

Review

The Role of Hsp70 in Adaptation to Adverse Conditions and Its Possible Medical Application

Michael B. Evgen'ev^{1,*}, Sergei B. Onikienko², Lubov N. Chuvakova¹, David G. Garbuz¹, Olga G. Zatsepina¹

¹Institute of Molecular Biology, Russian Academy of Science, 119991 Moscow, Russia

²Saint-Petersburg Scientific Center, Russian Academy of Science, 194064 Saint-Petersburg, Russia

*Correspondence: misha672011@yahoo.com (Michael B. Evgen'ev)

Academic Editors: Alexander E. Kabakov and Mateusz Maciejczyk

Submitted: 12 December 2022 Revised: 10 January 2023 Accepted: 13 January 2023 Published: 8 February 2023

Abstract

In the present era of global warming and dramatically increased environmental pollution posing a threat to animal life, the understanding and manipulation of organisms' resources of stress tolerance is apparently a question of survival. Heat stress and other forms of stressful factors induce a highly organized response of organisms at the cellular level where heat shock proteins (Hsps) and in particular Hsp70 family of chaperones are among the major players in the protection from the environmental challenge. The present review article summarizes the peculiarities of the Hsp70 family of proteins protective functions being a result of many millions of years of adaptive evolution. It discusses the molecular structure and specific details of *hsp70* gene regulation in various organisms, living in diverse climatic zones, with a special emphasis on the protective role of Hsp70 in adverse conditions of the environment. The review discusses the molecular mechanisms underlying Hsp70-specific properties that emerged in the course of adaptation to harsh environmental conditions. This review also includes the data on the anti-inflammatory role of Hsp70 and the involvement of endogenous and recombinant Hsp70 (recHsp70) in proteostatic machinery in various pathologies including neurodegenerative ones such as Alzheimer's and Parkinson's diseases in rodent model organisms and humans *in vivo* and *in vitro*. Specifically, the role of Hsp70 as an indicator of disease type and severity and the use of recHsp70 in several pathologies are discussed. The review discusses different roles exhibited by Hsp70 in various diseases including the dual and sometimes antagonistic role of this chaperone in various forms of cancer and viral infection including the SARS-Cov-2 case. Since Hsp70 apparently plays an important role in many diseases and pathologies and has significant therapeutic potential there is a dire need to develop cheap recombinant Hsp70 production and further investigate the interaction of externally supplied and endogenous Hsp70 in chaperonotherapy.

Keywords: heat shock proteins; adaptation to environmental stress; inflammation; proteostasis; recombinant Hsp70 biomedical applications

1. Introduction

Ritossa's description of morphological changes in the structure of *Drosophila* polytene chromosomes (formation of puffs) after heat shock (HS) opened a new page in the study of inducible genes and the entire eukaryotic genome [1]. Interestingly, this pioneering work was not immediately appreciated and was first regarded as a kind of amusing mishap, characteristic only for *Drosophila*. It was not until the mid-seventies, when the molecular consequences of HS were discovered [2,3], that an avalanche of studies began on this system, which appeared to be present in the genomes of all organisms studied [4–9]. Its simplicity and high reproducibility have made the HS gene system very popular for studying the subtle mechanisms of eukaryotic gene regulation and genome function in general [10–18].

A comprehensive study of the HS gene system carried out on different objects from bacteria and flies to humans has shown the presence of different groups of HS genes and demonstrated their role in cell and organism functioning in general. Excellent reviews are describing the structure and

function of different classes of HS genes [19–23].

In particular, the role of different classes of Hsp and especially Hsp70 in the maintenance of cellular proteostasis, both under normal physiological conditions and after stress, has been demonstrated in different laboratories [24–28]. It was also shown that the Hsp70 protein plays the central role in the folding and assembly of newly synthesized proteins, refolding of misfolded and aggregated proteins, membrane translocation of organelle and secretory proteins, and control of the activity of various regulatory proteins [29]. On the other hand, field biologists, as is often the case, did not immediately pay attention to the *hsp* gene system, and started to study these genes in an ecological aspect later [30–35].

It was logical to assume that if Hsps are really so important for the normal functioning of the cell and the organism as a whole under normal and stressful conditions, the *hsp* gene systems in organisms living in arid climatic zones or areas with rapid temperature fluctuations should characteristically differ from similar systems of forms from tem-



perate climate zones. In fact, it turned out to be so. Several examples have shown that thermophilic species are capable of synthesizing Hsps at higher temperatures compared to phylogenetically similar species from temperate zones [7,30,36,37].

In recent decades, molecular ecology has accumulated numerous facts and observations indicating the important role of various Hsps and especially the main stress protein Hsp70 for the adaptation of organisms to adverse conditions, as well as describing how the structure of *hsp* genes and correspondent proteins evolved for their optimal functioning under various conditions, including extreme ones [7,38,39]. Heat shock genes play an important and sometimes even pivotal role in adaptation to changing environmental conditions not only in eukaryotes, but also in bacteria, especially in pathogens and symbionts [8].

Although we are aware of the role of other Hsp groups in adaptation to various stress conditions [40–44], in this review we will pay special attention to the role of the major stress protein (Hsp70) in adaptation to adverse conditions. Besides, herein we consider the examples of the use of recombinant Hsp70 for the treatment of various diseases and pathologies in both model animals and humans available in the literature.

2. The Role of *hsp70* Genes in Adaptation

2.1 Peculiarities of Regulation of HS Gene Expression

It was established that the rapid activation of all groups of *hsp* genes under stress at the transcriptional level is provided by the same protein factor (Heat Shock Factor-HSF1). All *hsp* genes contain heat shock elements (HSE) consisting of several inverted repeats of the consensus nGAAn sequence in the proximal part of the promoter region [45,46]. In the absence of stress, HSF1 monomers interact with Hsp70 and Hsp90 to form an inactive complex. During heat shock, an increase in the number of denatured, unfolded proteins in the cell causes decay of the Hsps and HSF1 complex, and Hsps bind to the damaged proteins. It should be mentioned that activation of HSF1 involves several steps, including extensive posttranslational modification, translocation into the nucleus, trimerization, and binding to heat shock elements (HSE) at promoter sites, inducing transcription of all HS genes [47–49].

To date, it has been shown that the regulation of Hsps expression in particular is more complex and may differ in different tissues and organs [49–51]. The search for tissue-specific regulators of HS gene induction is important for understanding the role of Hsps in adaptation as well as in the treatment of many diseases, especially neurodegenerative ones.

2.2 Adaptation Mechanisms at the Genome Level

An important feature of genes belonging to the *hsp70* family is their multi-copy nature. Indeed, in the vast majority of the studied animal species, *hsp70* genes are represented

by several copies. Thus, in *Drosophila*, there are 5 to 6 copies of genes of this family induced under the action of HS [52,53]. Thirteen genes of the *hsp70* family were found in the human genome [54,55], and several of them are expressed constitutively, sometimes only in certain tissues, while other members are induced under various stress influences [54,56]. It is also important to note the unusually high homology between genes encoding *hsp70* in different organisms. Thus, the homology between *Drosophila* and human *hsp70* genes reaches 72%, and between human and *E. coli* *hsp70* genes – 50% [57]. Interestingly, in *Drosophila* the basic structure of *hsp70* genes is a pair of genes in an inverted position, and in thermophilic *Drosophila* species, the number of *hsp70* genes tends to be higher than in species from temperate zones [7,52,58]. Another important and obviously adaptive feature of *hsp70* genes is their cluster organization. It turned out that in thermophilic species these clusters have a more compact structure, which apparently provides their cumulative effect under stress. It was also shown that the high homology of the *hsp70* gene copies included in such a cluster is maintained by gene conversion [7,59,60].

Characteristic features have been described for *hsp70* genes at the chromatin level as well. Thus, back in the 1980s, it was shown that the regulatory regions of the *Drosophila* *hsp70* genes are represented by “naked” DNA, that is, devoid of nucleosomal structure [61–63]. Characteristically, the promoters of *hsp* genes under normal temperature are associated with a suspended, positioned RNA polymerase II (RNAPII) [64]. Such promoter arrangement is obviously necessary for rapid and efficient turn-on of all *hsp70* genes under stress. Interestingly, naked DNA regions in *hsp70* genes are hot spots for the insertion of mobile elements both in the case of P-mutagenesis and spontaneous transposition of mobile elements, which apparently ensures rapid evolution of these regulatory regions and *hsp70* genes in general [65,66]. At this end, in an *in vitro* system, it has been shown that *hsp70* gene promoters of the camel, whose cells are significantly more heat tolerant compared to human cells [32,67], have higher “strength” at elevated temperatures compared to orthologous human *hsp70* promoters [68].

2.3 Peculiarities of *hsp70* Genes Expression in Species from Contrasting Ecological Niches

Studies of *hsp70* gene expression in different organisms from temperature-contrasting climatic zones have allowed to describe characteristic features of this system functioning under normal physiological conditions and stress [7,34,37,39,69–73]. In this regard, we should recall the pioneering work of Ulmasov *et al.* [71] who showed that the level of Hsp70 in the body of desert, temperature-resistant lizards changes characteristically depending on the ambient temperature during the day. Adaptive changes in the level of Hsp70 expression depending on the season were

also shown in other studies [74,75], indicating the ecological importance of Hsps synthesis in natural populations. It should be emphasized that constitutive expression of inducible Hsp70 genes in different species from arid zones or regions with sharply changing temperatures during the day is probably the main feature characteristic for such species. This phenomenon has been described for such different groups of animals as lizards, ants, flies, tidal zone dwellers, Baikal amphipods, etc. [7,32–34,37,39,69–73].

The constitutive expression of Hsp70 and other chaperones in the cells of thermoresistant species apparently allows such forms to maintain proteostasis and function normally under high temperatures without switching on additional adaptogenic systems. An alternative system of regulation of *hsp70* genes has usually been observed in temperate forms, that are rarely exposed to sharp temperature fluctuations. In such species, of which *Drosophila* is a typical example, under normal temperature conditions Hsp70 synthesis is at a low, difficult-to-detect level, but when temperature or other stress influences are increased, rapid and intense activation of all *hsp* genes and, in particular, of the *hsp70* gene battery is observed [76]. Rapid and intense induction of Hsp70 in *Drosophila* species apparently determines their ability to acclimatize. *Drosophila* species inhabiting habitats with higher temperatures are characterized by a larger number of *hsp70* genes, a compact organization of their clusters, “stronger” promoters, and, consequently, more powerful induction of Hsp70 with increasing temperature, compared to species inhabiting temperate habitats [58,77,78]. It should be noted that a low, basal level of Hsp70 observed in *Drosophila* species is still, apparently, necessary for the normal functioning of flies under non-extreme physiological conditions. Thus, it has been shown that *D. melanogaster* flies with deletion of all copies of *hsp70* genes have impaired survival after severe HS, as well as a tendency to develop various neurodegenerative processes [79]. It has also been demonstrated that males of this strain with deleted *hsp70* genes have completely impaired memory and learning ability; moreover, low irradiation resulted in reduced viability in flies lacking *hsp70* genes [80,81]. It should be noted, that the observed patterns of Hsp70 synthesis of the “*Drosophila* type” characterized by the low constitutive synthesis of Hsp70 under normal physiological conditions and the “Lizard and Ant type”, when a pronounced constitutive synthesis of Hsp70 is observed in the cells of arid zone animals, by no means exhaust the spectrum of eukaryotic genome response to extreme environmental conditions. Several examples from different animal groups were described that do not fit into this simple scheme [82–85]. For example, it was shown that selected *D. melanogaster* flies capable of living and reproducing at 31 °C are characterized by lower Hsp70 induction during HS compared to other strains of this species [86,87]. Cold-living (e.g., Antarctic) species that dwelled for millions of years under stable low-temperature conditions and

completely lost the ability to induce *hsp* genes when temperature rises have also been described [83]. Such forms are characterized by a loss of canonical HSEs within *hsp*s promoters [88] (Fig. 1A).

It should be noted that the *hsp70* genes system present in all eukaryotic organisms is a complex, well-tuned adaptive mechanism that underwent millions of years of evolution, and any experimental manipulation with this balanced system can lead to unpredictable consequences. For example, transgenic *Drosophila* strains with the experimentally increased number of *hsp70* copies have reduced viability and other developmental disorders [89].

It is of note that heat shock genes play essential roles in adaptation to extreme environmental conditions in bacteria as well, particularly in pathogens and symbionts. In bacteria, Hsp70 proteins contribute to their survival in a hostile environment. It is noteworthy that the adhesion of bacteria to host cells is mediated by both host and bacterial Hsp70 (DnaK). After infecting the host, it is DnaK that initiates the processes of bacterial survival and induces the host’s immune response. Any mutation in the DnaK gene reduces the viability of bacteria in the host [9].

2.4 Adaptation at a Protein Level

Convergent evolution resulted in more similar amino acid sequences in inducible Hsp70 in different heat-tolerant, phylogenetically distant species compared to evolutionarily close forms living in milder climates [90,91]. Probably, in this case, natural selection acted at the amino acid level, selecting variants of Hsp70 capable of providing maximum protection under extreme temperatures and other forms of stress. To test this assumption, we introduced into the genome of *D. melanogaster* with deleted all *hsp70* genes a copy of the orthologous gene isolated from a Stratiomyidae (Diptera) species whose larvae live in hot, sulfurous springs of the Kuril Islands and can withstand temperatures up to 42–45 °C [70]. Our experiments showed that the larvae of such a transgenic strain containing a single *hsp70* gene isolated from the genome of this highly thermostable species can withstand higher temperatures compared to the *D. melanogaster* larvae containing one copy of its own endogenous *hsp70* gene [92]. Characteristically, experiments using *in vitro* refolding luciferase assay demonstrated that Stratiomyidae Hsp70 exhibited higher refolding capacity in comparison with *D. melanogaster* Hsp70 and even human paralog [91]. Moreover, using differential scanning calorimetry we showed that the ATP-binding domain of *Stratiomyidae singularior* Hsp70 is stable at temperature 4 degrees higher than that of *Drosophila* Hsp70 [91]. It is of note, that two isoforms of inducible Hsp70 were detected in exceptionally thermoresistant arid zone lizards, and the synthesis of each of these forms occurred at different elevated temperatures [71]. Thus, many researchers have shown using various objects the protective, chaperone role of Hsp70 under various extreme conditions, as well as the participa-

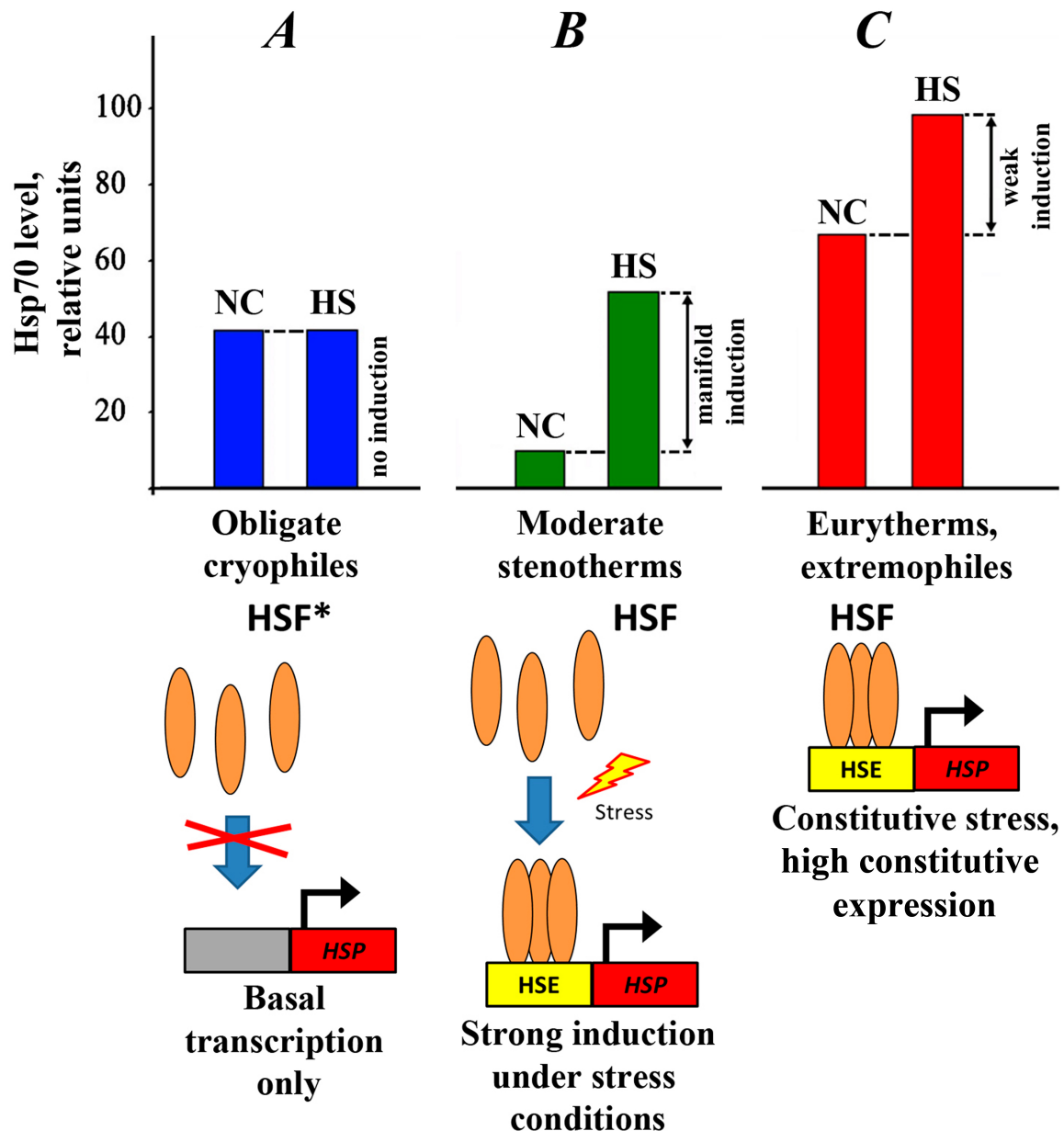


Fig. 1. Peculiarities of heat shock response regulation in organisms that dwell in temperature contrasting habitats. (A) Cold-living (obligate cryophiles) Antarctic forms dwelling in stable low-temperature conditions lost the ability to induce *hsp* genes when the temperature rises due to loss of HSE sequences within *hsp* genes promoters. (B) Species living in a temperate climate zone with rare sharp temperature fluctuations normally synthesize low levels of Hsps, that are dramatically induced after HS. (C) Extremophiles, thermoresistant species living in hot climates and constitutively synthesize high levels of Hsp70, which increases slightly after HS. HSF* - in some species from the group of obligate cryophiles, the ability to activate HSF is lost due to mutations in the *hsf* gene.

tion of Hsp70 in the maintenance of cellular proteostasis, both under normal and stressful conditions.

2.5 The Interaction of Hsp70 with Cellular Membranes in the Course of Adaptation

Generally speaking, Hsp70 plays an exceptionally important role in cell membrane function [93]. As early as 1986, Hightower suggested that Hsp70 might associate directly with plasma membrane lipids [94,95]. It was later

demonstrated, that Hsp70 plays an important role in whole organism protection under stress through the maintenance of barrier functions of endothelium and epithelium [96]. It is known that with elevated body temperature a general intoxication of the body usually occurred associated with the release of toxins and bacteria (e.g., endotoxins, lipopolysaccharides) from the gut, as well as an increase in circulating cytokines in response to the toxins. This is associated with an increase in the permeability of the intestinal epithelium

during intense exercise and heat stress [97–99]. Increased expression of Hsp70 has been shown to prevent heat shock-induced breakdown of the epithelial barrier by stabilizing tight junctions between epithelial cells and improving its recovery from HS and other forms of stress [100–102]. The role of Hsp70 in resistance to heat-induced damage in humans was demonstrated in a study of Kuennen *et al.* [103], which showed that inhibition of the HS response by Hsp70 inhibitor quercetin during 7 days of heat acclimatization resulted in a significant increase in gastrointestinal permeability and endotoxin leakage after acute heat stress.

In addition to protecting the intestinal epithelium from stressors, Hsp70 plays an important role in maintaining the integrity of the vascular epithelium [104,105]. More recently Hsp70 has been shown to interact selectively with negatively charged phospholipids, particularly phosphatidylserine (PS) [106–110] inside liposomes, after which Hsp70 is incorporated into the lipid bilayer, forming high molecular weight oligomers [107] that have ionic conduction channel properties [111,112].

Besides, phosphatidylserine in membranes, Hsp70 interacts with several other negatively charged phospholipids, most notably bis(monoacylglycero)phosphate (BMP), the main phospholipid of lysosome membranes [113,114]. The binding of Hsp70 in lysosomal membranes to the negatively charged phospholipid BMP imparts stability to this compartment by inhibiting lysosomal membrane permeabilization and preventing the release of lysosomal proteases and cathepsins into the cytosol in response to stressful conditions, thereby preventing cells and organ death [113,115].

There is extensive evidence that Hsp70 stabilizes biological membranes [116–118]. Thus, the temporal association of Hsps with membranes can restore bilayer fluidity and thus maintain membrane functionality under stress [116]. The accumulated data show that Hsp70 is preferentially embedded in cholesterol-rich microdomains (“rafts”) that serve as major platforms for the assembly and sorting of signal transduction complexes in the membranes (Fig. 2) [119]. It has been also demonstrated that Hsp70 can play a role in maintaining the stability of lipid raft-associated signal transduction complexes after stress [120]. Moreover, after various stresses and in some forms of cancer, free Hsp70 accumulated in the cell can be incorporated into the plasma membrane and be further released into the extracellular space both in free form and in the form of lipid vesicles of exosomes [121], lysosomal endosomes [122] or the context of cholesterol-rich microdomain-rafts (Fig. 2) [123].

It has been also shown, that extracellular Hsp70 (eHsp70) interacts with several receptors on the cell surface and may be internalized by target cells and initiate various signaling cascades. Thus, several receptors such as TLR2/4, CD91, CD40, and SR appear to be responsible for eHsp70 phagocytosis [124,125].

The unique protective characteristics described for Hsp70 in numerous *in vivo* and *in vitro* studies served as a

basis for the investigation of the involvement of this protein in various diseases and its possible therapeutic potential.

3. Role of Hsp70 in Various Pathologies, and Use in Medicine

3.1 Hsp70 as a Biomarker of Pathological Processes and Diseases

Studies of the role of Hsps, and Hsp70 in particular, under normal physiological conditions and after the stress has demonstrated the crucial role of this system in proteostasis at different steps of protein synthesis and degradation [5,7,24,126,127]. In substrate degradation, Hsp70 explores the ubiquitin-proteasome system as well as various autophagy pathways [27]. Naturally, it was interesting to find out how the Hsp70 family members function in the case of various pathologies accompanied by disturbances in cellular protein synthesis and degradation, and whether the levels of Hsp70 and other chaperones in various tissues can be used as an indicator of disease and physical condition in humans. Indeed, it was shown that during normal aging, the level of Hsp70 is significantly reduced in animals and humans [128–130]. Hsp70 levels decrease in humans during aging in most organs, including neurons, leading to the accumulation of damaged and unfolded proteins. This contributes to proteotoxicity, which may lead to the development of age-related diseases due to neurodegeneration. Aging also leads to the attenuation or alteration of many signaling pathways, as well as the expression of several transcription factors, including major heat shock factor (HSF1) [129–131]. Characteristically, aging decreases the level of Hsp70 induction after stress [132]. A decrease in the expression level of Hsp70 regardless of age is observed in such human diseases as pulmonary fibrosis, various types of “myopathies”, cholesterol sphingolipids, sphingolipidoses, diabetes, obesity, etc. [133–137]. In these diseases, reduced levels of both HSF1 and various members of the Hsp70 family can be observed in adipose tissue, liver, muscle, neurons, and vascular beds of patients. Interestingly, different baseline, constitutive levels of various chaperones, and particularly Hsp70, detected in patients often determine the severity of the disease course and the outcome of various diseases, including atherosclerosis and COVID-19 [138,139]. Therefore, the level of endogenous Hsp70 in cells is used, not only as a biomarker of various diseases but also to monitor the results of therapy for various human diseases [139,140]. In particular, Hsp70 levels have been used as a biomarker of prostate diseases and bronchopulmonary dysplasia (BPD) for targeted therapies [135,141]. Thus, it is known that Hsp70 levels are associated with a state of oxidative injury in the development of BPD [135]. Significant changes in Hsp70 levels in neuronal tissues have been reported in normal aging as well as in patients with neurodegenerative diseases accompanied by the formation of protein aggregates, including Alzheimer disease (AD), Parkinsonism, and amyotrophic lateral sclerosis.

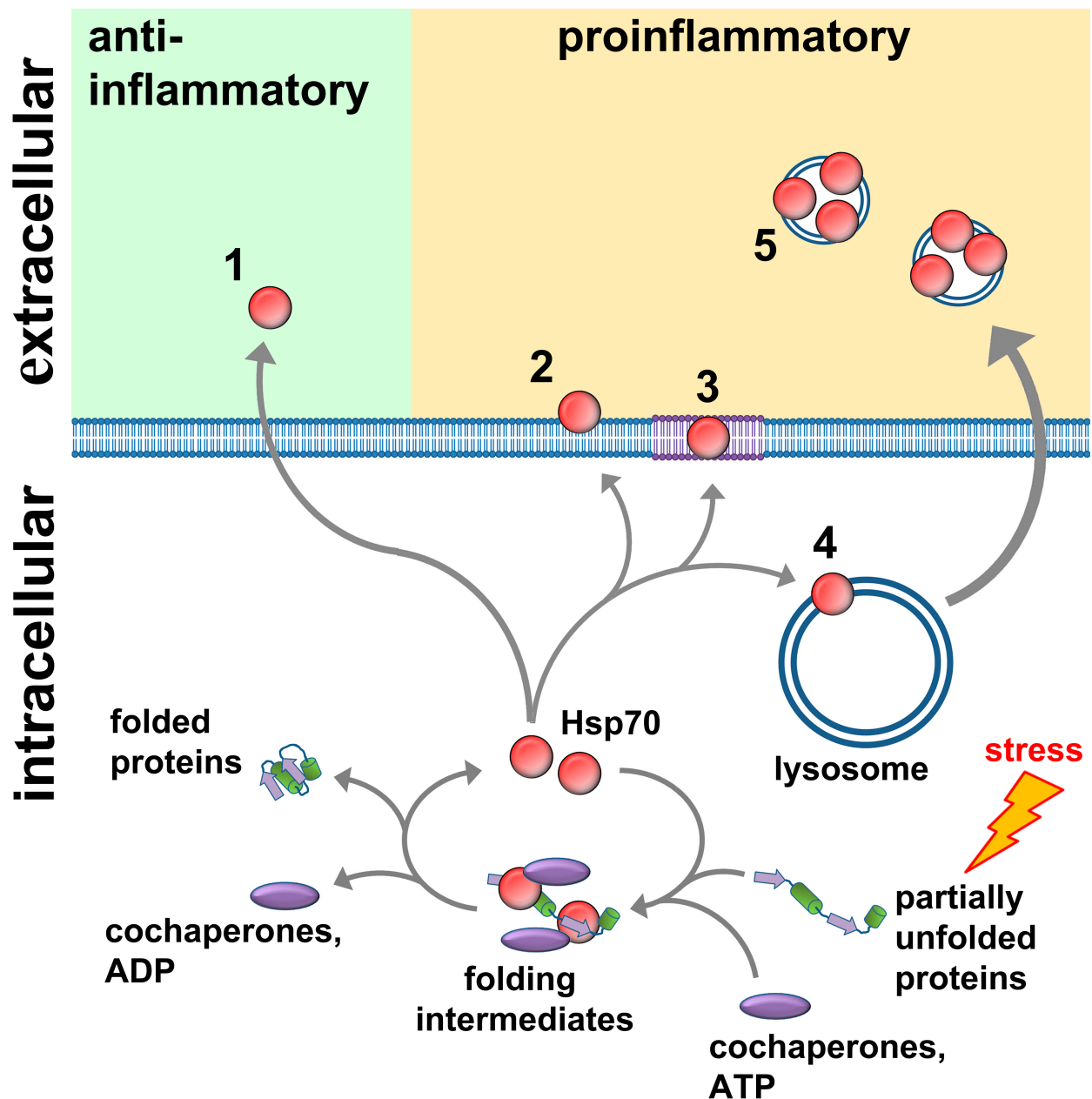


Fig. 2. Intracellular Hsp70 functions as a molecular chaperone, preventing aggregation and restoring native conformation of partially unfolded proteins with the participation of ATP and several co-chaperones. Extracellular Hsp70 may exist in two different forms, free (1) and membrane-bound (2–5). (1) Free protein releases from cells after necrosis under the action of various damaging factors, or due to transmembrane transport by a non-classical mechanism that does not require a signal sequence, involving the ABC transporter. (2, 3) Membrane-bound Hsp70 comes to the surface of the cell by interaction with phosphatidylserine as a part of the inner lipid layer of the membrane and subsequent transfers together with the phosphatidylserine molecule into the outer layer and lipid rafts. (4, 5) a result of secretion of Hsp70 as a part of membrane vesicles (ectosomes and exosomes), Free, unbound to membrane structures Hsp70 and exogenous recHsp70 predominantly play an anti-inflammatory role, while membrane-bound Hsp70 may play a pro-inflammatory role. By binding to the membranes of lysosomes, Hsp70 prevents their permeabilization and release of lysosomal secretion into the cell.

rosis (ALS) [142–144]. Determination of the Hsp70 levels and some other chaperones can serve as an indicator of the development of pathology in the case of AD. However, changes in Hsp70 levels in various pathologies may be multidirectional for members of the Hsp70 family in different

brain tissues of the patients. Thus, in AD, HSPA8 levels are increased in brain cells, while HSPA1A and HSPA2 levels are decreased [145]. This may be explained by the fact that members of the Hsp70 family are not equally expressed in different human tissues under normal conditions [54].

It is known that a significant increase in Hsp70 levels is often observed on the surface of cancer cells of different origin [143,146], which may hinder anti-cancer therapy. Thus, the presence of Hsp70 on the surface of cancer cells and its interaction with lysosome membranes have been envisioned as potential therapeutic targets [146,147]. Generally speaking, Hsp70 membrane association is a key component in the extracellular export of these proteins [148]. The secreted Hsp70 can additionally act as an effective neuroprotector, increasing the survival of neurons in various proteinopathies, as has been demonstrated in Alzheimer's and Parkinson's disease models. In this regard, recHsp70 and inducers of endogenous Hsp70 synthesis may be considered as candidate therapeutics with immune-modulating and neuroprotective properties. Since various members of the Hsp70 family, are predominantly expressed in different tissues and organs [54], it should be taken into account when using Hsp70 as an indicator of pathology and when performing therapies aimed at normalization or increase of Hsp70 levels in a particular tissue in various diseases. The changes in the levels of various Hsp70 family members in various diseases and pathologies demonstrated by many researchers served as a basis for conducting experiments on correction of Hsp70 levels in model animal and human cells both by induction or suppression of endogenous Hsp70 synthesis and by introduction of exogenous recHsp70 by various means.

3.2 Experimental Modulation of Endogenous Hsp70 Levels for Therapy of Various Diseases

A variety of experimental approaches has been used to normalize the level of endogenous Hsp70 impaired in various pathologies, including aging and chronic diseases. In some cases, researchers activated the synthesis of all Hsps groups, i.e., increased the heat shock response (HSR) in general, for example, by increasing the temperature of the body and proved, for example, the role of HS-induced Hsp70 for muscle regeneration in a mouse model [149]. Sometimes, simultaneous change of the expression level of all members of the Hsp70 family and other chaperones has been achieved by regulation of the main heat shock transcription factor (HSF1) synthesis [150]. In several other cases, the investigators have succeeded in selectively increasing the synthesis of only Hsp70. Such an approach has been recently applied to suppress a cytokine storm in COVID-19 patients [151]. Sometimes researchers used a combined approach, inducing HSR and administering recHsp70, for example, to treat diseases such as diabetes mellitus [137]. It is noteworthy to mention in this regard the results of Lydia Kitchen, who used photobiomodulation therapy (irradiation by 1060–1080 nm light), a noninvasive method of modulating Hsp70 and nitric oxide (NO) levels to treat infection in the case of COVID-19 and other coronaviruses [152], and the application of selective Hsp70 induction in the treatment of ultra-rare neuromuscular dis-

order *GNE* myopathy [134]. Along these lines, a specific low molecular weight inducer of Hsp70 ("KD-23") exhibited a therapeutic effect in a cellular model of craniocerebral injury [153]. Similarly, selective induction of endogenous Hsp70 with dioscin or geranylgeranylacetone (GGA) has been successfully applied in various lung injuries including fibrosis, and to restore blood flow after ischemia [154,155].

A separate group of naturally occurring substances capable of increasing the level of Hsp70 and other chaperones in the cell is represented by the so-called "adaptogens", i.e., natural substances such as ginseng, *Eleutherococcus*, *radiola rosea*, etc. These substances are usually of plant origin, and their use is often accompanied by induction of Hsp70 in various human tissues and organs [156–159]. Such substances have long been successfully used in China and other countries to heal wounds, as well as in several chronic and aging-related human diseases. The successful use of inducers of the entire HSR system as well as experimental selective modulation of the level of endogenous Hsp70 in the treatment of various diseases showed the promise of this major stress protein as a therapeutic agent and provided the basis for the production and use of recHsp70 in medicine.

3.3 The Use of Recombinant Hsp70 in Model Experiments and Clinical Application

Various laboratories have developed various methods for the expression and isolation of recHsp70. RecHsp70 has been isolated from a variety of animal and plant objects, produced in bacteria, and expressed in various cell cultures, including human cells, as well as in the milk of transgenic animals [139,160–163]. The use of recHsp70 has shown its efficacy in many models of human diseases with a variety of administration methods, from intranasal and intravenous to intracranial [161,164,165].

Thus, preventive intravenous administration of human recHsp70 in a rat model of sepsis normalized lipopolysaccharide (LPS)-induced blood pressure abnormalities and biochemical blood parameters and significantly reduced animal mortality [166]. Similar results were obtained with intravenous administration of a single dose of recHsp70 in an LPS-induced mouse peritonitis model [167].

In a mouse model of aging, we were able to show that intranasal subchronic administration of recombinant human Hsp70 significantly extends animal life and had a profound rejuvenation effect in neuronal cells of the cortex and hippocampus. Interestingly, an increase in lifespan when Hsp70 was sub-chronically injected was observed only in males [164]. In the mouse aging model used, as well as in two validated AD models, it was demonstrated that subchronic intranasal administration of recHsp70 improved memory and learning ability and reduced the level of such markers of aging as lipofuscin and amyloid level [164,168].

The method of intranasal administration of human recHsp70 used also gave encouraging results in a model

of photothrombotic stroke in mice [169]. Similarly, in a mouse model of diabetes, researchers were able to increase insulin sensitivity with intranasal injections of cheap Hsp70 isolated from alfalfa [161]. Kirkegaard *et al.* [162] first applied recHsp70 and its inducer to treat various forms of lysosome storage diseases, using both various animal models and cell cultures of patients, with various disorders of lysosome biogenesis (aggregation).

Administration of recHsp70 in different ways led to positive therapeutic effects when used *in vitro* and animal models for such different diseases and pathologies as diabetes, prostatitis, and various forms of lung damage, including various forms of fibrosis [135,137,141,170]. RecHsp70 administration also resulted in a reduction of cholesterol levels in human cells and model animals [171]. In this context, the exploration of the lentiviral constructs to express Hsp70 in different tissues in the case of lung injury should be mentioned [172]. It's of note, however, that in various forms of fibrosis, viral infections including COVID-19 cases, as well as in different forms of cancer, the administration of recombinant Hsp70 or its experimental induction may have different, sometimes multidirectional effects depending on the disease, dose of the chaperone and time of administration [140,146,147]. Thus, on the one hand, the increased content of endogenous Hsp70 on the cell membrane observed in many forms of cancer can hinder successful anti-cancer therapy, on the other hand, administration of recHsp70 in Margulis' laboratory and several other groups had a significant therapeutic effect both *in vitro* when used on various cancer cells and *in vivo* in pilot experiments [173–176], where injections of recombinant human Hsp70 were made directly into tumors of volunteers [176]. In their study of melanoma cells, this group showed that injection of recombinant Hsp70 triggers extracellular transport of its endogenous homolog in soluble form and as EVs (extracellular vesicles), that penetrate neighboring cells, resulting in a dramatic reduction in the growth rate of cancer cells [177]. In another study on carcinoma cases, the patients were injected with dendritic cells containing *hsp70* mRNA induced by electroporation, and a positive effect was also observed in several patients [178].

Although large-scale studies on the role of Hsps and especially Hsp70 in various diseases, including cancer, as well as in normal aging, are currently underway in various laboratories, there are only a relatively small number of researchers taking practical steps to apply recombinant Hsp70 in the clinic. This is due, firstly, to the high cost and low availability of the recombinant human Hsp70 preparation and, most importantly, to the insufficient data and analysis of the consequences of Hsp70 application in patients, given the variety of interactions of Hsp70 with different proteins and cellular signaling systems.

3.4 Molecular Mechanisms Underlying Protective Effects of Hsp70 in Different Diseases

Intensive research on the role of endogenous and recombinant Hsp70 in aging, as well as in a variety of diseases and pathologies in model animals and humans resulted in the description of a variety of factors and signaling pathways with which Hsp70 interacts in carrying out its protective functions. Numerous excellent reviews are describing the functions of Hsp70 under normal conditions and after stress [5,24,126,141,144,177]. Therefore, in this section, we will only briefly discuss the known properties of Hsp70 that can be used in the therapy of some human diseases.

It should be noted that generally speaking, Hsp70 has two main functions in the body. First, members of this family together with Hsp90 and co-chaperones (e.g., Hsp70-Hsp90 Organizing Protein (HOP)) provide proteostasis in cells under normal conditions and stress [7,24,126,127]. Secondly, it was shown that Hsp70 can excrete from many types of cells into the intercellular medium and interact with several receptors on the surface of cells responsible for innate immunity (specially TLR2/4 receptors). Initially, results indicated that extracellular Hsp70 (eHsp70) causes the activation of macrophages and neutrophils and the production of pro-inflammatory cytokines. Based on these data, it has been suggested that Hsp70 represents a “danger signal”, activating the immune system and thus playing a regulatory “cytokine” role therefore Hsp70 is often called “chaperokine” [179].

However, later it was shown that these pro-inflammatory effects are often caused by the action of residual amounts of LPS, with which recombinant Hsp70 preparations used in these works obtained in *E. coli* were contaminated. Subsequently, it was shown that free extracellular Hsp70 obtained in eukaryotic expression systems has rather an anti-inflammatory effect. In inflammatory processes occurring during various diseases, both the induction of endogenous Hsp70 and the administration of LPS-free recHsp70 reduce the levels of proinflammatory cytokines (TNF- α , etc.), diminish reactive oxygen species (ROS) and NO content, and restore cellular redox status [130,140,151,155,180]. It was further shown that different forms of exogenous Hsp70, in particular, free and membrane-bound ones, can have the opposite effect on the activity of the immune system cells (see below).

The ability of Hsp70 to bind to a variety of receptors and compete with other ligands plays a major role in several cases. An example of such interactions is the anti-inflammatory role of recHsp70 described in several works in LPS-induced sepsis, where recombinant Hsp70 competes with LPS for binding to TLR-4, which leads to restoration of basic blood parameters and increased animal survival [166,167].

Interestingly, in our experiments administration of recHsp70 significantly reduced LPS-induced TLR4 expression in human macrophage cells (THP1) [181].

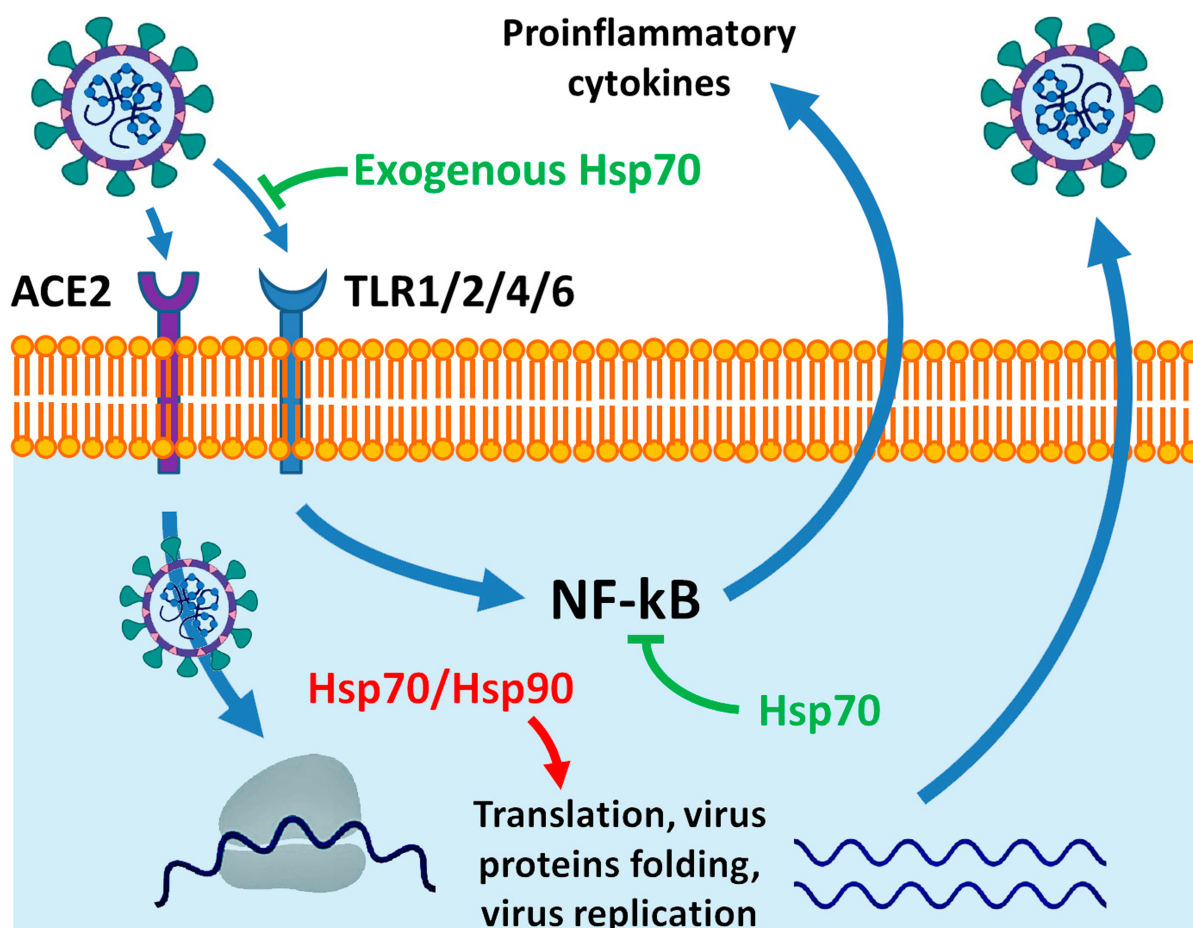


Fig. 3. In viral infection (SARS-CoV-2), extracellular HSP70, by binding to TLRs and probably to ACE2 receptors, blocks them and prevents the penetration of the virus inside the cell. Hsp70 also blocks NF- κ B activation and activates the synthesis of anti-inflammatory cytokines. On the other hand, several RNA viruses may explore the intracellular chaperone complex Hsp70/Hsp90 for their purposes to replicate and assemble viral particles in the host cells.

The ability to eliminate damaged proteins and prevent the formation of aggregates of different nature is obviously the most important protective function of Hsp70 described for such diseases as AD, Parkinsonism, ALS, and many others. It was shown that recHsp70 can compensate for the insufficient level of endogenous Hsp70 often observed in these pathologies [143,144]. Thus, in various models of AD, the intranasal administration of recHsp70 reduces β -amyloid levels and the number of amyloid plaques [168]. To this end, it has been described in detail how human Hsp70 and its co-chaperones DNAJB1 and Hsp110 interact to dissolve preformed fibrils of Parkinson's α -synuclein *in vitro* [182].

It is also worth mentioning an important and well-described role of Hsp70, which manifests itself as an anti-apoptotic agent at the early stages of this process [183,184]. This explains its crucial role in such diseases as cancer, pulmonary fibrosis, ischemia, neurodegeneration, etc. Thus, an investigation of the role of Hsp70 in lung dysfunction of model animals as well as in LPS-treated human alveolar epithelial cells showed that Hsp70 interacted with

KANK2 (ankyrin-repeat domain-containing protein), leading to reversed cell viability and reduced level of apoptosis-inducing factor (AIF) and, hence, apoptosis [172].

On the other hand, as we pointed out earlier, in some forms of cancer these anti-apoptotic properties of Hsp70 may interfere with anticancer therapy, forcing the use of Hsp70 inhibitors in these cases [185,186].

Hsp70 plays an important role in various viral diseases, including COVID-19. Here, as well as in the case of cancer cells Hsp70 can play a dual role being "a double agent". On the one hand, endogenous Hsp70 binding to TLR4 receptors may interfere with virus penetration into the cell. In addition, Hsp70 can prevent cytokine storm, which is the main cause of death in COVID patients, by causing degradation of the p65 subunit of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) subunit which is needed for cytokine storm [138]. On the other hand, some RNAi viruses use the host cell chaperone system Hsp70-Hsp90 at different stages of their cycle during viral infection [177,187] (Fig. 3).

As numerous studies demonstrated Hsp70 family members are highly effective vaccine adjuvants [188–190]. Thus, for example, the increase in the effectiveness of vaccination with laser irradiation of the skin in the area of influenza vaccine administration is associated with the induction of endogenous Hsp70, which has the properties of a vaccine adjuvant [191].

One of the most important protective functions of Hsp70 in many diseases is due to its role as an immunomodulator [192]. In particular, in our experiments exploring a mouse AD model, we showed that chronic Hsp70 administration changes the expression of several genes in the hippocampus of model animals. Most importantly using RNA-Seq, we identified a lot of differentially expressed genes in the hippocampus of old Tg mice compared with those of non-transgenic mice of the same age. Specifically, we observed Hsp70-induced upregulation of multiple genes participating in antigen processing and presentation especially the members of major histocompatibility complex (class I and II) in the brains of old 5XFAD Tg animals, suggesting that Hsp70 executes its beneficial role via activation of adaptive immunity [193].

In a model of infectious lung inflammation, it was shown that intraperitoneal injection of recHsp70 reduced the number of programmed death-1 (PD-1) positive T-lymphocytes in peripheral blood by several times and improved other important lung characteristics such as lung coefficient (Fig. 4) [181].

In a study of melanoma cells (B16), the addition of recHsp70 caused active synthesis and release of the endogenous analog from the cells in soluble and EVs forms, and both forms actively penetrated neighboring cells, leading to a sharp increase in Natural Killer (NK) cell toxicity towards melanoma cells [174]. This immunomodulatory effect of Hsp70 was due to the enhanced CD-8 positive response and massive anti-tumor cytokine accumulation.

In addition, numerous signaling systems and specific factors with which Hsp70 interacts in various diseases have been described in recent years. Thus, the interaction of Hsp70 with the androgen receptor (AR) and the important role of this chaperone in the pathogenesis of prostate cancer has been shown [141].

Another interesting example of the regulatory role of Hsp70 is the demonstrated ability of recHsp70 to lower cholesterol levels in primary human macrophage foam cells. Thus, RNA-seq analysis showed that the addition of Hsp70 led to the reprogramming of cell expression, including up-regulation of key targets of liver X receptors (LXR), master-regulators of whole-body cholesterol removal [171]. Valeria Calvaresi's group using native and mutant forms of recHsp70 described the details of Hsp70 binding to lysosomal membranes, which protects a whole series of lysosomal storage diseases [194].

It should also be noted that members of the Hsp70 family actively interact with other adaptive systems of the

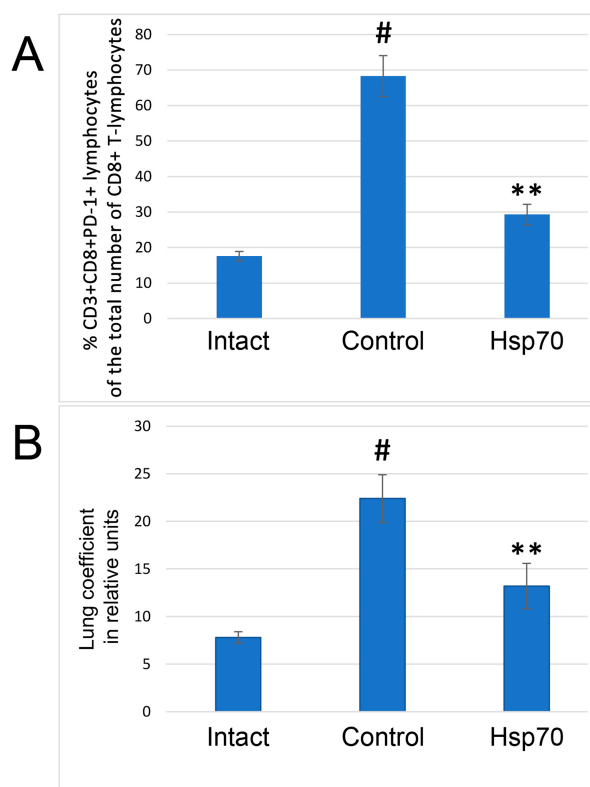


Fig. 4. Protective effect of Hsp70 in mouse the pneumonia model. RecHsp70 preparations decreased level of T lymphocytes with inhibitory PD-1 receptors in peripheral blood (A); and pulmonary coefficient (B) in the case of influenza-induced pneumonia in mice. [#] $p < 0.005$ versus intact; ^{**} $p < 0.01$ versus control.

body. In particular, they interact with the system of genes responsible for the synthesis and metabolism of hydrogen sulfide (H_2S) in the organism. Interestingly, the investigation of model animals has shown that inhalation of hydrogen sulfide mixtures has a protective effect during ischemia/reperfusion injury by induction of Hsp70 through P13/AKT/Nrf2 pathway [195].

Along these lines, in our experiments on a small sample of Covid patients [181], we have recently demonstrated that hot helium inhalation inducing Hsp70 followed by inhalation of hydrogen sulfide donor has a pronounced therapeutic effect preventing lung fibrosis in the patients. Moreover, in our *in vitro* and *in vivo* experiments we showed that different hydrogen sulfide donors as well as recHsp70 significantly reduced the level of proinflammatory cytokines, ROS, and other consequences of LPS-induced inflammation [181]. The contradictory and difficult-to-explain results sometimes obtained when analyzing the action of endogenous and recHsp70 may be related to the fact that different members of this family are preferentially synthesized in different organs and tissues and to the numerous tissue-specific posttranslational modifications to which members of the Hsp70 family are often subjected [180,196].

4. Conclusions

The discovery of the HS-inducible genes by Ferruccio Ritossa back in the early 1960s opened a new page in the study of the molecular mechanisms controlling the expression of the eukaryotic genome. While the main efforts of scientists were aimed at studying the structure and molecular regulation of *hsp* genes, the role of these genes and, in particular, the role of the main stress protein Hsp70 in adaptation to extreme environmental conditions attracted the attention of ecologists and molecular biologists much later. In the course of such studies, it was shown in a wide spectrum of model organisms and in nature that *hsp* genes, and *hsp70* in particular, play an important role in adaptation to extreme or rapidly changing environmental conditions. During adaptive evolution, the *hsp70* gene system in various eukaryotic organisms has undergone characteristic changes both at the level of structure, organization, and expression pattern of these genes in the genome and at the level of the structure of the Hsp70 family proteins themselves. Recent studies on the role of endogenous Hsp70 as well as recombinant Hsp70 in various diseases have demonstrated protective, anti-inflammatory functions of this protein to maintain proteostasis in the cells in various pathologies, including neurodegeneration and aging. The dual and sometimes antagonistic role of Hsp70 in cancer cells, fibrosis, and in infection with various viruses, including SARS, is discussed. The data accumulated in recent years represent a basis for the use of recHSP70 or its inducers in the therapy of many diseases.

Abbreviations

HS, heat shock; Hsp, heat shock proteins; eHsp70, extracellular Hsp70; recHsp70, recombinant Hsp70; HSF1, Heat Shock Factor; HSE, heat shock elements; RNAPII, RNA polymerase II; *D. melanogaster*, *Drosophila melanogaster*; TLR2/4, Toll-Like Receptor2/4; SR, scavenger receptors; EV, extracellular vesicles; BPD, bronchopulmonary Dysplasia; AD, Alzheimer disease; ALS, amyotrophic lateral sclerosis; NO, nitric oxide; HOP, Hsp70-Hsp90 Organizing Protein; ROS, reactive oxygen species; LPS, lipopolysaccharide; NF- κ B, Nuclear factor kappa-light-chain-enhancer of activated B cells; RNA Seq, RNA Sequencing; Tg, transgenic; PD-1, programmed death-1; NK, Natural Killer; AR, androgen receptor; LXR, liver X receptors; GGA, geranylgeranylacetone.

Author Contributions

Conceptualization and supervision—MBE; Writing—MBE, OGZ; Editing—DGG, LNC, OGZ, SBO; Formal analysis—OGZ, LNC, DGG; Visualisation—DGG. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

The work was funded by Russian Science Foundation Grants 17-74-30030 (MBE) and 19-14-00167 (DGG).

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Ritossa F. A new puffing pattern induced by temperature shock and DNP in *Drosophila*. *Experientia*. 1962; 18: 571–573.
- [2] Tissières A, Mitchell HK, Tracy UM. Protein synthesis in salivary glands of *Drosophila melanogaster*: relation to chromosome puffs. *Journal of Molecular Biology*. 1974; 84: 389–398.
- [3] Ashburner M, Bonner JJ. The induction of gene activity in *drosophila* by heat shock. *Cell*. 1979; 17: 241–254.
- [4] Craig EA. The stress response: changes in eukaryotic gene expression in response to environmental stress. *Science (New York, N.Y.)*. 1985; 230: 800–801.
- [5] Lindquist S. The heat-shock response. *Annual Review of Biochemistry*. 1986; 55: 1151–1191.
- [6] Feder ME, Hofmann GE. Heat-shock proteins, molecular chaperones, and the stress response: evolutionary and ecological physiology. *Annual Review of Physiology*. 1999; 61: 243–282.
- [7] Evgen'ev MB, Garbuz DG, Zatschina OG. Heat shock proteins and whole body adaptation to extreme environments. Springer: Berlin/Heidelberg, Germany. 2014.
- [8] Williams TA, Codoñer FM, Toft C, Fares MA. Two chaperonin systems in bacterial genomes with distinct ecological roles. *Trends in Genetics: TIG*. 2010; 26: 47–51.
- [9] Ghazaei C. Role and mechanism of the Hsp70 molecular chaperone machines in bacterial pathogens. *Journal of Medical Microbiology*. 2017; 66: 259–265.
- [10] Yost HJ, Lindquist S. RNA splicing is interrupted by heat shock and is rescued by heat shock protein synthesis. *Cell*. 1986; 45: 185–193.
- [11] Lee H, Kraus KW, Wolfner MF, Lis JT. DNA sequence requirements for generating paused polymerase at the start of *hsp70*. *Genes & Development*. 1992; 6: 284–295.
- [12] Lis J. Promoter-associated pausing in promoter architecture and postinitiation transcriptional regulation. *Cold Spring Harbor Symposia on Quantitative Biology*. 1998; 63: 347–356.
- [13] Shopland LS, Hirayoshi K, Fernandes M, Lis JT. HSF access to heat shock elements in vivo depends critically on promoter architecture defined by GAGA factor, TFIID, and RNA polymerase II binding sites. *Genes & Development*. 1995; 9: 2756–2769.
- [14] Lis J, Wu C. Transcriptional regulation of heat shock genes. *Transcription: Mechanisms and Regulation*. 1994; 459–475.
- [15] Tang H, Liu Y, Madabusi L, Gilmour DS. Promoter-proximal pausing on the *hsp70* promoter in *Drosophila melanogaster* depends on the upstream regulator. *Molecular and Cellular Biology*. 2000; 20: 2569–2580.
- [16] Lee C, Li X, Hechmer A, Eisen M, Biggin MD, Venters BJ, *et al*. NELF and GAGA factor are linked to promoter-proximal pausing at many genes in *Drosophila*. *Molecular and Cellular Biology*. 2008; 28: 3290–3300.

- [17] Petesch SJ, Lis JT. Rapid, transcription-independent loss of nucleosomes over a large chromatin domain at Hsp70 loci. *Cell*. 2008; 134: 74–84.
- [18] Lebedeva LA, Nabirochkina EN, Kurshakova MM, Robert F, Krasnov AN, Evgen'ev MB, *et al.* Occupancy of the *Drosophila* hsp70 promoter by a subset of basal transcription factors diminishes upon transcriptional activation. *Proceedings of the National Academy of Sciences of the United States of America*. 2005; 102: 18087–18092.
- [19] Pelham H. Heat-shock proteins. Coming in from the cold. *Nature*. 1988; 332: 776–777.
- [20] Morimoto RI, Kline MP, Bimston DN, Cotto JJ. The heat-shock response: regulation and function of heat-shock proteins and molecular chaperones. *Essays in Biochemistry*. 1997; 32: 17–29.
- [21] Jee H. Size dependent classification of heat shock proteins: a mini-review. *Journal of Exercise Rehabilitation*. 2016; 12: 255–259.
- [22] Richter K, Haslbeck M, Buchner J. The heat shock response: life on the verge of death. *Molecular Cell*. 2010; 40: 253–266.
- [23] Lang BJ, Guerrero ME, Prince TL, Okusha Y, Bonorino C, Calderwood SK. The functions and regulation of heat shock proteins; key orchestrators of proteostasis and the heat shock response. *Archives of Toxicology*. 2021; 95: 1943–1970.
- [24] Parsell DA, Lindquist S. The function of heat-shock proteins in stress tolerance: degradation and reactivation of damaged proteins. *Annual Review of Genetics*. 1993; 27: 437–496.
- [25] Parsell D. Heat shock proteins and stress tolerance. The biology of heat shock proteins and molecular chaperones. 1994; 457–494.
- [26] Morimoto RI, Tissières A. The biology of heat shock proteins and molecular chaperones. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1994.
- [27] Fernández-Fernández MR, Gragera M, Ochoa-Ibarrola L, Quintana-Gallardo L, Valpuesta JM. Hsp70 - a master regulator in protein degradation. *FEBS Letters*. 2017; 591: 2648–2660.
- [28] Kohler V, Andréasson C. Hsp70-mediated quality control: should I stay or should I go? *Biological Chemistry*. 2020; 401: 1233–1248.
- [29] Mayer MP. Gymnastics of molecular chaperones. *Molecular Cell*. 2010; 39: 321–331.
- [30] Evgen'ev M, Scheinker V, Levin A. Molecular mechanisms of adaptation to hyperthermia in eukaryotic organisms. I. Heat shock protein synthesis pattern in cell cultures and caterpillars of two silk worm species. *Molecular Biology*. 1987; 21: 484–494.
- [31] Lyashko VN, Vikulova VK, Chernicov VG, Ivanov VI, Ulmasov KA, Zatschina OG, *et al.* Comparison of the heat shock response in ethnically and ecologically different human populations. *Proceedings of the National Academy of Sciences of the United States of America*. 1994; 91: 12492–12495.
- [32] Ulmasov HA, Karaev KK, Lyashko VN, Evgen'ev MB. Heat-shock response in camel (*Camelus dromedarius*) blood cells and adaptation to hyperthermia. *Comparative Biochemistry and Physiology. B, Comparative Biochemistry*. 1993; 106: 867–872.
- [33] Gehring WJ, Wehner R. Heat shock protein synthesis and thermotolerance in *Cataglyphis*, an ant from the Sahara desert. *Proceedings of the National Academy of Sciences of the United States of America*. 1995; 92: 2994–2998.
- [34] Tomanek L, Somero G. Evolutionary and acclimation-induced variation in the heat-shock responses of congeneric marine snails (genus *Tegula*) from different thermal habitats: implications for limits of thermotolerance and biogeography. *The Journal of Experimental Biology*. 1999; 202: 2925–2936.
- [35] Krebs RA, Feder ME. Natural Variation In The Expression of The Heat-Shock Protein Hsp70 In A Population of *Drosophila melanogaster* And Its Correlation With Tolerance of Ecologically Relevant Thermal Stress. *Evolution*. 1997; 51: 173–179.
- [36] Loeschcke V, Krebs RA, Dahlgaard J, Michalak P. High-temperature stress and the evolution of thermal resistance in *Drosophila*. *EXS*. 1997; 83: 175–190.
- [37] Tomanek L. Variation in the heat shock response and its implication for predicting the effect of global climate change on species' biogeographical distribution ranges and metabolic costs. *The Journal of Experimental Biology*. 2010; 213: 971–979.
- [38] Sørensen JG, Kristensen TN, Loeschcke V. The evolutionary and ecological role of heat shock proteins. *Ecology letters*. 2003; 6: 1025–1037.
- [39] Sørensen JG. Application of heat shock protein expression for detecting natural adaptation and exposure to stress in natural populations. *Current Zoology*. 2010; 56: 703–713.
- [40] Oksala NKJ, Ekmekçi FG, Özsoy E, Kirankaya S, Kokkola T, Emecen G, *et al.* Natural thermal adaptation increases heat shock protein levels and decreases oxidative stress. *Redox Biology*. 2014; 3: 25–28.
- [41] Archana P, Aleena J, Pragna P, Vidya M, Niyas A, Bagath M, *et al.* Role of heat shock proteins in livestock adaptation to heat stress. *Journal of Dairy, Veterinary & Animal Research*. 2017; 5: 00127.
- [42] Shatilina ZM, Bedulina D, Protopopova M, Pavlichenko V, Pobezhimova T, Grabelnykh O, *et al.* Heat shock proteins in the mechanisms of stress adaptation in Baikal amphipods and Palaeartic *Gammarus lacustris* Sars II. Small HSP family. *Contemporary Problems of Ecology*. 2010; 3: 449–456.
- [43] Haslbeck M, Vierling E. A first line of stress defense: small heat shock proteins and their function in protein homeostasis. *Journal of Molecular Biology*. 2015; 427: 1537–1548.
- [44] Morrow G, Le Pécheur M, Tanguay RM. *Drosophila melanogaster* mitochondrial Hsp22: a role in resistance to oxidative stress, aging and the mitochondrial unfolding protein response. *Biogerontology*. 2016; 17: 61–70.
- [45] Sarge KD, Murphy SP, Morimoto RI. Activation of heat shock gene transcription by heat shock factor 1 involves oligomerization, acquisition of DNA-binding activity, and nuclear localization and can occur in the absence of stress. *Molecular and Cellular Biology*. 1993; 13: 1392–1407.
- [46] Pirkkala L, Nykänen P, Sistonen L. Roles of the heat shock transcription factors in regulation of the heat shock response and beyond. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2001; 15: 1118–1131.
- [47] Fernandes M. Structure and regulation of heat shock gene promoters. The biology of heat shock proteins and molecular chaperones. 1994; 375–393.
- [48] Morimoto RI. Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones, and negative regulators. *Genes & Development*. 1998; 12: 3788–3796.
- [49] Guisbert E, Czyz DM, Richter K, McMullen PD, Morimoto RI. Identification of a tissue-selective heat shock response regulatory network. *PLoS Genetics*. 2013; 9: e1003466.
- [50] Neef DW, Jaeger AM, Gomez-Pastor R, Willmund F, Frydman J, Thiele DJ. A direct regulatory interaction between chaperonin TRiC and stress-responsive transcription factor HSF1. *Cell Reports*. 2014; 9: 955–966.
- [51] Ma J, Grant CE, Plagens RN, Barrett LN, Kim Guisbert KS, Guisbert E. Cellular Proteomes Drive Tissue-Specific Regulation of the Heat Shock Response. *G3 (Bethesda, Md.)*. 2017; 7: 1011–1018.
- [52] Bettencourt BR, Feder ME. Hsp70 duplication in the *Drosophila melanogaster* species group: how and when did two become five? *Molecular Biology and Evolution*. 2001; 18: 1272–1282.

- [53] Gong WJ, Golic KG. Genomic deletions of the *Drosophila melanogaster* Hsp70 genes. *Genetics*. 2004; 168: 1467–1476.
- [54] Brocchieri L, Conway de Macario E, Macario AJL. hsp70 genes in the human genome: Conservation and differentiation patterns predict a wide array of overlapping and specialized functions. *BMC Evolutionary Biology*. 2008; 8: 19.
- [55] Kampinga HH, Hageman J, Vos MJ, Kubota H, Tanguay RM, Bruford EA, *et al.* Guidelines for the nomenclature of the human heat shock proteins. *Cell Stress & Chaperones*. 2009; 14: 105–111.
- [56] Daugaard M, Rohde M, Jäättelä M. The heat shock protein 70 family: Highly homologous proteins with overlapping and distinct functions. *FEBS Letters*. 2007; 581: 3702–3710.
- [57] Hunt C, Morimoto RI. Conserved features of eukaryotic hsp70 genes revealed by comparison with the nucleotide sequence of human hsp70. *Proceedings of the National Academy of Sciences of the United States of America*. 1985; 82: 6455–6459.
- [58] Evgen'ev MB, Zatssepina OG, Garbuz D, Lerman DN, Velikodvorskaya V, Zelentsova E, *et al.* Evolution and arrangement of the hsp70 gene cluster in two closely related species of the virilis group of *Drosophila*. *Chromosoma*. 2004; 113: 223–232.
- [59] Ish-Horowicz D, Burke JF. Rapid and efficient cosmid cloning. *Nucleic Acids Research*. 1981; 9: 2989–2998.
- [60] Garbuz DG, Yushenova IA, Zatssepina OG, Przhiboro AA, Bettencourt BR, Evgen'ev MB. Organization and evolution of hsp70 clusters strikingly differ in two species of Stratiomyidae (Diptera) inhabiting thermally contrasting environments. *BMC Evolutionary Biology*. 2011; 11: 74.
- [61] Karpov VL, Preobrazhenskaya OV, Mirzabekov AD. Chromatin structure of hsp 70 genes, activated by heat shock: selective removal of histones from the coding region and their absence from the 5' region. *Cell*. 1984; 36: 423–431.
- [62] Wu C. The 5' ends of *Drosophila* heat shock genes in chromatin are hypersensitive to DNase I. *Nature*. 1980; 286: 854–860.
- [63] Costlow N, Lis JT. High-resolution mapping of DNase I-hypersensitive sites of *Drosophila* heat shock genes in *Drosophila melanogaster* and *Saccharomyces cerevisiae*. *Molecular and Cellular Biology*. 1984; 4: 1853–1863.
- [64] Adelman K, Lis JT. Promoter-proximal pausing of RNA polymerase II: emerging roles in metazoans. *Nature Reviews. Genetics*. 2012; 13: 720–731.
- [65] Lerman DN, Michalak P, Helin AB, Bettencourt BR, Feder ME. Modification of heat-shock gene expression in *Drosophila melanogaster* populations via transposable elements. *Molecular Biology and Evolution*. 2003; 20: 135–144.
- [66] Shilova VY, Garbuz DG, Myasyankina EN, Chen B, Evgen'ev MB, Feder ME, *et al.* Remarkable site specificity of local transposition into the Hsp70 promoter of *Drosophila melanogaster*. *Genetics*. 2006; 173: 809–820.
- [67] Hoter A, Amiri M, Prince A, Amer H, Warda M, Naim HY. Differential Glycosylation and Modulation of Camel and Human HSP Isoforms in Response to Thermal and Hypoxic Stresses. *International Journal of Molecular Sciences*. 2018; 19: 402.
- [68] Astakhova LN, Zatssepina OG, Funikov SY, Zelentsova ES, Shostak NG, Orishchenko KE, *et al.* Activity of heat shock genes' promoters in thermally contrasting animal species. *PLoS ONE*. 2015; 10: e0115536.
- [69] Bedulina DS, Evgen'ev MB, Timofeyev MA, Protopopova MV, Garbuz DG, Pavlichenko VV, *et al.* Expression patterns and organization of the hsp70 genes correlate with thermotolerance in two congener endemic amphipod species (*Eulimnogammarus cyaneus* and *E. verrucosus*) from Lake Baikal. *Molecular Ecology*. 2013; 22: 1416–1430.
- [70] Garbuz DG, Zatssepina OG, Przhiboro AA, Yushenova I, Guzhova IV, Evgen'ev MB. Larvae of related Diptera species from thermally contrasting habitats exhibit continuous up-regulation of heat shock proteins and high thermotolerance. *Molecular Ecology*. 2008; 17: 4763–4777.
- [71] Ulmasov KA, Shammakov S, Karaev K, Evgen'ev MB. Heat shock proteins and thermoresistance in lizards. *Proceedings of the National Academy of Sciences of the United States of America*. 1992; 89: 1666–1670.
- [72] Garbuz DG, Evgen'ev MB. The evolution of heat shock genes and expression patterns of heat shock proteins in the species from temperature contrasting habitats. *Genetika*. 2017; 53: 12–30.
- [73] Barua D, Heckathorn SA. Acclimation of the temperature set-points of the heat-shock response. *Journal of Thermal Biology*. 2004; 29: 185–193.
- [74] Fader SC, Yu Z, Spotila JR. Seasonal variation in heat shock proteins (hsp 70) in stream fish under natural conditions. *Journal of Thermal Biology*. 1994; 19: 335–341.
- [75] Hofmann G, Somero G. Evidence for protein damage at environmental temperatures: seasonal changes in levels of ubiquitin conjugates and hsp70 in the intertidal mussel *Mytilus trossulus*. *The Journal of Experimental Biology*. 1995; 198: 1509–1518.
- [76] Pauli D, Arrigo AP, Tissières A. Heat shock response in *Drosophila*. *Experientia*. 1992; 48: 623–629.
- [77] Garbuz D, Evgen'ev MB, Feder ME, Zatssepina OG. Evolution of thermotolerance and the heat-shock response: evidence from inter/intraspecific comparison and interspecific hybridization in the virilis species group of *Drosophila*. I. Thermal phenotype. *The Journal of Experimental Biology*. 2003; 206: 2399–2408.
- [78] Sørensen JG, Giribets MP, Tarrío R, Rodríguez-Trelles F, Schou MF, Loeschke V. Expression of thermal tolerance genes in two *Drosophila* species with different acclimation capacities. *Journal of Thermal Biology*. 2019; 84: 200–207.
- [79] Gong WJ, Golic KG. Loss of Hsp70 in *Drosophila* is pleiotropic, with effects on thermotolerance, recovery from heat shock and neurodegeneration. *Genetics*. 2006; 172: 275–286.
- [80] Zatssepina OG, Evgen'ev MB, Garbuz DG. Role of a Heat Shock Transcription Factor and the Major Heat Shock Protein Hsp70 in Memory Formation and Neuroprotection. *Cells*. 2021; 10: 1638.
- [81] Gorenskaya O, Gavrilov A, Zatssepina O, Shchokorbatov YG, Evgen'ev M. The Role of Hsp70 Genes in Promoting Control of Viability in *Drosophila melanogaster* Subjected to Microwave Irradiation. *Biophysics*. 2021; 66: 541–549.
- [82] Brennecke T, Gellner K, Bosch TC. The lack of a stress response in *Hydra oligactis* is due to reduced hsp70 mRNA stability. *European Journal of Biochemistry*. 1998; 255: 703–709.
- [83] Clark MS, Peck LS. HSP70 heat shock proteins and environmental stress in Antarctic marine organisms: A mini-review. *Marine Genomics*. 2009; 2: 11–18.
- [84] Buckley BA, Somero GN. cDNA microarray analysis reveals the capacity of the cold-adapted Antarctic fish *Trematomus bernacchii* to alter gene expression in response to heat stress. *Polar Biology*. 2009; 32: 403–415.
- [85] Zatssepina OG, Przhiboro AA, Yushenova IA, Shilova V, Zelentsova ES, Shostak NG, *et al.* A *Drosophila* heat shock response represents an exception rather than a rule amongst Diptera species. *Insect Molecular Biology*. 2016; 25: 431–449.
- [86] Zatssepina OG, Velikodvorskaia VV, Molodtsov VB, Garbuz D, Lerman DN, Bettencourt BR, *et al.* A *Drosophila melanogaster* strain from sub-equatorial Africa has exceptional thermotolerance but decreased Hsp70 expression. *The Journal of Experimental Biology*. 2001; 204: 1869–1881.
- [87] Bettencourt BR, Feder ME, Cavicchi S. Experimental Evolution of Hsp70 Expression and Thermotolerance In *Drosophila melanogaster*. *Evolution; International Journal of Organic Evolution*. 1999; 53: 484–492.
- [88] Bogan SN, Place SP. Accelerated evolution at chaperone promoters among Antarctic notothenioid fishes. *BMC Evolutionary*

Biology. 2019; 19: 205.

- [89] Krebs RA, Feder ME. Deleterious consequences of Hsp70 overexpression in *Drosophila melanogaster* larvae. *Cell Stress & Chaperones*. 1997; 2: 60–71.
- [90] Fields PA. Review: Protein function at thermal extremes: balancing stability and flexibility. *Comparative Biochemistry and Physiology. Part A, Molecular & Integrative Physiology*. 2001; 129: 417–431.
- [91] Garbuz DG, Sverchinsky D, Davletshin A, Margulis BA, Mitkevich V, Kulikov AM, *et al.* The molecular chaperone Hsp70 from the thermotolerant Diptera species differs from the *Drosophila* paralog in its thermostability and higher refolding capacity at extreme temperatures. *Cell Stress & Chaperones*. 2019; 24: 1163–1173.
- [92] Shilova VY, Zatssepina OG, Garbuz DG, Funikov SY, Zelenkova ES, Schostak NG, *et al.* Heat shock protein 70 from a thermotolerant Diptera species provides higher thermoresistance to *Drosophila* larvae than correspondent endogenous gene. *Insect Molecular Biology*. 2018; 27: 61–72.
- [93] De Maio A, Hightower L. The interaction of heat shock proteins with cellular membranes: a historical perspective. *Cell Stress & Chaperones*. 2021; 26: 769–783.
- [94] Guidon PT, Hightower LE. Purification and initial characterization of the 71-kilodalton rat heat-shock protein and its cognate as fatty acid binding proteins. *Biochemistry*. 1986; 25: 3231–3239.
- [95] Hightower LE, Guidon PT. Selective release from cultured mammalian cells of heat-shock (stress) proteins that resemble glia-axon transfer proteins. *Journal of Cellular Physiology*. 1989; 138: 257–266.
- [96] Moseley PL. Heat shock proteins and heat adaptation of the whole organism. *Journal of Applied Physiology* (Bethesda, Md.: 1985). 1997; 83: 1413–1417.
- [97] Lim CL, Suzuki K. Systemic inflammation mediates the effects of endotoxemia in the mechanisms of heat stroke. *Biology and Medicine*. 2017; 9: 1.
- [98] Lambert GP, Gisolfi CV, Berg DJ, Moseley PL, Oberley LW, Kregel KC. Selected contribution: Hyperthermia-induced intestinal permeability and the role of oxidative and nitrosative stress. *Journal of Applied Physiology* (Bethesda, Md.: 1985). 2002; 92: 1750–61; discussion 1749.
- [99] Yeh YJ, Law LYL, Lim CL. Gastrointestinal response and endotoxemia during intense exercise in hot and cool environments. *European Journal of Applied Physiology*. 2013; 113: 1575–1583.
- [100] Musch MW, Ciancio MJ, Sarge K, Chang EB. Induction of heat shock protein 70 protects intestinal epithelial IEC-18 cells from oxidant and thermal injury. *The American Journal of Physiology*. 1996; 270: C429–C436.
- [101] Dokladny K, Wharton W, Lobb R, Ma TY, Moseley PL. Induction of physiological thermotolerance in MDCK monolayers: contribution of heat shock protein 70. *Cell Stress & Chaperones*. 2006; 11: 268–275.
- [102] Zuhl MN, Lanphere KR, Kravitz L, Mermier CM, Schneider S, Dokladny K, *et al.* Effects of oral glutamine supplementation on exercise-induced gastrointestinal permeability and tight junction protein expression. *Journal of Applied Physiology* (Bethesda, Md.: 1985). 2014; 116: 183–191.
- [103] Kuennen M, Gillum T, Dokladny K, Bedrick E, Schneider S, Moseley P. Thermotolerance and heat acclimation may share a common mechanism in humans. *American Journal of Physiology. Regulatory, Integrative and Comparative Physiology*. 2011; 301: R524–R533.
- [104] Pockley AG, Calderwood SK, Multhoff G. The atheroprotective properties of Hsp70: a role for Hsp70-endothelial interactions? *Cell Stress & Chaperones*. 2009; 14: 545–553.
- [105] Yuan X, Chen Y, Chen G, Liu G, Hang M, Wang P, *et al.* The Heat Shock Protein 70 Plays a Protective Role in Sepsis by Maintenance of the Endothelial Permeability. *BioMed Research International*. 2020; 2020: 2194090.
- [106] Arispe N, Doh M, Simakova O, Kurganov B, De Maio A. Hsc70 and Hsp70 interact with phosphatidylserine on the surface of PC12 cells resulting in a decrease of viability. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2004; 18: 1636–1645.
- [107] Armijo G, Okerblom J, Cauvi DM, Lopez V, Schlamadinger DE, Kim J, *et al.* Interaction of heat shock protein 70 with membranes depends on the lipid environment. *Cell Stress & Chaperones*. 2014; 19: 877–886.
- [108] Lopez V, Cauvi DM, Arispe N, De Maio A. Bacterial Hsp70 (DnaK) and mammalian Hsp70 interact differently with lipid membranes. *Cell Stress & Chaperones*. 2016; 21: 609–616.
- [109] Schilling D, Gehrmann M, Steinem C, De Maio A, Pockley AG, Abend M, *et al.* Binding of heat shock protein 70 to extracellular phosphatidylserine promotes killing of normoxic and hypoxic tumor cells. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2009; 23: 2467–2477.
- [110] McCallister C, Kdeiss B, Nikolaidis N. Biochemical characterization of the interaction between HspA1A and phospholipids. *Cell Stress & Chaperones*. 2016; 21: 41–53.
- [111] Arispe N, De Maio A. ATP and ADP modulate a cation channel formed by Hsc70 in acidic phospholipid membranes. *The Journal of Biological Chemistry*. 2000; 275: 30839–30843.
- [112] Macazo FC, White RJ. Monitoring charge flux to quantify unusual ligand-induced ion channel activity for use in biological nanopore-based sensors. *Analytical Chemistry*. 2014; 86: 5519–5525.
- [113] Kirkegaard T, Roth AG, Petersen NHT, Mahalka AK, Olsen OD, Moilanen I, *et al.* Hsp70 stabilizes lysosomes and reverts Niemann-Pick disease-associated lysosomal pathology. *Nature*. 2010; 463: 549–553.
- [114] Mahalka AK, Kirkegaard T, Jukola LTI, Jäättelä M, Kinnunen PKJ. Human heat shock protein 70 (Hsp70) as a peripheral membrane protein. *Biochimica Et Biophysica Acta*. 2014; 1838: 1344–1361.
- [115] Nylandsted J, Gyrd-Hansen M, Danielewicz A, Fehrenbacher N, Lademann U, Høyer-Hansen M, *et al.* Heat shock protein 70 promotes cell survival by inhibiting lysosomal membrane permeabilization. *The Journal of Experimental Medicine*. 2004; 200: 425–435.
- [116] Horváth I, Multhoff G, Sonnleitner A, Vigh L. Membrane-associated stress proteins: more than simply chaperones. *Biochimica Et Biophysica Acta*. 2008; 1778: 1653–1664.
- [117] Balogi Z, Multhoff G, Jensen TK, Lloyd-Evans E, Yamashita T, Jäättelä M, *et al.* Hsp70 interactions with membrane lipids regulate cellular functions in health and disease. *Progress in Lipid Research*. 2019; 74: 18–30.
- [118] Török Z, Crul T, Maresca B, Schütz GJ, Viana F, Dindia L, *et al.* Plasma membranes as heat stress sensors: from lipid-controlled molecular switches to therapeutic applications. *Biochimica Et Biophysica Acta*. 2014; 1838: 1594–1618.
- [119] Triantafilou M, Miyake K, Golenbock DT, Triantafilou K. Mediators of innate immune recognition of bacteria concentrate in lipid rafts and facilitate lipopolysaccharide-induced cell activation. *Journal of Cell Science*. 2002; 115: 2603–2611.
- [120] Chen S, Bawa D, Besshoh S, Gurd JW, Brown IR. Association of heat shock proteins and neuronal membrane components with lipid rafts from the rat brain. *Journal of Neuroscience Research*. 2005; 81: 522–529.
- [121] Gastpar R, Gehrmann M, Bausero MA, Asea A, Gross C, Schroeder JA, *et al.* Heat shock protein 70 surface-positive tumor exosomes stimulate migratory and cytolytic activity of nat-

- ural killer cells. *Cancer Research*. 2005; 65: 5238–5247.
- [122] Mambula SS, Calderwood SK. Heat shock protein 70 is secreted from tumor cells by a nonclassical pathway involving lysosomal endosomes. *Journal of Immunology* (Baltimore, Md.: 1950). 2006; 177: 7849–7857.
- [123] Broquet AH, Thomas G, Masliah J, Trugnan G, Bachelet M. Expression of the molecular chaperone Hsp70 in detergent-resistant microdomains correlates with its membrane delivery and release. *The Journal of Biological Chemistry*. 2003; 278: 21601–21606.
- [124] Srivastava P. Interaction of heat shock proteins with peptides and antigen presenting cells: chaperoning of the innate and adaptive immune responses. *Annual Review of Immunology*. 2002; 20: 395–425.
- [125] Takemoto S, Nishikawa M, Takakura Y. Pharmacokinetic and tissue distribution mechanism of mouse recombinant heat shock protein 70 in mice. *Pharmaceutical Research*. 2005; 22: 419–426.
- [126] Hartl FU, Bracher A, Hayer-Hartl M. Molecular chaperones in protein folding and proteostasis. *Nature*. 2011; 475: 324–332.
- [127] Garbuz DG, Zatsepina OG, Evgen'ev MB. The Major Human Stress Protein Hsp70 as a Factor of Protein Homeostasis and a Cytokine-Like Regulator. *Molekuliarnaia Biologiya*. 2019; 53: 200–217.
- [128] de Oliveira AA, Mendoza VO, Priviero F, Webb RC, Nunes KP. Age-Related Decline in Vascular Responses to Phenylephrine Is Associated with Reduced Levels of HSP70. *Biomolecules*. 2022; 12: 1125.
- [129] Peinado-Ruiz IC, Burgos-Molina AM, Sendra-Portero F, Ruiz-Gómez MJ. Relationship between heat shock proteins and cellular resistance to drugs and ageing. *Experimental Gerontology*. 2022; 167: 111896.
- [130] Morimoto RI. Proteotoxic stress and inducible chaperone networks in neurodegenerative disease and aging. *Genes & Development*. 2008; 22: 1427–1438.
- [131] Murshid A, Eguchi T, Calderwood SK. Stress proteins in aging and life span. *International Journal of Hyperthermia: the Official Journal of European Society for Hyperthermic Oncology, North American Hyperthermia Group*. 2013; 29: 442–447.
- [132] Singh R, Kølvrå S, Bross P, Jensen UB, Gregersen N, Tan Q, *et al.* Reduced heat shock response in human mononuclear cells during aging and its association with polymorphisms in HSP70 genes. *Cell Stress & Chaperones*. 2006; 11: 208–215.
- [133] Sellares J, Veraldi KL, Thiel KJ, Cárdenes N, Alvarez D, Schneider F, *et al.* Intracellular Heat Shock Protein 70 Deficiency in Pulmonary Fibrosis. *American Journal of Respiratory Cell and Molecular Biology*. 2019; 60: 629–636.
- [134] Yadav R, Devi SS, Oswalia J, Ramalingam S, Arya R. Role of HSP70 chaperone in protein aggregate phenomenon of GNE mutant cells: Therapeutic lead for GNE Myopathy. *The International Journal of Biochemistry & Cell Biology*. 2022; 149: 106258.
- [135] Hsiao C, Lee C, Yang R, Chen J, Su T, Chang Y, *et al.* Heat Shock Protein-70 Levels Are Associated With a State of Oxidative Damage in the Development of Bronchopulmonary Dysplasia. *Frontiers in Pediatrics*. 2021; 9: 616452.
- [136] Rodrigues-Krause J, Krause M, O'Hagan C, De Vito G, Boreham C, Murphy C, *et al.* Divergence of intracellular and extracellular HSP72 in type 2 diabetes: does fat matter? *Cell Stress & Chaperones*. 2012; 17: 293–302.
- [137] Mulyani WRW, Sanjiwani MID, Sandra, Prabawa IPY, Lestari AAW, Wihandani DM, *et al.* Chaperone-Based Therapeutic Target Innovation: Heat Shock Protein 70 (HSP70) for Type 2 Diabetes Mellitus. *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*. 2020; 13: 559–568.
- [138] Heck TG, Ludwig MS, Frizzo MN, Rasia-Filho AA, Homem de Bittencourt PI. Suppressed anti-inflammatory heat shock response in high-risk COVID-19 patients: lessons from basic research (inclusive bats), light on conceivable therapies. *Clinical Science* (London, England: 1979). 2020; 134: 1991–2017.
- [139] Wang JC, Bennett M. Aging and atherosclerosis: mechanisms, functional consequences, and potential therapeutics for cellular senescence. *Circulation Research*. 2012; 111: 245–259.
- [140] Hu C, Yang J, Qi Z, Wu H, Wang B, Zou F, *et al.* Heat shock proteins: Biological functions, pathological roles, and therapeutic opportunities. *MedComm*. 2022; 3: e161.
- [141] Fu X, Liu H, Liu J, DiSanto ME, Zhang X. The Role of Heat Shock Protein 70 Subfamily in the Hyperplastic Prostate: From Molecular Mechanisms to Therapeutic Opportunities. *Cells*. 2022; 11: 2052.
- [142] Aghazadeh N, Beilankouhi EAV, Fakhri F, Gargari MK, Bahari P, Moghadami A, *et al.* Involvement of heat shock proteins and parkin/ α -synuclein axis in Parkinson's disease. *Molecular Biology Reports*. 2022; 49: 11061–11070.
- [143] Calderwood SK, Murshid A. Molecular Chaperone Accumulation in Cancer and Decrease in Alzheimer's Disease: The Potential Roles of HSF1. *Frontiers in Neuroscience*. 2017; 11: 192.
- [144] Leak RK. Heat shock proteins in neurodegenerative disorders and aging. *Journal of Cell Communication and Signaling*. 2014; 8: 293–310.
- [145] Dong Y, Li T, Ma Z, Zhou C, Wang X, Li J. HSPA1A, HSPA2, and HSPA8 Are Potential Molecular Biomarkers for Prognosis among HSP70 Family in Alzheimer's Disease. *Disease Markers*. 2022; 2022: 9480398.
- [146] Albakova Z, Siam MKS, Sacitharan PK, Ziganshin RH, Ryazantsev DY, Sapozhnikov AM. Extracellular heat shock proteins and cancer: New perspectives. *Translational Oncology*. 2021; 14: 100995.
- [147] Vostakolaie MA, Hatami-Baroogh L, Babaei G, Molavi O, Kord S, Abdolalizadeh J. Hsp70 in cancer: A double agent in the battle between survival and death. *Journal of Cellular Physiology*. 2021; 236: 3420–3444.
- [148] De Maio A, Hightower LE. Heat shock proteins and the biogenesis of cellular membranes. *Cell Stress & Chaperones*. 2021; 26: 15–18.
- [149] Kami K, Ohira T, Oishi Y, Nakajima T, Goto K, Ohira Y. Role of 72-kDa Heat Shock Protein in Heat-stimulated Regeneration of Injured Muscle in Rat. *The Journal of Histochemistry and Cytochemistry: Official Journal of the Histochemistry Society*. 2019; 67: 791–799.
- [150] Kmiecik SW, Mayer MP. Molecular mechanisms of heat shock factor 1 regulation. *Trends in Biochemical Sciences*. 2022; 47: 218–234.
- [151] Rébé C, Ghiringhelli F, Garrido C. Can the hyperthermia-mediated heat shock factor/heat shock protein 70 pathway dampen the cytokine storm during SARS-CoV-2 infection? *British Journal of Pharmacology*. 2022; 179: 4910–4916.
- [152] Kitchen LC, Berman M, Halper J, Chazot P. Rationale for 1068 nm Photobiomodulation Therapy (PBMT) as a Novel, Non-Invasive Treatment for COVID-19 and Other Coronaviruses: Roles of NO and Hsp70. *International Journal of Molecular Sciences*. 2022; 23: 5221.
- [153] Lazarev VF, Dutsheva EA, Komarova EY, Mikhaylova ER, Guzhova IV, Margulis BA. GAPDH-targeted therapy - A new approach for secondary damage after traumatic brain injury on rats. *Biochemical and Biophysical Research Communications*. 2018; 501: 1003–1008.
- [154] Li F, Gong X, Yang B. Geranylgeranylacetone ameliorated ischemia/reperfusion induced-blood brain barrier breakdown through HSP70-dependent anti-apoptosis effect. *American Journal of Translational Research*. 2021; 13: 102–114.
- [155] Zeng H, Yang L, Zhang X, Chen Y, Cai J. Dioscin pre-

- vents LPS-induced acute lung injury through inhibiting the TLR4/MyD88 signaling pathway via upregulation of HSP70. *Molecular Medicine Reports*. 2018; 17: 6752–6758.
- [156] Panossian A, Wikman G, Kaur P, Asea A. Adaptogens exert a stress-protective effect by modulation of expression of molecular chaperones. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2009; 16: 617–622.
- [157] Panossian A, Seo E, Efferth T. Novel molecular mechanisms for the adaptogenic effects of herbal extracts on isolated brain cells using systems biology. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2018; 50: 257–284.
- [158] Yang Y, Sun Y, Zhang N, Li J, Zhang C, Duan X, *et al.* The up-regulation of two identified wound healing specific proteins-HSP70 and lysozyme in regenerated *Eisenia fetida* through transcriptome analysis. *Journal of Ethnopharmacology*. 2019; 237: 64–73.
- [159] Rouhollahi E, Moghadamtousi SZ, Hajiaghaalipour F, Zahedifard M, Tayeby F, Awang K, *et al.* Curcuma purpurascens BI. rhizome accelerates rat excisional wound healing: involvement of Hsp70/Bax proteins, antioxidant defense, and angiogenesis activity. *Drug Design, Development and Therapy*. 2015; 9: 5805–5813.
- [160] Boswell-Casteel RC, Johnson JM, Duggan KD, Tsutsui Y, Hays FA. Overproduction and biophysical characterization of human HSP70 proteins. *Protein Expression and Purification*. 2015; 106: 57–65.
- [161] Tytell M, Davis AT, Giles J, Snider LC, Xiao R, Dozier SG, *et al.* Alfalfa-derived HSP70 administered intranasally improves insulin sensitivity in mice. *Cell Stress & Chaperones*. 2018; 23: 189–194.
- [162] Kirkegaard T, Gray J, Priestman DA, Wallom K, Atkins J, Olsen OD, *et al.* Heat shock protein-based therapy as a potential candidate for treating the sphingolipidoses. *Science Translational Medicine*. 2016; 8: 355ra118.
- [163] Gurskiy YG, Garbuz DG, Soshnikova NV, Krasnov AN, Deikin A, Lazarev VF, *et al.* The development of modified human Hsp70 (HSPA1A) and its production in the milk of transgenic mice. *Cell Stress & Chaperones*. 2016; 21: 1055–1064.
- [164] Evgen'ev MB, Krasnov GS, Nesterova IV, Garbuz DG, Karpov VL, Morozov AV, *et al.* Molecular Mechanisms Underlying Neuroprotective Effect of Intranasal Administration of Human Hsp70 in Mouse Model of Alzheimer's Disease. *Journal of Alzheimer's Disease: JAD*. 2017; 59: 1415–1426.
- [165] Ekimova IV, Nitsinskaya LE, Romanova IV, Pastukhov YF, Margulis BA, Guzhova IV. Exogenous protein Hsp70/Hsc70 can penetrate into brain structures and attenuate the severity of chemically-induced seizures. *Journal of Neurochemistry*. 2010; 115: 1035–1044.
- [166] Kustanova GA, Murashev AN, Karpov VL, Margulis BA, Guzhova IV, Prokhorenko IR, *et al.* Exogenous heat shock protein 70 mediates sepsis manifestations and decreases the mortality rate in rats. *Cell Stress & Chaperones*. 2006; 11: 276–286.
- [167] Sulzbacher MM, Sulzbacher LM, Passos FR, Bilibio BLE, Althaus WF, Weizenmann L, *et al.* A single dose of eHSP72 attenuates sepsis severity in mice. *Scientific Reports*. 2020; 10: 9198.
- [168] Bobkova NV, Evgen'ev M, Garbuz DG, Kulikov AM, Morozov A, Samokhin A, *et al.* Exogenous Hsp70 delays senescence and improves cognitive function in aging mice. *Proceedings of the National Academy of Sciences of the United States of America*. 2015; 112: 16006–16011.
- [169] Demyanenko S, Nikul V, Rodkin S, Davletshin A, Evgen'ev MB, Garbuz DG. Exogenous recombinant Hsp70 mediates neuroprotection after photothrombotic stroke. *Cell Stress & Chaperones*. 2021; 26: 103–114.
- [170] Zhang X, Zhang X, Huang W, Ge X. The role of heat shock proteins in the regulation of fibrotic diseases. *Biomedicine & Pharmacotherapy*. 2021; 135: 111067.
- [171] Gungor B, Vanharanta L, Hölttä-Vuori M, Pirhonen J, Petersen NHT, Gramolelli S, *et al.* HSP70 induces liver X receptor pathway activation and cholesterol reduction *in vitro* and *in vivo*. *Molecular Metabolism*. 2019; 28: 135–143.
- [172] Pei Q, Ni W, Yuan Y, Yuan J, Zhang X, Yao M. HSP70 Ameliorates Septic Lung Injury via Inhibition of Apoptosis by Interacting with KANK2. *Biomolecules*. 2022; 12: 410.
- [173] Abkin SV, Ostroumova OS, Komarova EY, Meshalkina DA, Shevtsov MA, Margulis BA, *et al.* Phloretin increases the anti-tumor efficacy of intratumorally delivered heat-shock protein 70 kDa (HSP70) in a murine model of melanoma. *Cancer Immunology, Immunotherapy: CII*. 2016; 65: 83–92.
- [174] Komarova EY, Suezov RV, Nikotina AD, Aksenov ND, Garaeva LA, Shtam TA, *et al.* Hsp70-containing extracellular vesicles are capable of activating of adaptive immunity in models of mouse melanoma and colon carcinoma. *Scientific Reports*. 2021; 11: 21314.
- [175] Shevtsov MA, Komarova EY, Meshalkina DA, Bychkova NV, Aksenov ND, Abkin SV, *et al.* Exogenously delivered heat shock protein 70 displaces its endogenous analogue and sensitizes cancer cells to lymphocytes-mediated cytotoxicity. *Oncotarget*. 2014; 5: 3101–3114.
- [176] Shevtsov MA, Kim AV, Samochernych KA, Romanova IV, Margulis BA, Guzhova IV, *et al.* Pilot study of intratumoral injection of recombinant heat shock protein 70 in the treatment of malignant brain tumors in children. *Oncotargets and Therapy*. 2014; 7: 1071–1081.
- [177] Zhang X, Yu W. Heat shock proteins and viral infection. *Frontiers in Immunology*. 2022; 13: 947789.
- [178] Maeda Y, Yoshimura K, Matsui H, Shindo Y, Tamesa T, Tokumitsu Y, *et al.* Dendritic cells transfected with heat-shock protein 70 messenger RNA for patients with hepatitis C virus-related hepatocellular carcinoma: a phase I dose escalation clinical trial. *Cancer Immunology, Immunotherapy: CII*. 2015; 64: 1047–1056.
- [179] Asea A. Hsp70: a chaperokine. *Novartis Foundation Symposium*. 2008; 291: 173–224.
- [180] Zhang H, Gong W, Wu S, Perrett S. Hsp70 in Redox Homeostasis. *Cells*. 2022; 11: 829.
- [181] Onikienko S, Vinokurov M, Yurinskaya M, Zemlyanoi A, Abkin S, Shaykhutdinova E, *et al.* The Effects of H2S and Recombinant Human Hsp70 on Inflammation Induced by SARS and Other Agents *In Vitro* and *In Vivo*. *Biomedicines*. 2022; 10: 2155.
- [182] Wentink AS, Nilleghoda NB, Feufel J, Ubartaitė G, Schneider CP, De Los Rios P, *et al.* Molecular dissection of amyloid disaggregation by human HSP70. *Nature*. 2020; 587: 483–488.
- [183] Beere HM, Wolf BB, Cain K, Mosser DD, Mahboubi A, Kuwana T, *et al.* Heat-shock protein 70 inhibits apoptosis by preventing recruitment of procaspase-9 to the Apaf-1 apoptosome. *Nature Cell Biology*. 2000; 2: 469–475.
- [184] Mosser DD, Caron AW, Bourget L, Meriin AB, Sherman MY, Morimoto RI, *et al.* The chaperone function of hsp70 is required for protection against stress-induced apoptosis. *Molecular and Cellular Biology*. 2000; 20: 7146–7159.
- [185] Ferguson ID, Lin YT, Lam C, Shao H, Tharp KM, Hale M, *et al.* Allosteric HSP70 inhibitors perturb mitochondrial proteostasis and overcome proteasome inhibitor resistance in multiple myeloma. *Cell Chemical Biology*. 2022; 29: 1288–1302.e7.
- [186] Nikotina AD, Vladimirova SA, Komarova EY, Alexeev D, Efremov S, Leonova E, *et al.* Prevention of High Glucose-Mediated EMT by Inhibition of Hsp70 Chaperone. *International Journal of Molecular Sciences*. 2021; 22: 6902.

- [187] Lubkowska A, Pluta W, Strońska A, Lalko A. Role of Heat Shock Proteins (HSP70 and HSP90) in Viral Infection. *International Journal of Molecular Sciences*. 2021; 22: 9366.
- [188] Bolhassani A, Rafati S. Heat-shock proteins as powerful weapons in vaccine development. *Expert Review of Vaccines*. 2008; 7: 1185–1199.
- [189] McNulty S, Colaco CA, Blandford LE, Bailey CR, Baschieri S, Todryk S. Heat-shock proteins as dendritic cell-targeting vaccines—getting warmer. *Immunology*. 2013; 139: 407–415.
- [190] Dhakal J, Brah GS, Agrawal RK, Pawar HN, Kaur D, Verma R. Over-expression of gene encoding heat shock protein 70 from *Mycobacterium tuberculosis* and its evaluation as vaccine adjuvant. *Indian Journal of Medical Microbiology*. 2013; 31: 123–129.
- [191] Kashiwagi S, Brauns T, Gelfand J, Poznansky MC. Laser vaccine adjuvants. History, progress, and potential. *Human Vaccines & Immunotherapeutics*. 2014; 10: 1892–1907.
- [192] Borges TJ, Wieten L, van Herwijnen MJC, Broere F, van der Zee R, Bonorino C, *et al.* The anti-inflammatory mechanisms of Hsp70. *Frontiers in Immunology*. 2012; 3: 95.
- [193] Evgen'ev M, Bobkova N, Krasnov G, Garbuz D, Funikov S, Kudryavtseva A, *et al.* The Effect of Human HSP70 Administration on a Mouse Model of Alzheimer's Disease Strongly Depends on Transgenicity and Age. *Journal of Alzheimer's Disease: JAD*. 2019; 67: 1391–1404.
- [194] Calvaresi V, Truelsen LT, Larsen SB, Petersen NHT, Kirkegaard T, Rand KD. Conformational dynamics of free and membrane-bound human Hsp70 in model cytosolic and endolysosomal environments. *Communications Biology*. 2021; 4: 1369.
- [195] Ji K, Xue L, Cheng J, Bai Y. Preconditioning of H₂S inhalation protects against cerebral ischemia/reperfusion injury by induction of HSP70 through PI3K/Akt/Nrf2 pathway. *Brain Research Bulletin*. 2016; 121: 68–74.
- [196] Rigo MM, Borges TJ, Lang BJ, Murshid A, Nitika, Wolfgeher D, *et al.* Host expression system modulates recombinant Hsp70 activity through post-translational modifications. *The FEBS Journal*. 2020. (Online ahead of print)