgery. *Heart Surg Forum* 10: E131–135.
- van der Velden HM, Ausma J, Rook MB, Hellemons AJ, van Veen TA, Allessie MA, Jongsma HJ (2000) Gap junctional remodeling in relation to stabilization of atrial fibrillation in the goat. *Cardiovasc Res* 46: 476–486.
- Van Molle W, Wielockx B, Mahieu T, Takada M, Taniguchi T, Sekikawa K, Libert C (2002) HSP70 protects against TNF-induced lethal inflammatory shock. *Immunity* 16: 685–695.
- Vest JA, Wehrens XH, Reiken SR, Lehnart SE, Dobrev D, Chandra P, Danilo P, Ravens U, Rosen MR, Marks AR (2005) Defective cardiac ryanodine receptor regulation during atrial fibrillation. *Circulation* 111: 2025–2032.
- Voegeli TS, Wintink AJ, Chen Y, Currie RW (2008) Heat shock proteins 27 and 70 regulating angiotensin II-induced NF-kappaB: a possible connection to blood pressure control? *Appl Physiol Nutr Metab* 33: 1042–1049.
- Williams RS, Benjamin IJ (2000) Protective responses in the ischemic myocardium. *J Clin Invest* 106: 813–818.
- Wu ZK, Iivainen T, Pehkonen E, Laurikka J, Tarkka MR (2003) Arrhythmias in off-pump coronary artery bypass grafting and the antiarrhythmic effect of regional ischemic preconditioning. *J Cardiothorac Vasc Anesth* 17: 459–464.
- Yaglom JA, Gabai VL, Meriin AB, Mosser DD, Sherman MY (1999) The function of HSP72 in suppression of c-Jun N-terminal kinase activation can be dissociated from its role in prevention of protein damage. *J Biol Chem* 274: 20223–20228.
- Yue L, Feng J, Gaspo R, Li GR, Wang Z, Nattel S (1997) Ionic remodeling underlying action potential changes in a canine model of atrial fibrillation. *Circ Res* 81: 512–525.
- Zhao Y, Wang W, Qian L (2007) Hsp70 may protect cardiomyocytes from stress-induced injury by inhibiting Fas-mediated apoptosis. *Cell Stress Chaperones* 12: 83–95.
- Zou N, Ao L, Cleveland JC Jr, Yang X, Su X, Cai GY, Banerjee A, Fullerton DA, Meng X (2008) Critical role of extracellular heat shock cognate protein 70 in the myocardial inflammatory response and cardiac dysfunction after global ischemia-reperfusion. *Am J Physiol Heart Circ Physiol* 294: H2805–2813.