

Review

Understanding Multiple Sclerosis Pathophysiology and Current Disease-Modifying Therapies: A Review of Unaddressed Aspects

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Abstract

Multiple sclerosis (MS) is a complex autoimmune disorder of the central nervous system (CNS) with an unknown etiology and pathophysiology that is not completely understood. Although great strides have been made in developing disease-modifying therapies (DMTs) that have significantly improved the quality of life for MS patients, these treatments do not entirely prevent disease progression or relapse. Identifying the unaddressed pathophysiological aspects of MS and developing targeted therapies to fill in these gaps are essential in providing long-term relief for patients. Recent research has uncovered some aspects of MS that remain outside the scope of available DMTs, and as such, yield only limited benefits. Despite most MS pathophysiology being targeted by DMTs, many patients still experience disease progression or relapse, indicating that a more detailed understanding is necessary. Thus, this literature review seeks to explore the known aspects of MS pathophysiology, identify the gaps in present DMTs, and explain why current treatments cannot entirely arrest MS progression.

Keywords: multiple sclerosis (MS); experimental autoimmune encephalomyelitis; MS therapies; MS pathogenesis; immune system; neurodegenerative disorder

1. Introduction

Multiple sclerosis (MS) is a multifactorial disease that affects the central nervous system (CNS) and is believed to be caused by dysfunction of the immune system [1]. In 2023, more than 2.9 million people worldwide had MS [2].

MS can severely impact neurological function, causing varied symptoms. During remission, some show recovery, but chronic disabilities often become irreversible due to axonal loss [1,3]. There are four main subtypes: relapsing-remitting MS (RRMS) (most common with unpredictable relapses/remissions), primary-progressive MS (PPMS) (steady disability without relapses), secondary-progressive MS (SPMS) (progressive decline after RRMS), and progressive-relapsing MS (PRMS) (disability plus relapses). Clinically isolated syndrome (CIS) involves a first episode of MS-like CNS symptoms [1,3].

Despite ongoing research, the etiology of MS remains unclear. However, multiple factors such as genetics, environmental conditions, and infections are associated with development of the disease [1]. Effective disease treatment hinges on a comprehensive understanding of its pathophysiology. To date, research into the pathophysiology of MS has been extensive; however, a complete understanding of this complex process is lacking [3]. While initial theories suggested that MS resulted from a viral infection of the CNS, subsequent research shifted the perspective towards an autoimmune pathway as the primary cause of the disease [4]. Since then, our understanding of the disease has become much clearer, although more work is still needed to fully elucidate the underlying mechanisms [3].

MS is an incurable disease; however, since the introduction of interferon (IFN) in 1993, there has been a significant increase in the number of treatment options available to MS patients [5]. Currently, there are more than 19 United States Food and Drug Administration (FDA)- and European Medicines Agency (EMA)-approved disease modifying therapies (DMTs) available to treat patients with MS, each targeting specific immunological mechanisms (Tables 1,2,3,4) [6,7]. However, the existence of multiple DMTs for MS treatment, coupled with variations in international guidelines, has led to inconsistent decision-making across clinics, raising concern and requiring attention [7–10].

While current DMTs have significantly enhanced the quality of life for MS patients, unaddressed aspects remain unaddressed for the complete prevention of disease progression or relapse. Thus, developing targeted therapies thus necessitates a thorough understanding of the disease's pathophysiology and the identification of areas that current DMTs fail to address. This review examines the pathophysiology of MS and provides an overview of current DMTs that target each stage of disease development. Furthermore, it sheds light on those aspects of MS pathophysiology that existing DMTs still do not properly address. In doing so, the review explains the gaps and limitations in the studies on the efficacy of DMT for MS. While most steps of MS pathological development are targeted by current DMTs, this review will explain why disease progression and relapse often cannot be halted despite such treatment measures.



2. Overview of Currently Approved DMTs for MS Treatment

Although there is no cure for MS, various DMTs are available that can help reduce clinical and subclinical disease activity, slow disease progression, and delay disease development. Currently, DMTs are approved to treat CIS, RRMS, SPMS, and PPMS [11].

2.1 Currently Approved DMTs

DMTs can be categorized into four groups according to their mechanism of action: monoclonal antibodies such as mitoxantrone, natalizumab, alemtuzumab, daclizumab, ocrelizumab, and ofatumumab; sphingosine 1-phosphate receptor (S1PR) modulators including fingolimod, siponimod, ozanimod, and ponesimod; DNA replication and repair interfering agents such as teriflunomide and cladribine; and agents with an unknown mechanism of action, including interferon-beta (IFN β)-1b, IFN β -1a, dimethyl fumarate (DMF), monomethyl fumarate (MMF), diroximel fumarate (DRF), glatiramer acetate (GA), and pegylated IFN β -1a (Tables 1,2,3,4) [6,11–14].

2.2 DMTs Mechanisms of Action and Limitations

2.2.1 Monoclonal Antibodies

2.2.1.1 Natalizumab. Natalizumab works by blocking the interaction between the α 4 β 1 integrin on lymphocytes and vascular cell adhesion molecule-1 (VCAM1) on endothelial cells, which hinders the movement of immune cells into the CNS [6,11,15]. This blockade results in an increased number of mononuclear cells in the bloodstream and may also prompt the release of hematopoietic precursors from the bone marrow. The consequent rise in lymphocyte levels is a significant outcome of natalizumab therapy [6,11,15].

However, there is an important association between natalizumab use and a heightened risk of progressive multifocal leukoencephalopathy (PML), a rare demyelinating condition triggered by the John Cunningham virus (JCV) [6,11,15]. While JCV is common and generally asymptomatic, it can reactivate in individuals with weakened immune systems [6,11,15]. It has been proposed that the increase in circulating hematopoietic precursors and the buildup of pre-B and B cells due to natalizumab may help JCV transition from its latent state to a more virulent form, which can enhance viral activity and replication within lymphoid tissues [6,11,15]. Key risk factors for developing PML during treatment with natalizumab include prolonged therapy (>2 years), previous immunosuppressive treatments, and the presence of anti-JCV antibodies [6,11,15] (Table 1).

2.2.1.2 Ofatumumab. Ofatumumab is a monoclonal antibody designed to target the cluster of differentiation (CD20) protein found on the surface of B lymphocytes, although its precise mechanism of action remains unclear [6,11,16]. The fragment antigen-binding (Fab) region of ofatumumab specifically attaches to CD20, which leads to a depletion

in B cell populations, and possibly T cell reduction as well, through two primary pathways [16]. One pathway is complement-dependent cytotoxicity (CDC), where the binding of ofatumumab activates the complement system, leading to the destruction of B cells that express CD20. Alternatively, binding can trigger antibody-dependent cell-mediated cytotoxicity (ADCC), in which effector cells, such as natural killer (NK) cells, identify and eliminate the antibody-labeled B cells [16]. This dual mechanism highlights the role of ofatumumab in modulating immune responses by targeting B cells directly [6,11,16] (Table 1).

2.2.1.3 Ocrelizumab. Ocrelizumab is a humanized monoclonal antibody that, similar to ofatumumab, specifically targets the CD20 protein on B cells [6,11,17]. By binding to this protein, ocrelizumab facilitates the depletion of B cells that express CD20 [6,11,17]. However, unlike some therapies that aim to deplete B cells, ocrelizumab is formulated to allow preservation of the body's ability to regenerate B cells and maintain existing humoral immunity [17].

This means that although ocrelizumab effectively reduces the number of circulating B cells, it does not completely eradicate them. This strategy enables the eventual repopulation of B cells and conservation of the body's existing antibody-mediated immune functions. The selective approach to B cell depletion, combined with the maintenance of B cell reconstitution and humoral immunity, is believed to enhance the therapeutic effectiveness and improve the safety profile of ocrelizumab in treating a range of diseases associated with B cell activity [17] (Table 1).

2.2.1.4 Alemtuzumab. Alemtuzumab is a humanized monoclonal immunoglobulin G 1 (IgG1) antibody that targets the cluster of differentiation 52 (CD52) protein [6,11,18,19]. CD52 is a glycosylphosphatidylinositol-anchored protein expressed at high levels on T and B lymphocytes, and to a lesser extent on monocytes, macrophages, and eosinophils. It is minimally expressed on NK cells, plasma cells, neutrophils, and hematopoietic stem cells [18,19].

The binding of alemtuzumab to CD52 leads to the depletion of CD52-positive cells through two main mechanisms: ADCC (in animal models, this appears to be the predominant mechanism) and CDC of CD52-positive cells. The depletion of CD52-high CD4⁺ T cells may be particularly important in the development of secondary autoimmune conditions that can occur after alemtuzumab treatment, as these cells play a crucial role in regulating the immune response [18,19]. The exact physiological function of CD52 is still not fully understood, but it has been implicated in the activation and migration of T lymphocytes, despite the lack of intracellular domains [18,19] (Table 1).

2.2.2 S1PR Modulators

Sphingosine-1-phosphate receptors (S1PRs) are a family of five distinct G protein-coupled cell surface receptors [6,11,20]. Among them, S1P1R plays a crucial role in

Table 1. The currently FDA- and EMA-approved Monoclonal Antibodies-based DMTs for the treatment of MS.

Type	Drug name	Brand name	Mechanism of action	Method of administration	Approval year (FDA)	Approval year (EMA)	Approved for	Contraindications	Efficacy	Adverse effect
Monoclonal antibodies	Mitoxantrone	Novantrone	Synthetic antineoplastic anthracene-dione	Intravenous	2000	1998	SPMS PRMS Worsening RRMS &	Hypersensitivity reactions to mitoxantrone.	Reduces relapses, progression, and lesions in MS.	Myelosuppression, congestive heart failure, AML, serious skin damage (necrosis).
	Natalizumab	Tysabri	anti- α 4 integrin	Intravenous	2004	2006	Relapsing forms of MS	Hypersensitivity reactions to natalizumab, patients who have or had PML, immunocompromised patients, it should not be administered with immunoregulatory agents.	Reduce clinical relapses by 67%, new brain MRI lesions by 83%, and disability progression by 42%.	Bladder/urinary issues, skin reactions, respiratory symptoms, cardiovascular problems, genital/vaginal irritation, swelling (face, eyes, lips), fatigue and weakness.
	Alemtuzumab	Lemtrada	Anti-CD52	Intravenous	2014	2013	Relapsing forms of MS	Hypersensitivity reactions to alemtuzumab, HIV infection, active infection, uncontrolled hypertension, history of arterial dissection of the cervicocephalic arteries, history of stroke, history of angina pectoris or myocardial infarction, coagulopathy, on anti-platelets or anti-coagulant therapy, other concomitant autoimmune disease.	Reduce ARR (49%–55%), reduce MRI lesions (old 61%–63%, new: 17%–32%), and EDSS score (27%–42%).	Acquired haemophilia A, ITP, TTP, kidney, thyroid, red and white blood cells (RBC & WBC) disorders, autoimmune encephalitis.
	Daclizumab	Zinbrya	Anti-CD25	Subcutaneous injection	2016	2016	Relapsing forms of MS	Hypersensitivity reactions to daclizumab, hepatic disease or hepatic impairment, history of autoimmune hepatitis or other autoimmune condition involving the liver.	Significant ARR reduction (54%), reductions in new lesions (69%–78%).	Liver problems (including autoimmune-related liver problems), immune-system problems (immune-mediated disorders).
	Ocrelizumab	Ocrevus	Anti-CD20	Intravenous	2017	2018	Relapsing forms of MS, early PPMS	Hypersensitivity reactions to ocrelizumab, active hepatitis-B infection.	Reduce ARR (46%), reduce MRI lesions (old: 94%, new: 77%–83%) and EDSS score (40%).	Back pain, depression, herpes virus-associated infections, infusion reactions, lower respiratory infections, pain in extremities, upper respiratory infections, cough, diarrhea, edema peripheral, skin reaction.
	Ofatumumab	Kesimpta	Anti-CD20	Subcutaneous injection	2020	2021	Relapsing forms of MS	Hypersensitivity reactions to ofatumumab, hepatitis-B infection, active infection.	Reduce relapses (44%), reduces MRI lesions (old: 96.4%, new: 82.7%), and reduce disability worsening events (21.6%).	Upper respiratory infections, injection-related reaction, skin reaction, urinary tract infections, decrease in IgM levels, oral herpes.

FDA, Food and Drug Administration; EMA, European Medicines Agency; EDSS, expanded disability status scale; AML, acute myeloid leukemia; HIV, human immunodeficiency virus; ITP, immune thrombocytopenic purpura; TTP, thrombotic thrombocytopenic purpura; DMTs, disease-modifying therapies; MS, multiple sclerosis; SPMS, secondary-progressive MS; PRMS, progressive-relapsing MS; RRMS, relapsing-remitting MS; PPMS, primary-progressive MS; MRI, magnetic resonance imaging; PML, progressive multifocal leukoencephalopathy; ARR, annualized relapse rates; CD52, cluster of differentiation 52; IgM, immunoglobulin M. & patients whose neurologic status is significantly abnormal between relapses.

Table 2. The currently FDA- and EMA-approved S1PR Modulator for the treatment of MS.

Type	Drug name	Brand name	Mechanism of action	Method of administration	Approval year (FDA)	Approval year (EMA)	Approved for	Contraindications	Efficacy	Adverse effect
S1PR modulator	Fingolimod	Gilenya	S1P _{1,3,4,5R} modulator	Oral	2010	2011	Relapsing forms of MS	Hypersensitivity reactions to fingolimod, cardiovascular events or problems (including; recent myocardial infarction, unstable angina, stroke, transient ischemic attack, AV block, cardiac arrhythmias, etc.).	Reduce relapses (54% reduction in ARR) and slow disability worsening.	Cardiac and macular effects, posterior reversible encephalopathy syndrome, liver damage, breathing problems, severe worsening of MS after discontinuation of fingolimod, increase blood pressure, Basal cell carcinoma and melanoma, allergic reaction.
	Siponimod	Mayzent	S1P _{1,5R} modulator	Oral	2019	2020	Relapsing forms of MS, active SPMS	Hypersensitivity reactions to siponimod or any of its ingredients, immunodeficiency syndrome, PML or cryptococcal meningitis, active cancer, severe liver problems, heart problems, pregnant or planning for pregnancy.	Reduce ARR (52%), total (58.1%) and regional (58.3%–85.4%) brain atrophy, and confirmed disability progression (22%).	Skin reaction, basal cell carcinoma, lymphopenia, convulsions, macular oedema, atrioventricular block, bradcardia, squamous cell carcinoma.
	Ozanimod	Zeposia	S1P _{1,5R} modulator	Oral	2020	2020	RRMS, active SPMS, CIS	Hypersensitivity reactions to ozanimod or any of its ingredients, immunodeficiency, heart problems, severe infection, cancer, liver problem, pregnancy or planning for pregnancy.	Reduce ARR (38%–48%), reduce MRI lesions (old: 53%–63%, new: 42%–48%), and EDSS score (6 months: 8.6%).	Heart problems, urinary track infection, increase in blood pressure, skin reaction, PML (rare).
	Ponesimod	Ponvory	S1P _{1R} modulator	Oral	2021	2021	RRMS, active SPMS, CIS	Hypersensitivity reactions to ponesimod or any of its ingredients, immunodeficiency, heart problems, severe infection, cancer, liver problem, pregnancy or planning for pregnancy.	Reduce ARR (31%), reduce MRI lesions (56%), and EDSS score (17%).	Urinary tract infection, bronchitis, viral infections, herpes zoster viral infection, pneumonia, vertigo, pyrexia, macular oedema, seizures, bradcardia.

FDA, Food and Drug Administration; EMA, European Medicines Agency; S1PR, sphingosine 1-phosphate receptor; EDSS, expanded disability status scale; AV, atrioventricular; CIS, clinically isolated syndrome.

Table 3. The currently FDA- and EMA-approved DNA replication and repair interfering agents for the treatment of MS.

Type	Drug name	Brand name	Mechanism of action	Method of administration	Approval year (FDA)	Approval year (EMA)	Approved for	Contraindications	Efficacy	Adverse effect
Interfere with DNA replication and repair	Teriflunomide	Aubagio	Pyrimidine synthesis inhibitor	Oral	2012	2013	RRMS	Hypersensitivity reactions to teriflunomide or any of its ingredients, severe hepatic impairment, immunodeficiency, pregnancy or planning for pregnancy, breast-feeding, impaired bone marrow function, active infection, renal problems, severe hypoproteinaemia.	Reduce ARR (31%), reduce MRI lesions (old: 57%, new: 44%) and EDSS score (20%).	Pancreatic inflammation, allergic reaction, skin reaction, severe infection or sepsis, lung inflammation, transaminitis, teratogenicity, neuropathy, hypertension.
	Cladribine	Mavenclad	Purine antimetabolite	Oral	2019	2017	Relapsing forms of MS	Hypersensitivity reactions to cladribine or any of its ingredients, HIV infection, active tuberculosis, hepatitis, immunodeficiency or taking immunosuppression medication, active cancer, moderate or severe kidney problems, pregnancy or planning for pregnancy, breast-feeding.	Reduce ARR (55%–58%), reduce MRI lesions (old: 86%, new: 73%), and EDSS score (47%).	Lymphopenia, shingles, liver problems, skin reaction, allergic reaction, hair loss, tuberculosis (rare).

FDA, Food and Drug Administration; EMA, European Medicines Agency; EDSS, expanded disability status scale; HIV, human immunodeficiency virus.

Table 4. The currently FDA- and EMA-approved DMTs for MS treatment whose mechanisms of action remain unclear.

Type	Drug name	Brand name	Mechanism of action	Method of administration	Approval year (FDA)	Approval year (EMA)	Approved for	Contraindications	Efficacy	Adverse effect
Unknown	IFN β -1b	Betaferon	Unknown	Subcutaneous injection	1993	1995	RRMS, active SPMS, CIS	Hypersensitivity reactions to IFN β -1b or any of its ingredients, current severe depression or suicidal ideation, liver disease.	Reduce ARR (34%), reduce MRI lesions (old: 83%, new: 75%), and EDSS score (29%).	Skin reaction, allergic reaction, depression or suicidal ideation, excessive bleeding after injury, infections, GI problems, heart problems.
		Extavia			2009	2008		Hypersensitivity reactions to IFN β -1b or any of its ingredients.		Skin reaction, lymphopenia, flu-like symptoms, myalgia, leukopenia, neutropenia, increased liver enzymes, headache, hypertonia, pain, insomnia, abdominal pain, asthenia.
	IFN β -1a	Avonex	Unknown	Intramuscular injection	1996	1997	RRMS, SPMS	Hypersensitivity reactions to IFN β -1a or any of its ingredients, current severe depression or suicidal ideation.	Reduce ARR (low dose: 29%, high dose: 32%), reduce MRI lesions (old-low dose: 22%, old-high dose: 67%, new-low dose: 67%, new-high dose: 78%), and EDSS score (low dose: 32%, high dose: 28%).	Allergic reaction. Depression.
		Rebif		Subcutaneous injection	1998	1998	RRMS, active SPMS, CIS	Hypersensitivity reactions to IFN β -1a or any of its ingredients, current severe depression or suicidal ideation.		Liver problems. Allergic reaction, depression. Liver problems, skin reaction, flu-like symptoms, thyroid dysfunction, MS pseudo-relapse, GI problems, headache.
		Tecfidera		Oral	2013	2014	RRMS	Hypersensitivity reactions to DMF or any of its ingredients.	Reduce ARR (44%–53%), reduce MRI lesions (old: 74%–90%, new: 71%–85%), and EDSS score (38%).	Lymphopenia, PML, allergic reaction, GI problems, skin reaction, hair loss.
	DMF	Generics	Unknown		2020	2021		Suspicion or confirmed PML.		

Table 4. Continued.

Type	Drug name	Brand name	Mechanism of action	Method of administration	Approval year (FDA)	Approval year (EMA)	Approved for	Contraindications	Efficacy	Adverse effect
	MMF	Bafiertam	Unknown	Oral	2013	-	RRMS, active SPMS, CIS	Hypersensitivity reactions to MMF or any of its ingredients, liver problems, lymphopenia, active infection, pregnancy or planning for pregnancy.	-	Allergic reaction, PML, herpes zoster infection, lymphopenia, liver problem, skin reaction, GI problems.
	DRF	Vumerity	Unknown	Oral	2013	2021	RRMS	Hypersensitivity reactions to DRF or any of its ingredients, PML, severe liver disease, severe kidney disease, serious infection, GI disease.	Reduce ARR and reduce MRI old lesions (77%).	Allergic reaction, PML, skin reaction, GI problems, lymphopenia, pneumonia.
	GA	Copaxone Glatopa (US) Generic	Unknown	Subcutaneous injection	1996 2015 2017	2004 ? 2016	RRMS, active SPMS, CIS	Hypersensitivity reactions to GA or any of its ingredients.	Reduce ARR (29%).	Skin reaction, vasodilation, dyspnea, chest pain.
	Pegylated IFN β -1a	Plegridy	Unknown	Intramuscular injection	2021	2020	RRMS	Hypersensitivity reactions to Pegylated IFN β -1a or IFN β -1a or any of its ingredients, current severe depression or suicidal ideation.	Reduce ARR (27%), MRI lesions (old: 86%, new: 67%), and EDSS score (38%).	Liver problems, depression, allergic reaction, seizures, skin reaction, kidney problems, blood problems, flu-like symptoms, myalgia, arthralgia, asthenia.

FDA, Food and Drug Administration; EMA, European Medicines Agency; IFN β , interferon-beta; DMF, dimethyl fumarate; MMF, monomethyl fumarate; GA, glatiramer acetate; DRF, diroximel fumarate; EDSS, expanded disability status scale; GI, gastrointestinal.

various physiological processes including angiogenesis, neurogenesis, immune cell trafficking, endothelial barrier function, and vascular tone. S1P1R mediates the responsiveness of lymphocytes to the S1P chemotactic gradient, allowing them to traffic between secondary lymphoid tissues and efferent lymphatics [6,11,20]. Blocking S1P1R function leads to the sequestration of lymphocytes in the lymph nodes and thymus. S1P1R is also expressed on endothelial cells, where it regulates dendritic cell (DC) recruitment and vascular permeability [6,11,20]. In animal models, inhibiting S1P1R suppresses cytokine amplification and immune cell recruitment, which is the primary therapeutic target in MS treatment [6,11,20]. Moreover, the expression of S1PRs on various CNS cell populations suggests that their direct effects on the CNS may contribute to the therapeutic benefits observed. In the experimental autoimmune encephalomyelitis (EAE), MS mouse model, blockade of S1P1R enhances neuronal survival and inhibits gliosis and demyelination. The clinically relevant functions of the other S1PR subtypes (S1P2R, S1P3R, S1P4R, and S1P5R) are less well-established compared to S1P1R [20] (Table 2).

2.2.2.1 Fingolimod. Fingolimod, which targets S1P1R, S1P3R, S1P4R, S1P5R, was the first S1P1R modulator approved for clinical use [6,11,20]. It works by binding to S1P1R on lymphocytes, leading to receptor internalization and loss of responsiveness to the S1P gradient that drives lymphocyte migration from lymph nodes [6,11,20]. This reduces the number of circulating inflammatory cells, limiting their entry into the CNS. Beyond this systemic immunological effect, fingolimod may also stabilize the blood-brain barrier (BBB) and exert direct effects on CNS cells, such as reduced astrogliosis, axonal loss, and demyelination, in EAE models [6,11,20] (Table 2).

2.2.2.2 Siponimod, Ozanimod, and Ponesimod. Subsequently developed S1PR modulators, such as siponimod, ozanimod, and ponesimod, are more specific in targeting only S1P1, 5Rs or S1P1R, with the aim of maintaining efficacy while potentially reducing adverse effects [6,11,20]. All S1PR modulators are believed to have a similar impact on lymphocyte trafficking, but their direct CNS effects may vary and are not yet fully understood, particularly for S1P5R-mediated mechanisms [6,11,20] (Table 2).

2.2.3 DNA Replication and Repair Interfering Agents

2.2.3.1 Teriflunomide. Teriflunomide mechanism of action in MS is hypothesized to relate to its effects on the proliferation of stimulated lymphocytes [6,11,21]. During the cell cycle, lymphocytes undergo division, which involves an S phase (DNA synthesis) and M phase (cell division), separated by G phases. DNA synthesis in the S phase requires the provision of new building blocks, including pyrimidine and purine bases. Teriflunomide blocks *de novo* pyrimidine synthesis by specifically and reversibly inhibit-

ing the mitochondrial enzyme dihydro-orotate dehydrogenase (DHODH), which is expressed at high levels in proliferating lymphocytes [21]. This teriflunomide-mediated blockade of pyrimidine synthesis interrupts the cell cycle in the S phase, exerting a cytostatic effects on the proliferation of T and B cells, thereby limiting their involvement in the inflammatory processes underlying the pathogenesis of MS [21] (Table 3).

Teriflunomide selectively inhibits rapidly proliferating, activated lymphocytes. Resting lymphocytes can undergo homeostatic proliferation using the salvage pathway, bypassing the DHODH enzyme targeted by teriflunomide [21]. This selective targeting allows teriflunomide to inhibit the hyperactive, proliferating lymphocytes involved in MS, without affecting other proliferating cell types such as gastrointestinal (GI) mucosal cells that express lower DHODH levels and do not display the same rapid expansion [21] (Table 3).

2.2.3.2 Cladribine. Cladribine is a nucleoside analog resistant to adenosine deaminase, allowing its accumulation in cells [6,11,22]. Phosphorylation of cladribine to its active triphosphate form (2-CdATP) impairs both nucleic acid synthesis and repair [22]. Cladribine also inhibits ribonucleotide reductase, further disrupting nucleotide synthesis [22]. These combined effects are predicted to induce apoptotic cell death, particularly in lymphocytes that rely on both *de novo* and salvage nucleotide pathways for DNA replication during clonal expansion [22]. The slow depletion of lymphocytes observed with therapeutic doses of cladribine suggests that it affects both resting and dividing cells, although the precise mode of action in MS is not fully understood [22] (Table 3).

2.2.4 Other Agents

2.2.4.1 IFN β . IFN β , a natural cytokine, is widely used to treat relapsing forms of MS [6,11,23]. It has complex therapeutic mechanisms, including anti-inflammatory and neuroprotective effects [23]. IFN β modulates the inflammatory environment, reduces immune cell trafficking, and enhances nerve growth factors, supporting neuronal survival and repair [23]. It also increases the numbers of a specialized subset of NK cells, which produce anti-inflammatory molecules, potentially helping to curb neuroinflammation [23]. The multifaceted effects of IFN β , spanning inflammation regulation, cellular trafficking, neurotrophic support, and immune cell modulation, highlight its crucial role in managing this debilitating neurological condition [23] (Table 4).

2.2.4.2 DMF. DMF modifies a variety of proteins involved in T-cell activation through its electrophilic activity [6,11,24]. Additionally, DMF reduces the production of nitric oxide synthase and pro-inflammatory cytokines [24]. Beyond modulating T-cell function, DMF also affects the survival of immune cells themselves. This impact on cell

survival is likely the underlying cause of the high incidence of lymphopenia associated with DMF treatment [24] (Table 4).

2.2.4.3 MMF. MMF is the active metabolite of DMF. MMF is believed to exert its therapeutic effects through activation of nuclear factor erythroid 2-related factor 2 (Nrf2) and nuclear factor kappa B (NF- κ B) pathways, reducing oxidative stress and inflammation [25]. Specifically, Nrf2 activation by MMF provides cytoprotection in astrocytes, while MMF also decreases adhesion molecule expression, reducing monocyte migration across the BBB [25]. Additionally, MMF can modulate the immune response by impairing DC maturation and T cell activation [25]. Furthermore, Nrf2 activation by MMF has demonstrated neuroprotective effects on animal models of ischemia-reperfusion injury [25]. However, the precise mechanisms of action of DMF and its metabolite MMF in MS are not fully understood [6,11,25] (Table 4).

2.2.4.4 DRF. DRF is an oral fumarate approved for RRMS [6,11,26]. Similar to DMF, DRF is converted to the active metabolite MMF [6,11,26]. DMF has demonstrated efficacy in MS, but commonly causes GI adverse events in up to 40% of patients. By contrast, DRF has shown improved GI tolerability compared to DMF [6,11,26]. This may be due to the different chemical structure of DRF, resulting in lower production of the irritating metabolite methanol, as well as fewer off-target interactions. The better GI tolerability profile of DRF provides an alternative treatment option for RRMS patients unable to tolerate the GI side effects of DMF [11,26] (Table 4).

2.2.4.5 GA. GA has several proposed mechanisms of action. One mechanism is binding to major histocompatibility complex (MHC) class II molecules on antigen-presenting cells (APCs), preventing stimulation of myelin basic protein (MBP)-specific cells. Another mechanism is shifting the immune response from a pro-inflammatory to an anti-inflammatory pattern. GA inhibits the secretion of pro-inflammatory cytokines (interleukin 2 [IL-2], IL-12, IFN γ , tumor necrosis factor [TNF]) by T helper 1 (Th1) and induces the production of anti-inflammatory Th2 suppressor cells that can cross the BBB. Yet another mechanism is inducing the generation of T regulatory (Tregs), such as CD4⁺, CD8⁺, and CD4⁺CD25⁺ cells, which are associated with the suppression of MS [6,11,27]. Through these multifaceted effects on the immune system, GA aims to modulate the aberrant inflammatory processes underlying MS pathology [6,11,27] (Table 4).

2.3 Efficacy of the Currently Approved DMTs

Each DMT offers unique benefits along with safety concerns and substantial costs, providing vital treatment prospects for people with MS [11,14]. For example, IFN β and GA are considered “platform” therapies with mod-

est efficacy (~30% decrease in annualized relapse rates [ARRs]) but long-term safety [11,28]. Fingolimod has been more effective in reducing relapses (54% reduction in ARR) and slowing disability progression, but with possible adverse cardiac and macular effects [11,29]. Other DMTs, such as siponimod, ozanimod, and ponesimod, have shown promise in reducing disease disability progression by attenuating demyelination and promoting remyelination, with fewer adverse cardiovascular effects and cortical gray matter volume loss [11,30,31]. While ponesimod displays lower relapse rates and disability accumulation than teriflunomide, it has a higher incidence of adverse effects [11,32]. Teriflunomide has shown similar efficacy as IFN β -1a but with much worse adverse effects, while DMF has shown a 48–53% reduction in ARR compared to placebo but with side effects mostly related to the GI tract [11,33,34]. However, the newer fumarates, DRF and MMF, show promising efficacy with fewer GI side effects [11,35]. Natalizumab has been shown to reduce clinical relapses by 67%, new brain magnetic resonance imaging (MRI) lesions by 83%, and disability progression by 42%; however, due to the risk of PML in JCV-exposed patients, its use is restricted [11,15,36]. Ofatumumab and ocrelizumab have demonstrated significant ARR reduction, with notable reductions in MRI lesion rates, while alemtuzumab and cladribine also exhibit a high rate of relapse reduction, combined with decreased brain atrophy and lesion burden. However, each of these therapies also carries potential risks, such as the risk of developing secondary autoimmune disorders with alemtuzumab [11,18,19,37–39], and an increased incidence of lymphopenia with cladribine (Tables 1,2,3,4) [11,40].

3. Pathophysiology of MS

Understanding the pathophysiology of MS is crucial for determining the most suitable treatment for patients. MS begins with an immune response triggered by various unknown factors such as viruses, environmental factors, and genetics predisposing susceptibility factors [1,41]. Several viruses have been linked to MS, with Epstein-Barr virus (EBV) playing a particularly important role. The significance of EBV concerning disease susceptibility is well known [42]. EBV infiltrates the brain through B cells and establishes chronic infection, leading to periodic reactivation and inflammatory responses. The presence of memory B cells and the similarity between EBV and MBP epitopes can trigger autoimmune reactions against the myelin sheath, causing axonal demyelination [42]. Additionally, EBV expresses an abnormal autoantigen found in the brains of MS patients [42].

The environmental factors that have garnered significant attention regarding their impact on MS include vitamin D levels, obesity, cigarette smoking, and gut microbiomes [43–47]. Research indicates a strong connection between vitamin D deficiency and both the onset and progression of MS, with even baseline levels of vitamin D even influencing disease activity. Individuals with a genetic tendency for

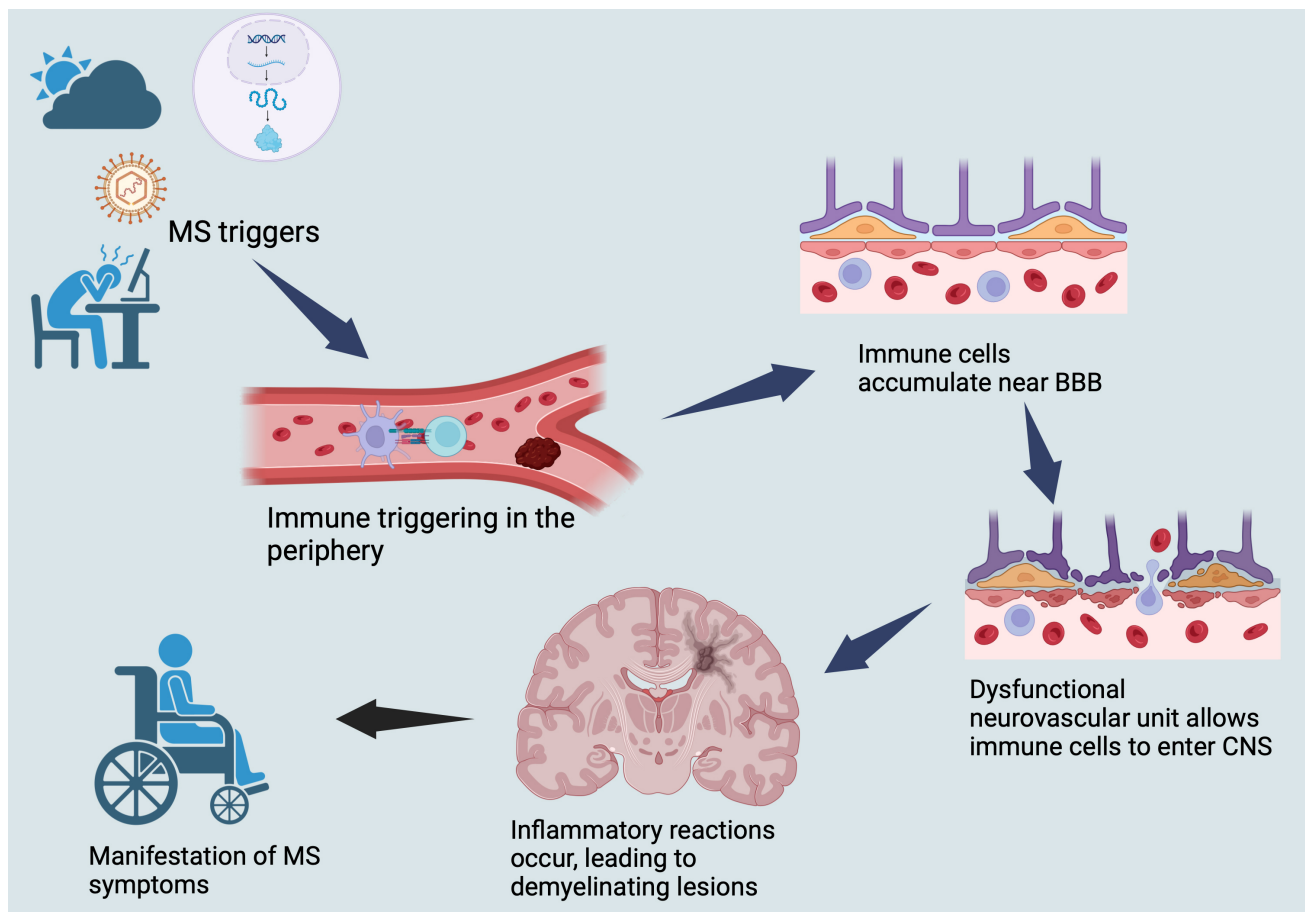


Fig. 1. Illustrates a brief overview of MS pathophysiology. MS, multiple sclerosis; CNS, central nervous system; BBB, blood-brain barrier.

higher vitamin D levels tend to have a reduced risk and later onset of disease [43,44].

Additionally, a link has been identified between obesity in early life and an increased risk of developing MS, regardless of other influencing factors [45]. Cigarette smoking also appears to contribute to the progression of MS, although the precise mechanisms underlying this relationship remain unknown, leading to ongoing disagreements among researchers regarding its effects across different populations [46].

Moreover, the gut microbiome is increasingly recognized as an important factor in the pathogenesis of MS. Research has demonstrated that the intestinal microbiome can influence the development of CNS-reactive T cells and modulate the function of CNS-resident cells. Notably, transplantation of gut microbiota from MS patients can exacerbate EAE [47]. Investigations into the gut microbiome of individuals with MS have revealed changes that relate to both pro-inflammatory and anti-inflammatory responses [47].

Several genetic factors have been linked to the risk of developing MS, with the human leukocyte antigen (HLA)-DRB1*15:01 allele being the most significant, although it explains less than half of the genetic predisposition to

the disease [1,48,49]. In addition to this allele, other regions within the HLA complex, as well as some non-HLA regions, have also been closely associated with MS risk [1,48,49]. Genome-wide association studies and linkage analyses have uncovered numerous genetic loci related to the immune, nervous, and hemostatic systems that contribute to susceptibility to MS. Furthermore, various epigenetic mechanisms, such as DNA methylation, histone modification, and mRNA methylation, are believed to play a role in the progression and development of MS [1,48,49].

MS is a complex autoimmune disease primarily affecting the CNS. The CNS is an immune-privileged site, with the BBB selectively regulating the entry of molecules and cells. In MS, this immune privilege is disrupted, leading to the undesirable activation of antigen-specific T and B cells. This triggers a cascade of self-damaging processes, resulting in CNS inflammation, demyelination, axonal damage, and neurodegeneration [1,3,48]. The exact mechanisms behind T⁺ cell activation in MS are not fully understood, but one theory suggests peripheral T⁺ cell activation, followed by their re-circulation into the CNS driven by the S1P gradient. The approved MS drug fingolimod exerts its action by blocking the S1P1R, supporting the importance of this pathway in disease activation [1,3,29,48].

Once in the CNS, activated T cells release immunoregulatory molecules, leading to demyelination through various hypothesized processes, such as receptor-mediated phagocytosis of myelin, myelin stripping, vesicular disruption of myelin sheaths, and oligodendroglial pathology [1,3,48,50,51]. During active stages of MS, oligodendrocytes are dramatically lost due to immune-mediated mechanisms, but large numbers of undifferentiated oligodendrocyte progenitor cells (OPCs) are recruited to the site of lesions [1,3,48,50]. In some cases, severe demyelination may occur with complete loss of oligodendrocytes and lack of remyelination, which may result from pathological mechanisms that effectively destroy all oligodendrocytes and their progenitors, or from antigen-mediated immune responses against these cells [1,3,48,50]. In cases of attempted remyelination, some stressed oligodendrocytes may be at risk of destruction, potentially driven by gamma-delta ($\gamma\delta$) T cells reacting to stress proteins or due to insufficient growth factors [1,3,48,50].

The disease course of MS typically involves episodic, relapsing-remitting phases driven by inflammation, followed by progressive phases characterized by demyelination and axonal loss (neurodegeneration) [1,48,50]. Understanding the complex interplay between these two underlying mechanisms is crucial for developing effective treatments for this debilitating disease (Fig. 1).

3.1 Immune Triggering in the Peripherally

The initiation of inflammatory events in MS is thought to occur in the periphery with the activation of T and B cells, which are subsequently transferred to the CNS [1,48,50].

3.1.1 APCs

The development of MS involves the activation of immune cells located in the periphery by APCs. When the self-antigens are presented, such activation results in the stimulation of auto-reactive T cells, which ultimately make their way to the CNS and initiate MS lesions. APCs comprise of several cell types such as DCs, monocytes, macrophages, and tissue-resident cells, which include CNS microglia [52,53]. DCs, a type of APC, are believed to be the primary APCs involved in the pathophysiology of MS and are known to be impacted by several DMTs used to treat MS [53] including IFN β , GA, S1PR modulators, DMF, natalizumab, daclizumab, mitoxantrone, and cladribine [12] (Table 5).

For example, IFN β therapy induces apoptosis and alters cytokines in DCs, reduces their migratory capacity, and affects T-cell differentiation [54]. GA reduces the antigen presentation capability of DCs [55], while S1PR modulators weaken their phagocytic and immunostimulatory capacity while affecting DC trafficking [12]. DMF inhibits the maturation and migration of DCs and affects T cell differentiation [56]. Natalizumab and alemtuzumab reduce the count of DCs in lesions and promote tolerogenic

functions in plasmacytoid DCs [57,58]. Daclizumab inhibits DC-mediated activation of T cells and induces anti-inflammatory cytokine production [59]. Mitoxantrone impairs the antigen-presenting capacity of DCs, and cladribine induces their apoptosis [60,61].

Monocytes and monocyte-derived macrophage were linked to advanced clinical disability, and disease progression in MS and EAE models. By contrast, microglia have been linked to early MS lesions and MS susceptibility genes, with evidence pointing toward them as being more closely associated with MS genes than neurons or astrocytes [62]. Various DMTs have been recorded to suppress or deactivate these cells during the course of MS (Table 5). For instance, mitoxantrone blocks myelin breakdown by macrophages [63], while natalizumab reduces microglial activation and suppresses the accumulation of microglia/macrophage populations in the CNS [64]. Alemtuzumab reduces the expression of major histocompatibility complex class II (MHC-II) and costimulatory molecules on microglia and infiltrating macrophages [65], while fingolimod impacts their activation and phagocytic ability [66]. Cladribine affects the differentiation of monocytes into macrophages, reduces their pro-inflammatory response, and induces microglial apoptosis [67]. Lastly, DMF suppresses pro-inflammatory macrophages [68], while GA inhibits the activation of monocytes and induces the differentiation of anti-inflammatory macrophages [69].

3.1.2 Tregs

Tregs are a type of lymphocyte that plays a vital role in maintaining self-tolerance and regulating immune responses in MS and EAE. Tregs are divided into two types: naturally occurring CD4⁺ CD25⁺ forkhead box P3⁺ (FOXP3⁺) Tregs that arise from the thymus, and adaptive Tregs that arise from immune responses outside the thymus. Both types of Tregs help to maintain immune homeostasis and regulate autoimmune inflammation. The differentiation of Treg precursor cells into effector Tregs is influenced by transforming growth factor beta (TGF- β) and IL-2, while the suppressive activity of Tregs is mediated by interleukin 10 (IL-10). Other regulatory cells, such as CD8⁺ Tregs, B regulatory cells (Bregs), and regulatory NK cells, also help to maintain immune homeostasis and regulate immune responses. However, studies have pointed to a depletion, dysregulation, or ineffectiveness of regulatory cells on effector T cells in MS [70–72]. Several DMTs reportedly influence the levels of Tregs, Bregs, and/or regulatory NK cells in MS, including IFN β therapy, GA treatment, fingolimod, siponimod, DMF, cladribine, alemtuzumab, and daclizumab (Table 5). These DMTs work through varying mechanisms to enhance Treg counts, increase Treg populations, or promote the increase of other regulatory cells, such as Bregs [73–82].

Table 5. An overview of the pathophysiological alterations linked to MS, along with the impact of existing DMTs on these changes.

Pathophysiological aspects of MS	Immune triggering in the periphery					Meningeal lymphatic vessels	Migration into the CNS	Crossing the BBB into CNS	Inflammatory factors	CNS demyelination and neurodegeneration			
	APCs		Treg	Activation of T ⁺ and B ⁺ cells	Glial cells					Neurons			
	DC	Monocytes and monocyte-derived macrophage			Oligodendrocytes						Astrocytes	Microglia	
Targeted DMTs	IFN β , S1PR modulators, natalizumab, daclizumab, mitoxantrone, and cladribine.	GA, DMF, fingolimod, cladribine, DMF, and GA.	Mitoxantrone, natalizumab, alemtuzumab, fingolimod, cladribine, alemtuzumab, and daclizumab.	IFN β , GA, fingolimod, siponimod, DMF, cladribine, alemtuzumab, and daclizumab.	Mitoxantrone, natalizumab, alemtuzumab, daclizumab, ocrelizumab, ofatumumab, fingolimod, INF β , siponimod, ponesimod, teriflunomide, cladribine, DMF, and GA.	There is no known impact of DMT.	Mitoxantrone, natalizumab, alemtuzumab, fingolimod, siponimod, ozanimod, ponesimod, and IFN β -1b.	Mitoxantrone, natalizumab, alemtuzumab, cladribine, fingolimod, siponimod, teriflunomide, and IFN β -1b.	Natalizumab, daclizumab, ocrelizumab, ofatumumab, S1PR modulators, teriflunomide, cladribine, IFN β , DMF, and GA.	Alemtuzumab, fingolimod, siponimod, ponesimod, teriflunomide, IFN β , GA, and DMF.	Fingolimod, natalizumab, ocrelizumab, siponimod, teriflunomide, IFN β , DMF, and MMF.	Natalizumab, fingolimod, ocrelizumab, ofatumumab, siponimod, teriflunomide, cladribine, IFN β , DMF, MMF, and GA.	Fingolimod, natalizumab, alemtuzumab, siponimod, teriflunomide, cladribine, IFN β , DMF, MMF, DRF, and GA.
DMTs Impact	<ul style="list-style-type: none">• Induction of apoptosis.• Alteration of cytokine profile.• Decrease in dendritic cell (DC) migratory capacity.• Impact on T⁺ cell differentiation.• Reduction in antigen presentation ability of DCs.• Weakening of DCs' phagocytic and immunostimulatory capacity.• Inhibition of DC maturation.• Decrease in the quantity of DCs in lesions.	<ul style="list-style-type: none">• Blocking myelin breakdown.• Decreasing activation and accumulation in the CNS.• Reducing the expression of MHC-II and costimulatory molecules.• Impacts phagocytic ability.• Affects differentiation.• Induction of apoptosis.• Promotion of anti-inflammatory profile.	<ul style="list-style-type: none">• Augmenting Treg and Bregs populations and efficacy.	<ul style="list-style-type: none">• Inhibiting the proliferation, differentiation, and activation of inflammatory T⁺ and B⁺ cells.• Increasing levels of naive and anti-inflammatory cells.	-	<ul style="list-style-type: none">• Decreasing leukocyte migratory activity through the BBB.• Inhibiting chemokines responsible for immune cell migration into the CNS.• Restricting cytokine signaling between the CNS and immune system.	<ul style="list-style-type: none">• Decreasing the infiltration of leukocytes through the BBB.• Inhibiting the migratory capacity of immune cells.• Restoring levels of tight junction proteins.• Promoting BBB integrity.• Reducing MMP activity.• Increasing levels of TIMPs and sVCAM-1.	<ul style="list-style-type: none">• Modifying markers associated with T⁺ and B⁺ cells.• Altering subsets of T⁺ cells.• Decreasing the quantity of inflammatory Th17 cells.• Reducing inflammation and the number of cytotoxic CD8⁺ T⁺ cells.• Enhancing the population of Tregs.• Increasing the levels of naive T⁺ cells.• Decreasing central memory T⁺ cells.	<ul style="list-style-type: none">• Decreasing apoptosis and promoting the differentiation, maturation, and remyelination of oligodendrocytes.	<ul style="list-style-type: none">• Suppressing astrocytic inflammatory reactions.	<ul style="list-style-type: none">• Decreasing microglial inflammasome activation.• Enhancing the anti-inflammatory phenotype.	<ul style="list-style-type: none">• Boosting axonal metabolism.• Restoring proper neuronal network function.• Normalizing excitability.• Providing protection against neuronal damage.• Ameliorating previous tissue damage.• Reducing axonal and neuronal degeneration.	

Table 5. Continued.

Pathophysiological aspects of MS	Immune triggering in the periphery				Meningeal lymphatic vessels	Migration into the CNS	Crossing the BBB into CNS	Inflammatory factors	CNS demyelination and neurodegeneration			
	APCs		Treg	Activation of T ⁺ and B ⁺ cells					Glial cells			Neurons
	DC	Monocytes and monocyte-derived macrophage							Oligodendrocytes	Astrocytes	Microglia	
								<ul style="list-style-type: none">• Amplifying the responsiveness of T⁺ effector cells to immune regulation.• Reduce pro-inflammatory cytokines, and increase anti-inflammatory cytokines.• Decrease levels of chemokines linked to both Th1 and Th2 responses.				

MS, multiple sclerosis; CNS, central nervous system; S1PR, sphingosine 1-phosphate receptor; IFN β , interferon-beta; DMF, dimethyl fumarate; MMF, monomethyl fumarate; GA, glatiramer acetate; DRF, diroximel fumarate; BBB, blood-brain barrier; APC, antigen-presenting cell; DC, dendritic cell; Treg, regulatory T⁺ cells; Breg, B regulatory cell; NK, natural killer; VCAM-1, vascular cell adhesion molecule-1; MMP, matrix metalloproteinase; TIMP, tissue inhibitors of metalloproteinase; Th, T helper; IFN γ , IFN-gamma; $\gamma\delta$ T cell, gamma-delta T cell; MHC-II, major histocompatibility complex class II.

3.2 Activation of T and B Cells

Inflammatory cells in MS patients are characterized by higher-than-normal levels of effector cytokines and expression of brain-homing chemokine receptors, as well as reactivity to antigens found in the CNS. It is believed that T and B cells in MS patients are prone to a deregulated inhibitory control, making the immune cells more prone to attack. These cells appear relatively resistant to regulation by regulatory cells. The disease pathogenesis implicates both T and B cells and their reactivity to multiple myelin antigens, such as MBP, myelin-associated glycoprotein, and proteolipid protein, which form multilamellar myelin sheaths around the axons and neuron cell bodies. Another protein, myelin oligodendrocyte glycoprotein, situated on the surface of the sheath, acts as an adhesive receptor, and connects neighboring myelinated fibers [51]. These cells infiltrate the BBB, activate local microglia and macrophages, thereby initiating a local inflammation process that leads to tissue damage and chronic inflammation [51].

Several DMTs, such as mitoxantrone, natalizumab, alemtuzumab, daclizumab, ocrelizumab, ofatumumab, fingolimod, INF- β , siponimod, ponesimod, teriflunomide, cladribine, DMF, and GA, have been shown to regulate the activity of these cells (Table 5). These DMTs work through various mechanisms to suppress the proliferation, differentiation, and activation of effector T and B cells and promote regulatory functions in CD4⁺ T cells, B cells, and NK cells. They also decrease the levels of immune cells, upregulate naïve and anti-inflammatory T and B cell counts, and alter cytokine profiles, ultimately leading to a shift towards Tregs and Bregs and a decrease in chronic inflammation [60,77,83–98].

3.3 Meningeal Lymphatic Vessels

The theory that encephalitogenic molecules travel to the peripheral areas to activate T and B cells is a subject of ongoing discussion. Lymphatic vessels located in the meninges play a role in transporting macromolecules and immune cells from the cerebrospinal fluid to the peripheral immune system via the cervical lymph nodes, potentially impacting the pathogenesis of MS. The hypothesis suggests that the pathogenicity of MS may be altered by the drainage of encephalitogens out of the CNS, which then triggers the activation of peripheral immune cells. To support this notion, the obstruction of meningeal lymphatic function has been linked to a mitigated course of EAE, as it reduces interactions between T cells and APCs [99,100]. Additionally, differences in the anteroposterior diameters of cervical lymph nodes between MS patients and healthy individuals lend further credence to the idea of lymphatic drainage contributing to the disease pathology [101]. To the best of the author's knowledge, no studies have yet explored the effect of DMTs on the meningeal lymphatic vessel system in MS, likely due to its recent discovery and the novelty of this area of research.

3.4 Migration into the CNS

Immune cells are believed to be directed towards the CNS where demyelination and neurodegeneration occur. The involvement of S1P and S1P1R has been suggested in this process. T cells express S1P1R on their plasma membrane, which gets downregulated while entering the lymph node. Following activation and clonal expansion, S1P1R expression increases, facilitating T cell migration from the lymph nodes. This establishes a gradient of S1P expression, which is sensed by S1P1R and can shuttle activated T cells back to the circulation possibly leading to the CNS [102]. Chemokines are crucial for guiding leukocytes to sites of inflammation within the CNS. Various chemokines have specific receptor-binding affinities for different leukocyte types, such as neutrophils, eosinophils, monocytes, DCs, and T cells. They work by directly attracting and activating integrins on leukocytes to bind to adhesion receptors found on endothelial cells [103]. DMTs impact this pathway in MS, such as mitoxantrone, natalizumab, alemtuzumab, fingolimod, siponimod, ozanimod, ponesimod, and IFN β -1b. These DMTs reduce leukocyte migration through the BBB and suppress the activity of chemokines involved in cell migration into the CNS. They also affect gene expression changes in immune cells, limit cytokine communication between the immune system and CNS, and alter the migration of specific immune cell subsets into the CNS [20,84,104–108] (Table 5).

3.5 Crossing the BBB into CNS

The mechanism by which immune cells cross the BBB into the CNS still remains to be resolved, but some studies suggest impaired junctional integrity with proteins such as occludin, claudin-5, and vascular endothelial-cadherin, barrier properties demonstrated with serum leakage, and reduced or impaired efflux pump activity such as P-glycoprotein (P-gp) as possible reasons [109].

Moreover, during MS, the endothelial cells of the BBB shift their immune phenotype and increase expression of adhesion molecules such as E-selectin, L-selectin, intercellular adhesion molecule-1 (ICAM-1), VCAM-1, and atypical chemokine receptor 1, thus enabling immune cell infiltration [99]. ICAM-1 and VCAM-1 bind the integrins leukocyte functional antigen 1 and very late antigen-4 (VLA-4) and mediate cellular transfer through the BBB [110]. This leads to infiltration of immune cells, facilitated by matrix metalloproteinases (MMPs) expressed by leukocytes that tunnel through the tight junctions and cleave transmembrane β -dystroglycan, which anchors astrocytes' end feet to the basement membrane [111]. Gadolinium-enhanced MRI can detect focal BBB dysfunction, which arises early in the pathogenesis of new lesions in MS [109]. DMTs targeting this step include mitoxantrone, natalizumab, alemtuzumab, cladribine, fingolimod, siponimod, teriflunomide, and IFN β -1b. These DMTs work through various mechanisms to reduce leukocyte infiltration through the BBB, inhibit the migratory capacity of im-

immune cells, restore levels of tight junctional proteins, promote BBB integrity, reduce MMP activity, and enhance the levels of tissue inhibitors of metalloproteinases and soluble VCAM-1 [112–122] (Table 5).

3.6 Inflammatory Factors

Immune cell-driven inflammation is considered a subsequent phase in the development of MS. The prevailing view is that the immunological underpinnings of MS involve the mediating actions of T cells, B cells, monocytes, and occasional plasma cells [99]. MS lesions typically form around CNS veins and venules, with active demyelination closely linked to inflammatory infiltrates. These perivascular infiltrates contain lymphocytes and plasma cells, while active tissue damage is associated with macrophages and microglia, all orchestrated by various cytokines, chemokines, and their receptors [50].

3.6.1 B Cells

Numerous investigations have pointed to the involvement of B cells in the development of MS, as indicated by the detection of B cells, plasma cells, immunoglobulins, and complement deposition within MS lesions, along with the positive response to B cell-targeted therapies [123]. While B cells undergo affinity maturation and proliferation in germinal centers, the direction of B cell destiny is largely influenced by CD4⁺ T cells, specifically follicular Th cells, which guide their fate through a combination of membrane-bound and soluble stimulatory and inhibitory cues [124].

3.6.2 CD4⁺ T Cells

The significant role of CD4⁺ T cells and Th cells in MS is supported by evidence from EAE models. These cells precede the arrival of B cells in the CNS parenchyma, often reaching the site before clinical symptoms manifest, and they exhibit the capacity to trigger disease in otherwise healthy organisms. Furthermore, the targeting of antigens specific to Th cells has been shown to partially or even wholly suppress the progression of the disease [125].

Past studies have also highlighted those multiple subsets of CD4⁺ T cells, including Th17, Th1, and Th2 cells, are linked to MS lesions. It is primarily the Th1 cells, which produce IFN γ , and Th17 cells, that secrete IL-17, whose activities are closely associated with the pathogenic mechanisms underlying MS [51,126]. Th1 cells play a crucial role in the inflammatory process by secreting cytokines, primarily IFN γ and TNF- β . These cytokines activate macrophages, which subsequently damage oligodendrocytes, leading to pathological myelination. Th1 hyperactivation results in tissue damage and chronic inflammation, with IFN γ being essential for T-cell responses [51].

Th17 cells, another subset of Th cells, express the VLA-4 surface antigen, facilitating their migration into the CNS [51]. These cells exert multiple effects in the CNS, including the production of high levels of glutamate, which is harmful to oligodendrocytes. A subpopulation

of Th17 cells, known as Th17.1, secretes high levels of the IL-23 receptor. When stimulated, these cells produce granulocyte-macrophage colony-stimulating factor (GM-CSF), transforming into more encephalitogenic cells and intensifying CNS inflammation [51]. Both Th1 and Th17 cell populations increase during the onset of MS [51]. Th9 cells, an additional subset of CD4⁺ T cells, produce IL-9 and appear to have an anti-inflammatory effects on MS. Th9 cells differentiation is promoted by TGF- β and IL-9, which are produced by both Th9 and non-pathogenic Th17 cells [51]. GM-CSF⁺ C-X-C chemokine receptor type 4-positive (CXCR4⁺) T cells represent a distinct subset of Th cells characterized by their expression of both GM-CSF and CXCR4. In the context of MS, CXCR4 plays a crucial role in facilitating the migration of encephalitogenic lymphocytes across the BBB [51].

3.6.3 CD8⁺ T Cells

While initial studies largely concentrated on the pathogenic role of CD4⁺ T cells in MS, recent research has progressively acknowledged the significant involvement of CD8⁺ T cells in the disease [127]. In MS brain lesions, CD8⁺ T cells are regarded as the most prominent T cell populations and have been found to closely correlate with the degree of axonal damage [127].

3.6.4 $\gamma\delta$ T Cells

The $\gamma\delta$ T cells are a relatively small subset of T cells, with the majority (>70%) having double negative markers (CD8⁻CD4⁻), while a portion has CD8⁺ (~30%), and only a few have CD4⁺ (~1%) markers [128,129]. The involvement of $\gamma\delta$ T cells in the pathogenesis of both EAE and MS has also been established. Clonal populations of $\gamma\delta$ T cells have been detected in MS lesions and cerebrospinal fluid during the early stage of the disease. These cells are believed to have a significant correlation with MS pathogenicity [128,129]. DMTs can elevate inflammatory status in MS and improve patient symptoms, such as natalizumab, daclizumab, ocrelizumab, ofatumumab, S1PR modulators, teriflunomide, cladribine, IFN β , DMF, and GA (Table 5). These DMTs work through various mechanisms to influence immune cells, such as impacting markers associated with T and B cells, altering subsets of T cells, reducing the number of inflammatory Th17 and GM-CSF⁺ CXCR4⁺ T cells, decreasing inflammation and the number of cytotoxic CD8⁺ T cells, enhancing the number of Tregs, increasing the levels of naive T cells, decreasing central memory T cells, and enhancing the responsiveness of T-effector cells to immune regulation [87,130–145].

3.6.5 Macrophages

Macrophages are a dynamic and varied group of cells that play essential roles in immune responses and various disease processes [146]. They are considered professional phagocytes and are abundant within the CNS lesions of MS and EAE. These macrophages originate either from glial

cells or from infiltrating peripheral monocytes. In a healthy CNS, peripheral monocytes or macrophages are typically absent; their presence in EAE or MS lesions is often linked to disease onset. Their involvement in the CNS underscores their vital role in the pathogenesis of MS [146].

Depleting monocytes before the onset of symptoms can delay the emergence of EAE and lead to a milder disease progression, while depleting them after disease onset can slow the disease progression [146]. Early in EAE, an increase in peripheral monocytes correlates with greater clinical severity and a greater likelihood of progressing to paralysis. The recruitment of monocytes into the CNS is primarily mediated by chemokine signaling, particularly through the engagement of C-C motif chemokine ligand 2 (CCL2) and C-C motif chemokine receptor 2 (CCR2) in the myeloid compartment. CCR2 levels rise in the blood of EAE patients in response to GM-CSF stimulation [146]. Once monocytes reach the CNS parenchyma, they become activated and differentiate into either myeloid DCs or macrophages. Depending on the signals present within the CNS lesions, macrophages can differentiate into a pro-inflammatory phenotype (M1) or an anti-inflammatory phenotype (M2) [146].

M1 cells secrete various cytokines that are crucial for antigen presentation and the sustained activation of T-cells, contributing to damage to the myelin sheath and surrounding cells, which leads to axonal degeneration [146]. During the remission phase, M2 cells predominate in the CNS lesions. M2 cells suppress pro-inflammatory cytokines and antigen presentation, thereby inhibiting CD4 T cell responses and facilitating remyelination and the differentiation of oligodendrocytes within CNS lesions. The polarization of M2 cells is beneficial in preventing the development of EAE [146].

DMTs impact the pro-inflammatory activity of macrophages. For instance, mitoxantrone halts myelin degradation by macrophages [63], while natalizumab decreases their activation and presence in the CNS [64]. Alemtuzumab reduces MHC-II expression on macrophages [65], fingolimod modifies their activation and phagocytic ability [66]. Cladribine affect monocyte-to-macrophage differentiation and promotes their apoptosis [67]. DMF suppresses pro-inflammatory macrophages [68], and GA encourages the development of anti-inflammatory macrophages [69].

3.6.6 Neutrophils

Neutrophils play a significant role in the pathogenesis of MS. Animal models have demonstrated that neutrophils contribute to disease development and pathogenicity [147]. Following just 24 h of disease induction, neutrophils accumulate near the meninges in EAE, with their increased presence observed during both preclinical and peak phases of the disease [147]. GM-CSF and the secretion of chemokines (CXCL1, CXCL2, and CXCL6) from

Th17 cells promote the recruitment of neutrophils to the brain and spinal cord in EAE [147].

In the blood of MS patients, increased levels of neutrophil-activating chemokines and neutrophil-derived enzymes (e.g., CXCL1, CXCL8, neutrophil elastase, and myeloperoxidase [MPO]) are observed, which are linked to the formation of new inflammatory lesions [147]. The neutrophil-to-lymphocyte ratio is proposed as a marker of disease activity, as it tends to be elevated in MS patients, particularly during relapse compared to remission [147]. Neutrophils possess a diverse array of effector functions that contribute to disease pathogenesis, including the release of inflammatory mediators and enzymes such as IL-1 β , MMPs, and MPO; the destruction and phagocytosis of myelin debris; the release of neutrophil extracellular traps; the generation of reactive oxygen species; disruption of the BBB; and the production and presentation of autoantigens [147].

In addition to their role as pro-inflammatory cells, neutrophils also function as anti-inflammatory cells. They infiltrate tissues and enhance the uptake of apoptotic neutrophils by macrophages. Neutrophils scavenge inflammatory chemokines and cytokines using decoy and scavenger receptors [147]. The process of efferocytosis, where macrophages engulf apoptotic neutrophils, promotes the polarization of macrophages to an M2-like phenotype and negatively regulates inflammation [147]. Several DMTs have been observed to influence neutrophil function, counts and migratory capacity across the BBB. These include: IFN β -1b, GA, fingolimod, natalizumab, alemtuzumab, ofatumumab, DMF and MMF [148–152].

3.6.7 Cytokines and Chemokines

Cytokines are small, versatile proteins that play a crucial role in cellular communication. These molecular messengers exert a wide range of effects, known as pleiotropic functions, influencing various biological processes in both normal physiological states and disease conditions. By facilitating intercellular signaling, cytokines orchestrate complex interactions within the immune system and beyond, contributing to the maintenance of health and the development of pathological states [51]. In MS, cytokines play pivotal roles in the pathogenesis of the disease. They are instrumental in activating encephalitogenic lymphocytes, particularly Th1 and Th17 cells, and in facilitating the migration of these lymphocytes into the CNS [51]. The differentiation of Th1 cells is orchestrated by a triad of cytokines: IFN γ , IL-2, and TNF- α . Once activated, Th1 cells themselves produce IFN γ and TNF- α , establishing a self-perpetuating inflammatory environment. This cytokine milieu not only maintains the inflammatory state but also provides positive feedback, promoting the further differentiation of naive T cells into Th1 cells [51]. The pathogenic effects of IFN γ and TNF- α on MS are particularly significant. These cytokines contribute to the demyelination process, a hallmark of MS pathology. Their levels have been found to correlate

with disease severity and progression, often associated with exacerbation of MS symptoms [51]. This underscores the critical role these cytokines play in the inflammatory cascade that drives MS pathogenesis.

The differentiation of Th17 cells is subject to tight regulation, involving a complex interplay of cytokines. Primarily, this process requires the presence of TGF- β and IL-6. Additionally, IL-21 and IL-23 play crucial roles in supporting the proliferation and stabilization of Th17 cells [51]. Interestingly, in the absence of TGF- β , an alternative pathway for Th17 differentiation exists. This pathway is driven by a combination of IL-6, IL-23, and IL-1 β [51].

In the context of MS, researchers have observed significant alterations in the cytokine profile. Specifically, increased production levels of IL-6, IL-23, and IL-1 β have been noted. These elevated levels likely contribute to enhanced Th17 differentiation, and consequently, to the pathogenesis of MS [51]. Moreover, the role of TGF- β in MS appears to be complex. Elevated levels of TGF- β in the cerebrospinal fluid correlate with disease relapse. This observation suggests that while TGF- β is necessary for Th17 differentiation, its increased presence in the CNS might also be indicative of ongoing inflammatory processes or attempted regulatory responses in [51].

Cytokines also play a crucial role in modulating lymphocyte migration across the BBB. This process is intricate and involves multiple mechanisms [51]. TNF- α exerts a dual effect on the BBB. On one hand, it can disrupt the tight and adherent junction molecules that maintain the integrity of endothelial cells, thereby reducing BBB permeability. On the other hand, when TNF- α acts in concert with IFN γ , it can significantly alter the architecture of these junction proteins [51]. Moreover, the synergistic action of TNF- α and IFN γ enhances the expression of including ICAM-1, VCAM-1, and selectins. This increased expression facilitates lymphocyte adhesion and subsequent migration across the BBB [51]. Cytokines involved in Th17 cell differentiation also contribute to lymphocyte migration. For instance, IL-17 stimulates endothelial cells of the BBB to express chemokines such as CCL2 and CXCL8. These chemokines, move lymphocytes to the BBB, potentially exacerbating the inflammatory process [51]. This complex interplay of cytokines and their effects on the BBB and lymphocyte migration underscores the multifaceted nature of inflammatory processes in neurological disorders such as multiple sclerosis.

Several DMTs deplete cells that produce pro-inflammatory cytokines, reducing their levels in cerebrospinal fluid and plasma, while increasing the levels of anti-inflammatory cytokines. These pro-inflammatory cytokines include GM-CSF, IFN γ , CXCR3, TNF- α , IL-6, IL-10, IL-1 β , IL-8, IL-9, IL-17A, IL-17F, IL-21, and IL-22, while anti-inflammatory cytokines include IL-7, IL-4, and TGF- β . Additionally, DMTs are associated with a decline in cerebrospinal fluid levels of chemokines linked to both Th1 (CXCL9, CXCL10, CXCL11) and Th2 (CCL22) re-

sponses. DMTs that are known to cause these effects include natalizumab, alemtuzumab, fingolimod, ponesimod, teriflunomide, cladribine, IFN β -1a, IFN β -1b, DMF, and MMF [56,138,153–160] (Table 5).

3.7 CNS Inflammation, Demyelination, and Neurodegeneration

Astrocytes, microglia, oligodendrocytes, and neurons interact to control key processes in the CNS including development, inflammation, metabolism, and neuronal activity. These diverse cell types work together in complex networks to maintain homeostasis and support proper CNS function. In the context of MS, disruptions or imbalances in these intricate cell-cell interactions can contribute to the pathogenesis of the disease. Disruptions in the coordinated functions of astrocytes, microglia, oligodendrocytes, and neurons can contribute to the development and progression of the disease [47].

3.7.1 Glial Cells

The CNS comprises both neuronal and non-neuronal cells including glial cells such as astrocytes, microglia, and oligodendrocytes, which play a critical supportive role in proper functioning of the nervous system [161]. Astrocytes and microglia play an essential part in the inflammation observed in the CNS during MS. Oligodendrocytes, the myelin-producing cells, are crucial in the demyelination process that characterizes the disease [47,161].

3.7.1.1 Astrocytes. Astrocytes are crucial to CNS homeostasis, supporting neurotransmitter recycling, maintaining the BBB, and balancing ionic distributions. MS pathogenesis involves astrocytes playing either a pro- or anti-inflammatory role [161]. Some astrocytes exhibit neurotoxicity in EAE and MS, exacerbating neurodegeneration and inflammation. By contrast, others induce T cell apoptosis as a regulatory measure against inflammation and neurodegeneration. The immune actions of astrocytes vary depending on modulation by Tregs or promotion by pathogenic T cells. Astrocytes expression of S1PR affect the disease severity, demyelination, and axonal loss [162], and the dysregulated astrocyte-specific ion channels may also influence its pathogenicity [161]. Astrocyte heterogeneity in MS/EAE may provide valuable insights into the disease [161]. DMTs such as fingolimod, natalizumab, ocrelizumab, siponimod, teriflunomide, IFN β , DMF, and MMF can help reduce astrocytic inflammatory responses [163–171] (Table 5).

3.7.1.2 Microglia. Microglia, a myeloid cell in the CNS derived from the yolk sac, regulate synapse development, pruning, and excitability, acting as principal phagocytes and regulators of inflammation [161]. In MS, microglia play a significant role in remyelination, promoting oligodendrocyte precursor cells differentiation and remyelination, but this function may be impaired by age-dependent

increases in circulating TGF- β [172]. Furthermore, microglial clearance of myelin debris enhances the remyelination process. Similar to other glia cells, microglia exhibit diversity within the CNS [173]. It is a recognized fact that macrophages and microglia can cause inflammation in the CNS. However, paradoxically, these cells have also been associated with supporting functional recovery and promoting nerve regeneration. The debate surrounding the role of macrophages/microglia in the CNS is speculated to be linked to the way in which they become activated. Classically activated cells (M1 pro-inflammatory microglia) reportedly promote inflammation and demyelination, whereas alternatively activated cells (M2 anti-inflammatory microglia) appear to support functional recovery and nerve regeneration [174]. DMTs such as natalizumab, fingolimod, ocrelizumab, ofatumumab, siponimod, teriflunomide, cladribine, IFN β , DMF, MMF, and GA can help reduce microglial inflammasome activation, and promote anti-inflammatory microglial phenotype [64, 175–186] (Table 5).

3.7.1.3 Oligodendrocytes. Oligodendrocytes are the predominant cells responsible for the production of myelin and play a crucial role in the formation of myelin sheaths around nerve axons. While some oligodendrocytes may be preserved from damage during direct attacks by active T cells, severe demyelination can lead to the complete loss of these cells and a lack of remyelination. This might occur as a result of anomalies affecting OPC, an increased expression of specific immunoproteasomes in oligodendrocytes, or as a consequence of antigen-mediated immune responses launched against these cells [161]. Factors such as astrocyte signals, chondroitin sulfate proteoglycans, and hyaluronan oligomers may impede oligodendrocyte maturation and remyelination, while aged oligodendrocytes may also limit the remyelination process [161].

Recent research has suggested that individuals with MS demonstrate altered oligodendroglial heterogeneity, potentially explaining the disease progression in predisposed individuals [187]. Moreover, in MS, oligodendrocytes can function as innate immune cells that execute phagocytosis and antigen cross-presentation to cytotoxic CD8⁺ T cells, as well as activating memory and effector CD4⁺ T cells [188]. DMTs such as alemtuzumab, fingolimod, siponimod, ponesimod, teriflunomide, IFN β , and DMF can help reduce apoptosis and enhance remyelination, differentiation, and maturation of oligodendrocytes [96,189–195]. GA treatment can increase oligodendrocyte proliferation, survival, and maturation [196] (Table 5).

3.7.2 Neurons

The primary cause of clinical manifestation in MS is neuronal damage, particularly axonal damage. Various mechanisms contribute to axonal damage and are linked to ion channel dysregulation, the selective vulnerability and loss of excitatory neurons, and loss of inhibitory synap-

tic inputs [197]. Additionally, mitochondrial dysfunction, genetic predisposition, and defects in trophic and anti-inflammatory intercellular communication are involved in neuronal death [198]. However, certain DMTs have been shown to alleviate neuronal damage and promote neuron survival, axonal projections, and synaptic function. For instance, fingolimod, natalizumab, alemtuzumab, siponimod, teriflunomide, cladribine, IFN β , DMF, MMF, DRF, and GA, reportedly to enhance axonal metabolism, restore neuronal network function, normalize excitability, protect against neuronal damage, improve previous tissue damage, and/or decrease axonal and neuronal degeneration [96,189,199–207] (Table 5).

4. DMTs Unaddressed Aspects

Although researchers have made significant strides in investigating the pathophysiology of MS, it remains a complex disease that is not yet fully understood. Available DMTs target different aspects of this disease pathophysiology; nonetheless, it is unclear why these treatments alone are not sufficient to fully resolve the disease or mitigate its effects on patients.

Exploring the insufficiency of DMTs in fully resolving or mitigating the effects of MS requires consideration of various factors. These include the pathophysiology of the disease, cellular communication, the dynamic functionality of the cells, persistent autoantigen presentation, the optimal timing and mechanism of treatment, accounting for environmental factors, comorbidities, lack of diversity in DMTs clinical trials, risk-benefit profile, the potential role of pharmacogenomics in future MS treatment, the risk of administered DMTs causing new somatic mutations, the likelihood of developing anti-drug antibodies (ADA), and the wearing-off phenomenon.

4.1 Unaddressed Aspects of MS Pathophysiology

The current available DMTs for MS target various aspects of the disease pathophysiology. However, there are still key components that remain incompletely addressed by the existing DMTs. One emerging area of interest is the role of meningeal lymphatic vessels in MS pathology. Recent research has identified the meningeal lymphatic system as a new aspect of MS pathology that requires further elucidation. These meningeal lymphatic vessels may play a crucial, and potentially primary, role in the development and progression of the disease. Although the exact function of the meningeal lymphatic system in MS is still under investigation, it is possible that this component of the disease pathophysiology has been overlooked or underappreciated [99–101].

Existing DMTs may not adequately target or modulate the processes occurring within the meningeal lymphatic vessels. Continued research is needed to fully characterize the involvement of the meningeal lymphatic system in MS pathogenesis. Understanding this newly identified aspect of the disease could open up new therapeutic avenues and

highlight the limitations of current DMTs in addressing all the underlying drivers of MS. Targeting the meningeal lymphatic vessels may become an important consideration for the development of future emerging therapies.

4.2 Cellular Communication

While the current studies on DMTs for MS have provided valuable insights, they often fall short in fully addressing the role of cellular communication in the underlying pathology of the disease. The complex interactions and dynamic communication between the various cell types in the CNS, including astrocytes, oligodendrocytes, microglia, and neurons, are crucial in maintaining homeostasis and driving pathological processes [47]. However, the current understanding of how disruptions in these cellular communication networks contribute to the development and progression of MS remains limited.

Existing DMTs primarily focus on targeting specific immune mechanisms or modulating broader inflammatory pathways [11]. However, the intricacies of how the crosstalk between different CNS cell populations is altered in MS, and how this may impact disease pathogenesis, have not been extensively explored. Delving deeper into the nuances of cellular communication, signaling, and coordination within the CNS could unveil new opportunities for therapeutic interventions. Identifying key nodes or pathways in the dysregulated cellular networks may lead to the development of more targeted and comprehensive treatment strategies for MS.

4.3 The Dynamic Functionality of the Cells

The effects of DMTs on immune cells must be considered in the context of their dynamic functionality. Astrocytes and microglia, for example, can act as both pro-inflammatory and anti-inflammatory cells, depending on the specific environmental cues and signaling pathways activated. When targeting these immune cells with DMTs, it is important to take into account their multifaceted roles in the CNS. Astrocytes and microglia can play a protective, homeostatic function by clearing cellular debris and supporting neuronal health, but they can also become reactive and contribute to inflammation and tissue damage under certain conditions [161,174]. The functional derivatives of these immune cells must be carefully evaluated when designing and implementing DMT strategies. Therapies that aim to modulate astrocyte or microglial activity should consider their potential to shift the balance between pro-inflammatory and anti-inflammatory states, as this can have significant implications for disease progression and neurological outcomes. By understanding the time-dependent and context-dependent functionality of key immune cell types, researchers and clinicians can develop more targeted and personalized DMT approaches that take into account the dynamic nature of the cellular landscape in the CNS.

4.4 Persistent Autoantigen Presentation

The persistent presentation of autoantigens to the immune system is a key driver of the pathogenesis in MS. In MS, myelin proteins and other CNS components are recognized as foreign by the patient's own immune cells, leading to chronic inflammation and autoimmune attack on the brain and spinal cord [3,48,50]. This ongoing autoantigen presentation sustains the activation of autoreactive T and B cells, fueling the cyclical pattern of immune cell infiltration, demyelination, neurodegeneration, and disease exacerbations that characterize MS [3,48,50]. Importantly, the continuous availability of these CNS autoantigens for recognition by the immune system poses a significant challenge for the efficacy of current DMTs.

Many DMTs work by modulating the immune system, for example, by sequestering lymphocytes in lymphoid tissues, depleting specific immune cell subsets, or altering their trafficking and activation [11]. However, as long as the target autoantigens remain present and accessible, the autoimmune process can resume even in the face of these immunomodulatory therapies. This may help explain why, despite the availability of numerous effective DMTs, disease progression and relapses often persist in many MS patients. The underlying autoimmune response, driven by the perpetual exposure to CNS autoantigens, is not fully addressed by the current DMT approaches. Therefore, strategies to interrupt this cycle of autoantigen presentation may therefore be an important frontier in MS therapeutics. Approaches such as antigen-specific immunotherapies, which aim to induce tolerance to key myelin and neuronal components, or therapies that can clear pathogenic autoantigen deposits, may hold promise in enhancing the long-term efficacy of DMTs [47,208,209].

4.5 Optimal Timing and Mechanism of Treatment

To optimize the use of DMTs in MS treatment, physicians must consider both the disease stage and progression level. Different DMTs have varying mechanisms of action and are beneficial for different stages of the disease. In cases where immune cells are triggered in the periphery, and immune cell education with encephalitogenic potency is a concern, IFN β , GA, S1PR modulators, DMF, natalizumab, daclizumab, mitoxantrone, and cladribine may be considered [12,54,56,57,210,211]. On the other hand, for more advanced stages of the disease where neurologic recovery and remyelination are the goals, alemtuzumab, fingolimod, siponimod, ponesimod, teriflunomide, IFN β , DMF, and GA may be more appropriate [96,191–195,212,213]. Determining the disease stage and progression level can be vital in selecting the most beneficial DMT for the patient.

4.6 Accounting for Environmental Susceptibility Factors in MS

When evaluating DMTs for patients with MS, it is crucial to consider not only the direct effects of the ther-

apies, but also the broader environmental and genetic factors that can influence disease susceptibility and progression. Several key environmental factors have been implicated in MS risk and pathogenesis including prior infections, vitamin D deficiency, obesity, smoking, and disruptions to the gut microbiome. These environmental susceptibility factors can significantly impact genetic transcription and immune cell trafficking into the central nervous system, ultimately shaping the individual's disease profile and response to treatment [4,41,47]. For example, vitamin D has been shown to influence the genetic expression of numerous genes linked to MS susceptibility and immunological processes [1,41,48]. Similarly, alterations to the gut microbiome modulate immune cell trafficking, which can have downstream effects on disease activity and treatment efficacy [47].

However, current research and clinical practices surrounding DMT selection often fail to adequately address these important environmental and genetic factors. By overlooking these broader disease determinants, clinicians may be missing critical opportunities to personalize treatment approaches and optimize outcomes for patients with MS. Environmental factors, such as geographic location and sunlight exposure, play a significant role in MS epidemiology. Patients living in sunny, equatorial regions tend to have different disease characteristics and progression compared to those in countries with more pronounced seasonal variations. These environmental influences can impact an individual's risk of developing MS, as well as their response to various DMTs [1]. Similarly, genetic predisposition is a well-established determinant of MS susceptibility and phenotype. Certain genetic markers and profiles may confer differential risks and inform the optimal therapeutic approach for a given patient. However, these important genetic factors are often overlooked in the current DMT selection process [1].

Integrating the assessment of environmental susceptibility factors and their genetic and immunological implications into the DMT selection process can provide valuable insights to guide more tailored, comprehensive treatment strategies [47,214]. This holistic approach recognizes the complex interplay between a patient's genetic predisposition, environmental exposures, and the mechanism of action of the chosen DMT, ultimately enhancing the potential for successful long-term disease management.

4.7 Comorbidities in MS

People with MS often have other coexisting medical conditions, known as comorbidities. An individual with MS have higher rates of ischemic heart disease, stroke, peripheral vascular disease, and psychiatric disorders compared to the general population [215–217]. Comorbidities are especially common in underrepresented minority and immigrant groups with MS. Comorbidities impact the entire disease course. At an individual level, comorbidities are associated with more relapses, greater physical/cognitive im-

pairment, reduced quality of life, and increased mortality [215]. Notably, the incidence of comorbid autoimmune diseases seems higher in MS patients compared to healthy individuals, with up to 18% of all MS cases in the New York MS Consortium registry suffering from additional autoimmune conditions [216].

Comorbidities may develop due to shared risk factors, one condition causing another, or an underlying third condition. However, clinical trials of DMTs often exclude individuals with comorbidities, limiting the real-world applicability of trial results [215]. The most common autoimmune comorbidities in MS include rheumatoid arthritis, psoriasis, and inflammatory bowel diseases [217]. This overlap in pathogenesis between MS and other autoimmune disorders suggests that some DMTs could potentially address both disease entities, potentially simplifying treatment for these patients. For example, DMF may be a useful option for MS patients also suffering from necrobiosis lipoidica associated with diabetes or non-infectious uveitis [218,219]. Similarly, siponimod was well-tolerated and showed benefits in a phase II study of patients with refractory dermatomyositis [220]. Comorbidities can also affect DMT initiation, adherence, and effectiveness [221]. Studies show higher comorbidity burden and the presence of specific comorbidities such as anxiety or ischemic heart disease are associated with lower likelihood of starting injectable DMTs and increased risk of discontinuing DMTs such as IFN β and fingolimod [222,223]. The relationship is bidirectional, as some DMTs may also trigger new-onset comorbidities such as secondary autoimmune disorders, PML, or lymphopenia [11,15,18,19,26,215].

Understanding and managing comorbidities is crucial for optimizing outcomes in people living with MS. Prioritizing the patient's symptom profile and optimizing their medication use are essential first steps in effectively evaluating and managing comorbidities. This foundation allows the clinician to develop a comprehensive, patient-centered approach to address this challenging and often complex symptom.

4.8 Lack of Diversity of DMTs in Clinical Trials

Another concerning aspect of DMTs for MS is the lack of representation from racial and ethnic minority populations in clinical trials [224,225]. This presents a significant challenge in understanding the true effects, both beneficial and adverse, of these drugs among minority MS patients. The patient populations enrolled in pivotal DMT clinical trials have historically been predominantly white, with underrepresentation of racial and ethnic minorities who bear a disproportionate burden of MS. This lack of diversity limits the generalizability of the trial findings to the broader MS patient population, especially given the known differences in disease course, treatment response, and comorbidity profiles across racial/ethnic groups [224,225]. Without robust data from diverse clinical trial cohorts, clinicians face uncertainty when prescribing DMTs for minority patients with

MS. The effects of these therapies, including efficacy in reducing relapses and disability progression, as well as the incidence and severity of side effects, may differ significantly in non-white individuals compared to the trial populations [224,225]. This knowledge gap presents substantial barriers to delivering equitable, personalized care for racial and ethnic minority patients with MS. Addressing the lack of diversity in DMT clinical research must be a priority to ensure these powerful therapies can be utilized safely and effectively across all populations affected by this debilitating neurological condition.

4.9 Risk-Benefit Profile

The complexity of MS precludes a simple, one-size-fits-all approach to DMT selection. Instead, a careful, personalized evaluation of the risk-benefit profile for each DMT is necessary when choosing the optimal treatment for a patient. When selecting a DMT clinicians must consider a multitude of factors, including the patient's specific disease characteristics, personal preferences, presence of comorbid conditions, and reproductive plans [226]. This individualized approach is crucial to ensure the chosen DMT effectively manages the patient's MS while minimizing the potential for harm.

One such consideration is the patient's susceptibility to opportunistic infections, such as PML associated with prior JCV exposure [15]. The development of secondary autoimmune disorders or treatment-induced lymphopenia also merit careful evaluation, as these complications can significantly impact the tolerability and continued use of certain DMTs [18,19,24]. Furthermore, GI irritability should not be overlooked, as this side effect can also affect a patient's ability to adhere to and derive long-term benefit from their prescribed DMT regimen [11,26]. These types of treatment-emergent adverse events have the potential to disrupt the continuity of care, ultimately compromising the overall efficacy of the available DMT options.

Ultimately, the selection of a DMT for an individual patient with MS requires a comprehensive evaluation of the complex interplay between the patient's disease profile, comorbidities, and the unique risk-benefit characteristics of each therapeutic option. Only by employing a personalized, nuanced approach can clinicians ensure the chosen DMT strikes the delicate balance between effective disease management and minimizing potential adverse outcome.

4.10 Potential Role of Pharmacogenomics in Future MS Treatment

Personalized treatment is emerging as a potentially beneficial approach in the field of MS. Pharmacogenomics, a growing area in medicine that targets patient genetics when determining treatments, could be effective in treating MS. The disease is complex and has genetic susceptibility factors that influence progression, and variability in patient response to available DMTs can be attributed to genetics. Personalizing medicine can optimize patient response

and help deliver the best treatment options. However, this field is still in its early stages and requires further research [227,228].

4.11 New Somatic Mutations

The evidence suggests that certain DMTs used in MS treatment have the potential to induce new somatic mutations that can impact disease pathophysiology. Research indicates that fingolimod, teriflunomide, cladribine, alemtuzumab, and ocrelizumab can cause teratogenicity and embryonic lethality [229]. This leads to concerns that DMTs may affect genomic integrity and induce new mutations not present during disease development. In particular, cladribine has been found to have mutagenic effects on lymphocytes in patients with RRMS [230], while alemtuzumab has been linked to a risk of secondary autoimmunity [19]. These findings underscore the risk these drugs pose to genomic makeup. Due to these potential risks, patients taking these drugs must undergo regular monitoring for potential adverse effects. However, further research is necessary to determine the potential impact of certain DMTs on the development of new somatic mutations and how they might affect treatment outcomes.

4.12 ADAs

ADAs represents a critical factor that impacts treatment discontinuation in MS patients. When administering monoclonal antibody DMTs, which comprise chimeric, humanized, and fully human antibodies, there is the potential for therapeutic resistance from the development of binding and neutralizing antibodies. This phenomenon can occur at rates ranging from under 1% to over 80% within a year of starting treatment [231–233]. Close monitoring of ADAs is crucial to optimize treatment outcomes for patients with MS and to ensure that treatment can be adjusted as needed.

4.13 Wearing-Off Phenomenon

Recent literature has reported the wearing-off phenomena in patients using monoclonal antibody treatment, manifesting as a reduction in perceived benefits before the next dose is due, often accompanied by depression and reduced treatment satisfaction. The definition of wearing off can vary significantly among patients and clinicians, making it difficult to interpret this phenomenon accurately [234,235]. Interestingly, findings from a study indicate that patients treated with ocrelizumab, who reported wearing-off during treatment, showed reduced immunomodulation and increased neuroaxonal damage, potentially indicating a reduction in treatment response during the wearing-off period [236]. Further research is necessary to standardize the definition of wearing-off phenomena and develop strategies to address this challenge, optimizing treatment outcomes for patients with MS.

5. MS Emerging Therapies: Potential Benefits and Limitations

The current DMTs for MS have limitations in fully treating the disease, necessitating the development of new emerging therapies that aim to overcome the shortcomings of the existing DMTs. Several promising strategies are currently being explored, which may help address some of the key limitations of the current DMTs. These include therapies that target cellular communication, induce antigen-specific immune tolerance, and modulate environmental factors that contribute to MS pathogenesis [47].

Bruton's tyrosine kinase (BTK) inhibitors target B cells and peripheral myeloid cells, limiting their pathogenic crosstalk with T cells. Some BTK inhibitors can also cross the BBB, potentially suppressing detrimental microglial functions and promoting remyelination, offering hope for treating progressive MS [47,237].

Antigen-specific tolerance induction includes cell-based methods, such as administration of antigen-loaded tolerogenic DCs, autoantigen-specific regulatory T-cells, or chimeric antigen receptor T cells. However, logistical challenges limit the clinical implementation of cell-based therapies. Alternatively, nanoparticle-based, and mRNA-based approaches that deliver myelin antigens and tolerogenic adjuvants have shown promise in inducing bystander immune tolerance, an important feature considering the heterogeneity of autoantigens targeted in MS [47,208,209].

Environmental factor modulation identifies environmental factors associated with MS development, such as EBV and specific gut microbiome compositions, opening new therapeutic avenues including vaccines and probiotic supplementation. Additionally, factors such as circadian rhythms and pollution may provide insights into environmentally controlled pathways involved in MS pathogenesis [47,214]. While these emerging therapies hold significant promise, their long-term efficacy, safety, and practical implementation remain to be fully established through continued research and clinical evaluation.

6. Conclusions

The main objective of this review was to assess the pathophysiology of MS, provide an overview of the current DMTs that target each stage of disease development, and highlight the aspects of MS pathophysiology that still remain inadequately addressed by existing DMTs. Although the review indicates that the presently approved DMTs target most stages of MS pathophysiology, it also brings to light that some newly discovered aspects of the disease may either be influenced by the present DMTs or perhaps need to be addressed in future treatment options. This further emphasizes that there may be other unexplored facets of MS pathology that are not currently targeted by the available DMTs. This highlights the importance of gaining a deeper understanding of MS pathology to treat the ailment effectively.

While current DMTs have significantly improved the management of MS, this review suggests that important gaps remain in our knowledge and therapeutic armamentarium. Specifically, the review highlights the need to better understand the effects of DMTs on cellular communication, cellular dynamics, persistence of autoantigen presentation and environmental influences on disease progression. Additionally, the impact of comorbidities on treatment management, the influence of ethnic and racial factors on disease course and DMT efficacy, and the risk-benefit profiles of these therapies require further examination. The role of pharmacogenomics, the effect of DMTs on the emergence of new somatic mutations, and the potential impact of ADAs on therapies, such as humanized or chimeric antibodies, are other areas that warrant deeper exploration. Furthermore, the wearing-off phenomenon of DMT efficacy over time also deserves closer attention.

Addressing these critical knowledge gaps will be essential for developing more personalized, comprehensive, and effective treatment strategies for individuals living with MS. A deeper understanding of the multifaceted aspects of MS pathophysiology and the complex interactions among the disease, the patient, and the therapeutic interventions is necessary to optimize outcomes and overcome the remaining unmet needs in this field. Continued research and clinical investigation into these understudied areas of MS management will empower clinicians to make more informed, tailored decisions and provide patients with the most appropriate, safe, and efficacious DMT options to manage this challenging neurological condition.

Abbreviations

MS, multiple sclerosis; RRMS, relapsing-remitting MS; PPMS, primary-progressive MS; SPMS, secondary-progressive MS; PRMS, progressive-relapsing MS; CIS, clinically isolated syndrome; EAE, experimental autoimmune encephalomyelitis; CNS, central nervous system; FDA, Food and Drug Administration; EMA, European Medicines Agency; S1PR, sphingosine 1-phosphate receptor; IFN β , interferon-beta; DMF, dimethyl fumarate; MMF, monomethyl fumarate; GA, glatiramer acetate; DRF, diroximel fumarate; BBB, blood-brain barrier; ADCC, antibody-dependent cell-mediated cytotoxicity; CDC, complement-dependent cytotoxicity; ARR, annualized relapse rates; GI, gastrointestinal; PML, multifocal leukoencephalopathy; JCV, John Cunningham virus; OPC, oligodendrocyte progenitor cells; APC, antigen-presenting cell; DC, dendritic cell; Treg, regulatory T cell; FOXP3⁺, forkhead box P3⁺; TGF- β , transforming growth factor-beta; IL, interleukin; Breg, B regulatory cell; NK, natural killer; MBP, myelin basic protein; MAG, myelin-associated glycoprotein; PLP, proteolipid protein; MOG, myelin oligodendrocyte glycoprotein; P-gp, P-glycoprotein; ICAM-1, intercellular adhesion molecule-1; VCAM-1, vascular cell adhesion molecule-1; VLA-4, very late antigen-4; MMP, matrix metalloproteinase; TIMP, tissue inhibitors of met-

alloproteinase; Th, T helper; IFN γ , IFN-gamma; $\gamma\delta$ T-cell, gamma-delta T cell; ADA, anti-drug antibodies; BTK, Bruton's tyrosine kinase; EDSS, expanded disability status scale; AML, acute myeloid leukemia; HIV, human immunodeficiency virus; ITP, immune thrombocytopenic purpura; TTP, thrombotic thrombocytopenic purpura; GM-CSF, granulocyte-macrophage colony-stimulating factor; TNF- β , tumor necrosis factor-beta; ICAM-1, intercellular adhesion molecule-1; CCL2, C-C motif chemokine ligand 2; CXCL8, C-X-C motif chemokine ligand 8.

Author Contributions

EMAM confirms sole responsibility for the following aspects: the conception and design of the review, synthesis of relevant literature, and the preparation of the manuscript. EMAM read and approved the final manuscript. EMAM participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

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Conflict of Interest

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References

- [1] Mohammed EMA. Multiple sclerosis is prominent in the Gulf states: Review. *Pathogenesis*. 2016; 3: 19–38.
- [2] MS International Federation. Atlas of MS. Available at: <https://www.atlasofms.org/map/global/epidemiology/number-of-people-with-ms> (Accessed: 22 July 2024).
- [3] Dobson R, Giovannoni G. Multiple sclerosis - a review. *European Journal of Neurology*. 2019; 26: 27–40.
- [4] Brass SD, Weiner HL, Hafler DA. Multiple Sclerosis. In Rose NR, Mackay IR (eds.) *The Autoimmune Diseases* (pp. 615–632). 4th edn. Academic Press: New York. 2006.
- [5] Panitch HS, Bever CT, Jr. Clinical trials of interferons in multiple sclerosis. What have we learned? *Journal of Neuroimmunology*. 1993; 46: 155–164.
- [6] Freeman L, Longbrake EE, Coyle PK, Hendin B, Vollmer T. High-Efficacy Therapies for Treatment-Naïve Individuals with Relapsing-Remitting Multiple Sclerosis. *CNS Drugs*. 2022; 36: 1285–1299.
- [7] Montalban X, Gold R, Thompson AJ, Otero-Romero S, Amato MP, Chandraratna D, *et al.*ECTRIMS/EAN Guideline on the pharmacological treatment of people with multiple sclerosis. *Multiple Sclerosis Journal*. 2018; 24: 96–120.
- [8] Wiendl H, Gold R, Berger T, Derfuss T, Linker R, Mäurer M, *et al.* Multiple Sclerosis Therapy Consensus Group (MSTCG): position statement on disease-modifying therapies for multiple sclerosis (white paper). *Therapeutic Advances in Neurological Disorders*. 2021; 14: 17562864211039648.
- [9] Olek M, Mowry E. Initial disease-modifying therapy for relapsing-remitting multiple sclerosis in adults. Up To Date. 2023. Available at: <https://www.uptodate.com/contents/initial-disease-modifying-therapy-for-relapsing-remitting-multiple-sclerosis-in-adults#H35> (Accessed: 20 January 2024).
- [10] Freedman MS, Devonshire V, Duquette P, Giacomini PS, Giuliani F, Levin MC, *et al.* Treatment Optimization in Multiple Sclerosis: Canadian MS Working Group Recommendations. *The Canadian Journal of Neurological Sciences. Le Journal Canadien des Sciences Neurologiques*. 2020; 47: 437–455.
- [11] Amin M, Hersh CM. Updates and advances in multiple sclerosis neurotherapeutics. *Neurodegenerative Disease Management*. 2023; 13: 47–70.
- [12] Liu C, Zhu J, Mi Y, Jin T. Impact of disease-modifying therapy on dendritic cells and exploring their immunotherapeutic potential in multiple sclerosis. *Journal of Neuroinflammation*. 2022; 19: 298.
- [13] Rommer PS, Berger K, Ellenberger D, Fneish F, Simbrich A, Stahmann A, *et al.* Management of MS Patients Treated With Daclizumab - a Case Series of 267 Patients. *Frontiers in Neurology*. 2020; 11: 996.
- [14] Gallehzan NA, Khosravi M, Jamebozorgi K, Mir N, Jalilian H, Soleimanpour S, *et al.* Cost-utility and cost-effectiveness analysis of disease-modifying drugs of relapsing-remitting multiple sclerosis: a systematic review. *Health Economics Review*. 2024; 14: 12.
- [15] Khoy K, Mariotte D, Defer G, Petit G, Toutirais O, Le Mauff B. Natalizumab in Multiple Sclerosis Treatment: From Biological Effects to Immune Monitoring. *Frontiers in Immunology*. 2020; 11: 549842.
- [16] Kang C, Blair HA. Ofatumumab: A Review in Relapsing Forms of Multiple Sclerosis. *Drugs*. 2022; 82: 55–62.
- [17] Hauser SL, Kappos L, Montalban X, Craveiro L, Chognot C, Hughes R, *et al.* Safety of Ocrelizumab in Patients With Relapsing and Primary Progressive Multiple Sclerosis. *Neurology*. 2021; 97: e1546–e1559.
- [18] Ruck T, Bittner S, Wiendl H, Meuth SG. Alemtuzumab in Multiple Sclerosis: Mechanism of Action and Beyond. *International Journal of Molecular Sciences*. 2015; 16: 16414–16439.
- [19] Ruck T, Barman S, Schulte-Mecklenbeck A, Pfeuffer S, Steffen F, Nelke C, *et al.* Alemtuzumab-induced immune phenotype and repertoire changes: implications for secondary autoimmunity. *Brain: a Journal of Neurology*. 2022; 145: 1711–1725.
- [20] McGinley MP, Cohen JA. Sphingosine 1-phosphate receptor modulators in multiple sclerosis and other conditions. *Lancet (London, England)*. 2021; 398: 1184–1194.
- [21] Bar-Or A, Pachner A, Menguy-Vacheron F, Kaplan J, Wiendl H. Teriflunomide and its mechanism of action in multiple sclerosis. *Drugs*. 2014; 74: 659–674.
- [22] Järvinen E, Suomi F, Stewart JB, Guala D, Valori M, Jansson L, *et al.* Cultured lymphocytes' mitochondrial genome integrity is not altered by cladribine. *Clinical and Experimental Immunology*. 2023; 214: 304–313.
- [23] Kieseier BC. The mechanism of action of interferon- β in relapsing multiple sclerosis. *CNS Drugs*. 2011; 25: 491–502.
- [24] Mills EA, Ogrodnik MA, Plave A, Mao-Draayer Y. Emerging Understanding of the Mechanism of Action for Dimethyl Fumarate in the Treatment of Multiple Sclerosis. *Frontiers in Neurology*. 2018; 9: 5.
- [25] Berger AA, Sottosanti ER, Winnick A, Izygon J, Bernardino K, Cornett EM, *et al.* Monomethyl Fumarate (MMF, Bafiertam) for the Treatment of Relapsing Forms of Multiple Sclerosis (MS). *Neurology International*. 2021; 13: 207–223.
- [26] Singer BA, Arnold DL, Drulovic J, Freedman MS, Gold R, Gudesblatt M, *et al.* Diroximel fumarate in patients with

- relapsing-remitting multiple sclerosis: Final safety and efficacy results from the phase 3 EVOLVE-MS-1 study. *Multiple Sclerosis*. 2023; 29: 1795–1807.
- [27] McKeage K. Glatiramer Acetate 40 mg/mL in Relapsing-Remitting Multiple Sclerosis: A Review. *CNS Drugs*. 2015; 29: 425–432.
 - [28] Jacobs LD, Cookfair DL, Rudick RA, Herndon RM, Richert JR, Salazar AM, *et al.* Intramuscular interferon beta-1a for disease progression in relapsing multiple sclerosis. The Multiple Sclerosis Collaborative Research Group (MSCRG). *Annals of Neurology*. 1996; 39: 285–294.
 - [29] Kappos L, Radue EW, O'Connor P, Polman C, Hohlfeld R, Calabresi P, *et al.* A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis. *The New England Journal of Medicine*. 2010; 362: 387–401.
 - [30] Kappos L, Bar-Or A, Cree BAC, Fox RJ, Giovannoni G, Gold R, *et al.* Siponimod versus placebo in secondary progressive multiple sclerosis (EXPAND): a double-blind, randomised, phase 3 study. *Lancet (London, England)*. 2018; 391: 1263–1273.
 - [31] Comi G, Kappos L, Selmaj KW, Bar-Or A, Arnold DL, Steinman L, *et al.* Safety and efficacy of ozanimod versus interferon beta-1a in relapsing multiple sclerosis (SUNBEAM): a multicentre, randomised, minimum 12-month, phase 3 trial. *The Lancet. Neurology*. 2019; 18: 1009–1020.
 - [32] Kappos L, Fox RJ, Burcklen M, Freedman MS, Havrdová EK, Hennessy B, *et al.* Ponesimod Compared With Teriflunomide in Patients With Relapsing Multiple Sclerosis in the Active-Comparator Phase 3 OPTIMUM Study: A Randomized Clinical Trial. *JAMA Neurology*. 2021; 78: 558–567.
 - [33] Vermersch P, Czlonkowska A, Grimaldi LME, Confavreux C, Comi G, Kappos L, *et al.* Teriflunomide versus subcutaneous interferon beta-1a in patients with relapsing multiple sclerosis: a randomised, controlled phase 3 trial. *Multiple Sclerosis*. 2014; 20: 705–716.
 - [34] Fox RJ, Miller DH, Phillips JT, Hutchinson M, Havrdova E, Kita M, *et al.* Placebo-controlled phase 3 study of oral BG-12 or glatiramer in multiple sclerosis. *The New England Journal of Medicine*. 2012; 367: 1087–1097.
 - [35] Naismith RT, Wolinsky JS, Wundes A, LaGanke C, Arnold DL, Obradovic D, *et al.* Diroximel fumarate (DRF) in patients with relapsing-remitting multiple sclerosis: Interim safety and efficacy results from the phase 3 EVOLVE-MS-1 study. *Multiple Sclerosis*. 2020; 26: 1729–1739.
 - [36] Polman CH, O'Connor PW, Havrdova E, Hutchinson M, Kappos L, Miller DH, *et al.* A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *The New England Journal of Medicine*. 2006; 354: 899–910.
 - [37] Wolinsky JS, Arnold DL, Brochet B, Hartung HP, Montalban X, Naismith RT, *et al.* Long-term follow-up from the ORATORIO trial of ocrelizumab for primary progressive multiple sclerosis: a post-hoc analysis from the ongoing open-label extension of the randomised, placebo-controlled, phase 3 trial. *The Lancet. Neurology*. 2020; 19: 998–1009.
 - [38] Hauser SL, Bar-Or A, Cohen JA, Comi G, Correale J, Coyle PK, *et al.* Ofatumumab versus Teriflunomide in Multiple Sclerosis. *The New England Journal of Medicine*. 2020; 383: 546–557.
 - [39] Ziemssen T, Bass AD, Berkovich R, Comi G, Eichau S, Hobart J, *et al.* Efficacy and Safety of Alemtuzumab Through 9 Years of Follow-up in Patients with Highly Active Disease: Post Hoc Analysis of CARE-MS I and II Patients in the TOPAZ Extension Study. *CNS Drugs*. 2020; 34: 973–988.
 - [40] Giovannoni G, Comi G, Cook S, Rammohan K, Rieckmann P, Soelberg Sørensen P, *et al.* A placebo-controlled trial of oral cladribine for relapsing multiple sclerosis. *The New England Journal of Medicine*. 2010; 362: 416–426.
 - [41] Milo R, Kahana E. Multiple sclerosis: geoepidemiology, genetics and the environment. *Autoimmunity Reviews*. 2010; 9: A387–A394.
 - [42] Jakimovski D, Weinstock-Guttman B, Ramanathan M, Dwyer MG, Zivadinov R. Infections, Vaccines and Autoimmunity: A Multiple Sclerosis Perspective. *Vaccines*. 2020; 8: 50.
 - [43] Gandhi F, Jhaveri S, Avanthika C, Singh A, Jain N, Gulraiz A, *et al.* Impact of Vitamin D Supplementation on Multiple Sclerosis. *Cureus*. 2021; 13: e18487.
 - [44] Giordano A, Clarelli F, Pignolet B, Mascia E, Sorosina M, Misra K, *et al.* Vitamin D affects the risk of disease activity in multiple sclerosis. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2024; jnnp-2024-334062.
 - [45] Schreiner TG, Genes TM. Obesity and Multiple Sclerosis-A Multifaceted Association. *Journal of Clinical Medicine*. 2021; 10: 2689.
 - [46] Arnetz B. Multiple Sclerosis and Smoking. *The American Journal of Medicine*. 2020; 133: 783–788.
 - [47] Charabati M, Wheeler MA, Weiner HL, Quintana FJ. Multiple sclerosis: Neuroimmune crosstalk and therapeutic targeting. *Cell*. 2023; 186: 1309–1327.
 - [48] Mohammed EMA. Elucidating the Molecular Basis of Multiple Sclerosis and Understanding the Disease Pathophysiology. *Immunome Research*. 2016; 12: 1.
 - [49] Brynedal B, Duvefelt K, Jonasdottir G, Roos IM, Akesson E, Palmgren J, *et al.* HLA-A confers an HLA-DRB1 independent influence on the risk of multiple sclerosis. *PloS One*. 2007; 2: e664.
 - [50] Lassmann H. Multiple Sclerosis Pathology. *Cold Spring Harbor Perspectives in Medicine*. 2018; 8: a028936.
 - [51] Liu R, Du S, Zhao L, Jain S, Sahay K, Rizvanov A, *et al.* Autoreactive lymphocytes in multiple sclerosis: Pathogenesis and treatment target. *Frontiers in Immunology*. 2022; 13: 996469.
 - [52] Bar-Or A, Li R. Cellular immunology of relapsing multiple sclerosis: interactions, checks, and balances. *The Lancet. Neurology*. 2021; 20: 470–483.
 - [53] Alakhra NS, Kaplan MH. Dendritic Cells as a Nexus for the Development of Multiple Sclerosis and Models of Disease. *Advanced Biology*. 2023; 7: e2300073.
 - [54] Cauwels A, Van Lint S, Rogge E, Verhee A, Van Den Eeckhout B, Pang S, *et al.* Targeting IFN activity to both B cells and plasmacytoid dendritic cells induces a robust tolerogenic response and protection against EAE. *Scientific Reports*. 2021; 11: 21575.
 - [55] Sanna A, Fois ML, Arru G, Huang YM, Link H, Pugliatti M, *et al.* Glatiramer acetate reduces lymphocyte proliferation and enhances IL-5 and IL-13 production through modulation of monocyte-derived dendritic cells in multiple sclerosis. *Clinical and Experimental Immunology*. 2006; 143: 357–362.
 - [56] Mazzola MA, Raheja R, Regev K, Beynon V, von Glehn F, Paul A, *et al.* Monomethyl fumarate treatment impairs maturation of human myeloid dendritic cells and their ability to activate T cells. *Multiple Sclerosis*. 2019; 25: 63–71.
 - [57] Häusler D, Akgün K, Stork L, Lassmann H, Ziemssen T, Brück W, *et al.* CNS inflammation after natalizumab therapy for multiple sclerosis: A retrospective histopathological and CSF cohort study. *Brain Pathology (Zurich, Switzerland)*. 2021; 31: e12969.
 - [58] Gross CC, Ahmetspahic D, Ruck T, Schulte-Mecklenbeck A, Schwarte K, Jörgens S, *et al.* Alemtuzumab treatment alters circulating innate immune cells in multiple sclerosis. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2016; 3: e289.
 - [59] Wuest SC, Edwan JH, Martin JF, Han S, Perry JSA, Cartagena CM, *et al.* A role for interleukin-2 trans-presentation in dendritic cell-mediated T cell activation in humans, as revealed by dactilizumab therapy. *Nature Medicine*. 2011; 17: 604–609.
 - [60] Neuhaus O, Wiendl H, Kieseier BC, Archelos JJ, Hemmer B, Stüve O, *et al.* Multiple sclerosis: Mitoxantrone promotes differential effects on immunocompetent cells in vitro. *Journal of Neuroimmunology*. 2005; 168: 128–137.
 - [61] Singh V, Prajeeth CK, Gudi V, Bénardais K, Voss EV, Stangel M. 2-Chlorodeoxyadenosine (cladribine) induces apoptosis

in human monocyte-derived dendritic cells. *Clinical and Experimental Immunology*. 2013; 173: 288–297.

- [62] International Multiple Sclerosis Genetics Consortium. Multiple sclerosis genomic map implicates peripheral immune cells and microglia in susceptibility. *Science (New York, N.Y.)*. 2019; 365: eaav7188.
- [63] Watson CM, Davison AN, Baker D, O'Neill JK, Turk JL. Suppression of demyelination by mitoxantrone. *International Journal of Immunopharmacology*. 1991; 13: 923–930.
- [64] Sucksdorff M, Tuisku J, Matilainen M, Vuorimaa A, Smith S, Keitilä J, *et al.* Natalizumab treatment reduces microglial activation in the white matter of the MS brain. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2019; 6: e574.
- [65] Barbour M, Wood R, Harte T, Bushell TJ, Jiang HR. Anti-CD52 antibody treatment in murine experimental autoimmune encephalomyelitis induces dynamic and differential modulation of innate immune cells in peripheral immune and central nervous systems. *Immunology*. 2022; 165: 312–327.
- [66] Sucksdorff M, Rissanen E, Tuisku J, Nuutinen S, Paavilainen T, Rokka J, *et al.* Evaluation of the Effect of Fingolimod Treatment on Microglial Activation Using Serial PET Imaging in Multiple Sclerosis. *Journal of Nuclear Medicine: Official Publication, Society of Nuclear Medicine*. 2017; 58: 1646–1651.
- [67] Medeiros-Furquim T, Ayoub S, Johnson LJ, Aprico A, Nwoke E, Binder MD, *et al.* Cladribine Treatment for MS Preserves the Differentiative Capacity of Subsequently Generated Monocytes, Whereas Its Administration *In Vitro* Acutely Influences Monocyte Differentiation but Not Microglial Activation. *Frontiers in Immunology*. 2022; 13: 678817.
- [68] Michell-Robinson MA, Moore CS, Healy LM, Osso LA, Zorko N, Grouza V, *et al.* Effects of fumarates on circulating and CNS myeloid cells in multiple sclerosis. *Annals of Clinical and Translational Neurology*. 2015; 3: 27–41.
- [69] Pul R, Moharreg-Khiabani D, Škuljec J, Skripuletz T, Garde N, Voss EV, *et al.* Glatiramer acetate modulates TNF- α and IL-10 secretion in microglia and promotes their phagocytic activity. *Journal of Neuroimmune Pharmacology: the Official Journal of the Society on NeuroImmune Pharmacology*. 2011; 6: 381–388.
- [70] Verreycken J, Baeten P, Broux B. Regulatory T cell therapy for multiple sclerosis: Breaching (blood-brain) barriers. *Human Vaccines & Immunotherapeutics*. 2022; 18: 2153534.
- [71] Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4⁺CD25⁺ regulatory T cells in patients with multiple sclerosis. *The Journal of Experimental Medicine*. 2004; 199: 971–979.
- [72] Beliën J, Goris A, Matthys P. Natural Killer Cells in Multiple Sclerosis: Entering the Stage. *Frontiers in Immunology*. 2022; 13: 869447.
- [73] Ebrahimimofared M, Ganji A, Zahedi S, Nourbakhsh P, Ghasami K, Mosayebi G. Characterization of Regulatory T-Cells in Multiple Sclerosis Patients Treated with Interferon Beta-1a. *CNS & Neurological Disorders Drug Targets*. 2018; 17: 113–118.
- [74] Aharoni R, Eilam R, Schottlender N, Radomir L, Leistner-Segal S, Feferman T, *et al.* Glatiramer acetate increases T- and B-regulatory cells and decreases granulocyte-macrophage colony-stimulating factor (GM-CSF) in an animal model of multiple sclerosis. *Journal of Neuroimmunology*. 2020; 345: 577281.
- [75] Ferraro D, De Biasi S, Simone AM, Orlandi R, Nasi M, Vitetta F, *et al.* Modulation of Tregs and iNKT by Fingolimod in Multiple Sclerosis Patients. *Cells*. 2021; 10: 3324.
- [76] Cellerino M, Ivaldi F, Pardini M, Rotta G, Vila G, Bäcker-Koduah P, *et al.* Impact of treatment on cellular immunophenotype in MS: A cross-sectional study. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2020; 7: e693.
- [77] Wu Q, Mills EA, Wang Q, Dowling CA, Fisher C, Kirch B, *et al.* Siponimod enriches regulatory T and B lymphocytes in secondary progressive multiple sclerosis. *JCI Insight*. 2020; 5: e134251.
- [78] Sulaimani J, Cluxton D, Clowry J, Petrasca A, Molloy OE, Moran B, *et al.* Dimethyl fumarate modulates the Treg-Th17 cell axis in patients with psoriasis. *The British Journal of Dermatology*. 2021; 184: 495–503.
- [79] Paolicelli D, Ruggieri M, Manni A, Gargano CD, Carleo G, Palazzo C, *et al.* Real-Life Experience of the Effects of Cladribine Tablets on Lymphocyte Subsets and Serum Neurofilament Light Chain Levels in Relapsing Multiple Sclerosis Patients. *Brain Sciences*. 2022; 12: 1595.
- [80] Remez L, Ganelin-Cohen E, Safina D, Hellmann MA, Lotan I, Bosak N, *et al.* Alemtuzumab mediates the CD39⁺ T-regulatory cell response via CD23⁺ macrophages. *Immunology and Cell Biology*. 2021; 99: 521–531.
- [81] Haas J, Würthwein C, Korporal-Kuhnke M, Viehoveer A, Jarius S, Ruck T, *et al.* Alemtuzumab in Multiple Sclerosis: Short- and Long-Term Effects of Immunodepletion on the Peripheral Treg Compartment. *Frontiers in Immunology*. 2019; 10: 1204.
- [82] Bielekova B, Richert N, Herman ML, Ohayon J, Waldmann TA, McFarland H, *et al.* Intrathecal effects of daclizumab treatment of multiple sclerosis. *Neurology*. 2011; 77: 1877–1886.
- [83] von Essen MR, Chow HH, Holm Hansen R, Buhelt S, Sellebjerg F. Immune reconstitution following alemtuzumab therapy is characterized by exhausted T cells, increased regulatory control of proinflammatory T cells and reduced B cell control. *Frontiers in Immunology*. 2023; 14: 1249201.
- [84] Tanasescu R, Frakich N, Chou JJ, Filippini P, Podda G, Xin G, *et al.* Natalizumab Treatment of Relapsing Remitting Multiple Sclerosis Has No Long-Term Effects on the Proportion of Circulating Regulatory T Cells. *Neurology and Therapy*. 2023; 12: 2041–2052.
- [85] Gilmore W, Lund BT, Li P, Levy AM, Kelland EE, Akbari O, *et al.* Repopulation of T, B, and NK cells following alemtuzumab treatment in relapsing-remitting multiple sclerosis. *Journal of Neuroinflammation*. 2020; 17: 189.
- [86] Laurent SA, Strauli NB, Eggers EL, Wu H, Michel B, Demuth S, *et al.* Effect of Ocrelizumab on B- and T-Cell Receptor Repertoire Diversity in Patients With Relapsing Multiple Sclerosis From the Randomized Phase III OPERA Trial. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2023; 10: e200118.
- [87] Ho S, Oswald E, Wong HK, Vural A, Yilmaz V, Tüzün E, *et al.* Ocrelizumab Treatment Modulates B-Cell Regulating Factors in Multiple Sclerosis. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2023; 10: e200083.
- [88] Shinoda K, Li R, Rezk A, Mexhitaj I, Patterson KR, Kakara M, *et al.* Differential effects of anti-CD20 therapy on CD4 and CD8 T cells and implication of CD20-expressing CD8 T cells in MS disease activity. *Proceedings of the National Academy of Sciences of the United States of America*. 2023; 120: e2207291120.
- [89] Torres JB, Roodselaar J, Sealey M, Ziehn M, Bigaud M, Kneuer R, *et al.* Distribution and efficacy of ofatumumab and ocrelizumab in humanized CD20 mice following subcutaneous or intravenous administration. *Frontiers in Immunology*. 2022; 13: 814064.
- [90] Muls N, Dang HA, Sindic CJM, van Pesch V. Fingolimod increases CD39-expressing regulatory T cells in multiple sclerosis patients. *PloS One*. 2014; 9: e113025.
- [91] D'Ambrosio D, Steinmann J, Brossard P, Dingemans J. Differential effects of ponesimod, a selective S1P1 receptor modulator, on blood-circulating human T cell subpopulations. *Immunopharmacology and Immunotoxicology*. 2015; 37: 103–109.
- [92] Wu Q, Wang Q, Yang J, Mills EA, Chilukuri P, Saad A, *et al.* Teriflunomide modulates both innate and adaptive immune capacities in multiple sclerosis. *Multiple Sclerosis and Related Disorders*. 2023; 75: 104719.
- [93] Klotz L, Eschborn M, Lindner M, Liebmann M, Herold M, Janoschka C, *et al.* Teriflunomide treatment for multiple sclerosis.

- rosis modulates T cell mitochondrial respiration with affinity-dependent effects. *Science Translational Medicine*. 2019; 11: eaao5563.
- [94] Holm Hansen R, von Essen MR, Mahler MR, Cobanovic S, Binko TS, Sellebjerg F. Cladribine Effects on T and B Cells and T Cell Reactivity in Multiple Sclerosis. *Annals of Neurology*. 2023; 94: 518–530.
- [95] Rolfes L, Pfeuffer S, Huntemann N, Schmidt M, Su C, Skuljec J, *et al*. Immunological consequences of cladribine treatment in multiple sclerosis: A real-world study. *Multiple Sclerosis and Related Disorders*. 2022; 64: 103931.
- [96] Traub J, Traffehn S, Ochs J, Häusser-Kinzel S, Stephan S, Scannevin R, *et al*. Dimethyl fumarate impairs differentiated B cells and fosters central nervous system integrity in treatment of multiple sclerosis. *Brain Pathology (Zurich, Switzerland)*. 2019; 29: 640–657.
- [97] Sy M, Newton BL, Pawling J, Hayama KL, Cordon A, Yu Z, *et al*. N-acetylglucosamine inhibits inflammation and neurodegeneration markers in multiple sclerosis: a mechanistic trial. *Journal of Neuroinflammation*. 2023; 20: 209.
- [98] Häusler D, Hajiyeveva Z, Traub JW, Zamvil SS, Lalive PH, Brück W, *et al*. Glatiramer acetate immune modulates B-cell antigen presentation in treatment of MS. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2020; 7: e698.
- [99] Matthews PM. Chronic inflammation in multiple sclerosis - seeing what was always there. *Nature Reviews. Neurology*. 2019; 15: 582–593.
- [100] Alghanimy A, Work LM, Holmes WM. The glymphatic system and multiple sclerosis: An evolving connection. *Multiple Sclerosis and Related Disorders*. 2024; 83: 105456.
- [101] Tuulasvaara A, Kurdo G, Martola J, Laakso SM. Cervical lymph node diameter reflects disease progression in multiple sclerosis. *Multiple Sclerosis and Related Disorders*. 2024; 84: 105496.
- [102] Colombo E, Farina C. Lessons from S1P receptor targeting in multiple sclerosis. *Pharmacology & Therapeutics*. 2022; 230: 107971.
- [103] Burns SA, Lee Archer R, Chavis JA, Tull CA, Hensley LL, Drew PD. Mitoxantrone repression of astrocyte activation: relevance to multiple sclerosis. *Brain Research*. 2012; 1473: 236–241.
- [104] Jalousinski M, Karolczak K, Mazurek A, Glabinski A. The effects of methylprednisolone and mitoxantrone on CCL5-induced migration of lymphocytes in multiple sclerosis. *Acta Neurologica Scandinavica*. 2008; 118: 120–125.
- [105] Aglas-Leitner F, Juillard P, Juillard A, Byrne SN, Hawke S, Grau GE, *et al*. Circulating CCR6⁺ ILC proportions are lower in multiple sclerosis patients. *Clinical & Translational Immunology*. 2022; 11: e1426.
- [106] O'Sullivan S, Dev KK. Sphingosine-1-phosphate receptor therapies: Advances in clinical trials for CNS-related diseases. *Neuropharmacology*. 2017; 113: 597–607.
- [107] Chun J, Hartung HP. Mechanism of action of oral fingolimod (FTY720) in multiple sclerosis. *Clinical Neuropharmacology*. 2010; 33: 91–101.
- [108] Yen JH, Kong W, Ganea D. IFN-beta inhibits dendritic cell migration through STAT-1-mediated transcriptional suppression of CCR7 and matrix metalloproteinase 9. *Journal of Immunology (Baltimore, Md.: 1950)*. 2010; 184: 3478–3486.
- [109] Nishihara H, Perriot S, Gastfriend BD, Steinfort M, Cibien C, Soldati S, *et al*. Intrinsic blood-brain barrier dysfunction contributes to multiple sclerosis pathogenesis. *Brain: a Journal of Neurology*. 2022; 145: 4334–4348.
- [110] Pyka-Fosciak G, Lis GJ, Litwin JA. Adhesion Molecule Profile and the Effect of Anti-VLA-4 mAb Treatment in Experimental Autoimmune Encephalomyelitis, a Mouse Model of Multiple Sclerosis. *International Journal of Molecular Sciences*. 2022; 23: 4637.
- [111] Rempe RG, Hartz AMS, Bauer B. Matrix metalloproteinases in the brain and blood-brain barrier: Versatile breakers and makers. *Journal of Cerebral Blood Flow and Metabolism: Official Journal of the International Society of Cerebral Blood Flow and Metabolism*. 2016; 36: 1481–1507.
- [112] Petersen ER, Ammitzbøll C, Søndergaard HB, Oturai AB, Sørensen PS, Nilsson AC, *et al*. Expression of melanoma cell adhesion molecule-1 (MCAM-1) in natalizumab-treated multiple sclerosis. *Journal of Neuroimmunology*. 2019; 337: 577085.
- [113] Pyka-Fosciak G, Lis GJ, Litwin JA. Effect of natalizumab treatment on metalloproteinases and their inhibitors in a mouse model of multiple sclerosis. *Journal of Physiology and Pharmacology: an Official Journal of the Polish Physiological Society*. 2020; 71: 10.26402/jpp.2020.2.11.
- [114] Nguyen K, Juillard P, Hawke S, Grau GE, Marsh-Wakefield F. Trans-Endothelial Migration of Memory T Cells Is Impaired in Alemtuzumab-Treated Multiple Sclerosis Patients. *Journal of Clinical Medicine*. 2022; 11: 6266.
- [115] Lin LY, Juillard P, Hawke S, Marsh-Wakefield F, Grau GE. Oral Cladribine Impairs Intermediate, but Not Conventional, Monocyte Transmigration in Multiple Sclerosis Patients across a Model Blood-Brain Barrier. *International Journal of Molecular Sciences*. 2023; 24: 6487.
- [116] Ford RK, Juillard P, Hawke S, Grau GE, Marsh-Wakefield F. Cladribine Reduces Trans-Endothelial Migration of Memory T Cells across an In Vitro Blood-Brain Barrier. *Journal of Clinical Medicine*. 2022; 11: 6006.
- [117] Annunziata P, Cioni C, Masi G, Tassi M, Marotta G, Severi S. Fingolimod reduces circulating tight-junction protein levels and in vitro peripheral blood mononuclear cells migration in multiple sclerosis patients. *Scientific Reports*. 2018; 8: 15371.
- [118] Spampinato SF, Costantino G, Merlo S, Canonico PL, Sortino MA. Microglia Contributes to BAF-312 Effects on Blood-Brain Barrier Stability. *Biomolecules*. 2022; 12: 1174.
- [119] Spampinato SF, Merlo S, Sano Y, Kanda T, Sortino MA. Protective effect of the sphingosine-1 phosphate receptor agonist siponimod on disrupted blood brain barrier function. *Biochemical Pharmacology*. 2021; 186: 114465.
- [120] Zhao Y, Chen C, Xiao X, Fang L, Cheng X, Chang Y, *et al*. Teriflunomide Promotes Blood-Brain Barrier Integrity by Up-regulating Claudin-1 via the Wnt/ β -catenin Signaling Pathway in Multiple Sclerosis. *Molecular Neurobiology*. 2024; 61: 1936–1952.
- [121] Breuer J, Herich S, Schneider-Hohendorf T, Chasan AI, Wettschureck N, Gross CC, *et al*. Dual action by fumaric acid esters synergistically reduces adhesion to human endothelium. *Multiple Sclerosis*. 2018; 24: 1871–1882.
- [122] Hamana A, Takahashi Y, Tanioka A, Nishikawa M, Takakura Y. Amelioration of Experimental Autoimmune Encephalomyelitis in Mice by Interferon-Beta Gene Therapy, Using a Long-Term Expression Plasmid Vector. *Molecular Pharmaceutics*. 2017; 14: 1212–1217.
- [123] Cencioni MT, Mattoscio M, Magliozzi R, Bar-Or A, Muraro PA. B cells in multiple sclerosis - from targeted depletion to immune reconstitution therapies. *Nature Reviews. Neurology*. 2021; 17: 399–414.
- [124] Negron A, Stüve O, Forsthuber TG. Ectopic Lymphoid Follicles in Multiple Sclerosis: Centers for Disease Control? *Frontiers in Neurology*. 2020; 11: 607766.
- [125] Cross AH, Raine CS. Serial adoptive transfer of murine experimental allergic encephalomyelitis: successful transfer is dependent on active disease in the donor. *Journal of Neuroimmunology*. 1990; 28: 27–37.
- [126] Moser T, Akgün K, Proschmann U, Sellner J, Ziemssen T. The role of TH17 cells in multiple sclerosis: Therapeutic implications. *Autoimmunity Reviews*. 2020; 19: 102647.
- [127] Mockus TE, Munie A, Atkinson JR, Segal BM. Encephalito-

- genic and Regulatory CD8 T Cells in Multiple Sclerosis and Its Animal Models. *Journal of Immunology* (Baltimore, Md.: 1950). 2021; 206: 3–10.
- [128] Garcillán B, Marin AVM, Jiménez-Reinoso A, Briones AC, Muñoz-Ruiz M, García-León MJ, *et al.* $\gamma\delta$ T Lymphocytes in the Diagnosis of Human T Cell Receptor Immunodeficiencies. *Frontiers in Immunology*. 2015; 6: 20.
- [129] Malik S, Want MY, Awasthi A. The Emerging Roles of Gamma-Delta T Cells in Tissue Inflammation in Experimental Autoimmune Encephalomyelitis. *Frontiers in Immunology*. 2016; 7: 14.
- [130] Kaufmann M, Haase R, Proschmann U, Ziemssen T, Akgün K. Real-World Lab Data in Natalizumab Treated Multiple Sclerosis Patients Up to 6 Years Long-Term Follow Up. *Frontiers in Neurology*. 2018; 9: 1071.
- [131] Wynn D, Kaufman M, Montalban X, Vollmer T, Simon J, Elkins J, *et al.* Daclizumab in active relapsing multiple sclerosis (CHOICE study): a phase 2, randomised, double-blind, placebo-controlled, add-on trial with interferon beta. *The Lancet. Neurology*. 2010; 9: 381–390.
- [132] Ranganath T, Simpson LJ, Ferreira AM, Seiler C, Vendrame E, Zhao N, *et al.* Characterization of the Impact of Daclizumab Beta on Circulating Natural Killer Cells by Mass Cytometry. *Frontiers in Immunology*. 2020; 11: 714.
- [133] Hauser SL, Bar-Or A, Weber MS, Kletzl H, Günther A, Manfrini M, *et al.* Association of Higher Ocrelizumab Exposure With Reduced Disability Progression in Multiple Sclerosis. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2023; 10: e200094.
- [134] Feige J, Moser T, Akgün K, Schwenker K, Hitzl W, Haschke-Becher E, *et al.* Repeated iv anti-CD20 treatment in multiple sclerosis: Long-term effects on peripheral immune cell subsets. *Annals of Clinical and Translational Neurology*. 2024; 11: 450–465.
- [135] Bar-Or A, Wiendl H, Montalban X, Alvarez E, Davydovskaya M, Delgado SR, *et al.* Rapid and sustained B-cell depletion with subcutaneous ofatumumab in relapsing multiple sclerosis: APLIOS, a randomized phase-2 study. *Multiple Sclerosis*. 2022; 28: 910–924.
- [136] Blumenfeld-Kan S, Staun-Ram E, Miller A. Fingolimod reduces CXCR4-mediated B cell migration and induces regulatory B cells-mediated anti-inflammatory immune repertoire. *Multiple Sclerosis and Related Disorders*. 2019; 34: 29–37.
- [137] Kamyani D, Hassane M, Alnaqbi A, Souid AK, Rasbi ZA, Tahrawi AA, *et al.* Olanimod-mediated remission in experimental autoimmune encephalomyelitis is associated with enhanced activity of CNS CD27^{low} NK cell subset. *Frontiers in Immunology*. 2024; 15: 1230735.
- [138] Hou H, Sun Y, Miao J, Gao M, Guo L, Song X. Ponesimod modulates the Th1/Th17/Treg cell balance and ameliorates disease in experimental autoimmune encephalomyelitis. *Journal of Neuroimmunology*. 2021; 356: 577583.
- [139] von Essen MR, Talbot J, Hansen RHH, Chow HH, Lundell H, Siebner HR, *et al.* Intrathecal CD8⁺CD20⁺ T Cells in Primary Progressive Multiple Sclerosis. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2023; 10: e200140.
- [140] Holm Hansen R, Højsgaard Chow H, Christensen JR, Sellebjerg F, von Essen MR. Dimethyl fumarate therapy reduces memory T cells and the CNS migration potential in patients with multiple sclerosis. *Multiple Sclerosis and Related Disorders*. 2020; 37: 101451.
- [141] Baker D, Herrod SS, Alvarez-Gonzalez C, Zalewski L, Albor C, Schmierer K. Both cladribine and alemtuzumab may effect MS via B-cell depletion. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2017; 4: e360.
- [142] D'Angelo C, Reale M, Costantini E, Di Nicola M, Porfilio I, de Andrés C, *et al.* Profiling of Canonical and Non-Traditional Cytokine Levels in Interferon- β -Treated Relapsing-Remitting Multiple Sclerosis Patients. *Frontiers in Immunology*. 2018; 9: 1240.
- [143] Aharoni R, Eilam R, Stock A, Vainshtein A, Shezen E, Gal H, *et al.* Glatiramer acetate reduces Th-17 inflammation and induces regulatory T-cells in the CNS of mice with relapsing-remitting or chronic EAE. *Journal of Neuroimmunology*. 2010; 225: 100–111.
- [144] Groh J, Hörner M, Martini R. Teriflunomide attenuates neuroinflammation-related neural damage in mice carrying human PLP1 mutations. *Journal of Neuroinflammation*. 2018; 15: 194.
- [145] Ingelfinger F, Gerdes LA, Kavaka V, Krishnarajah S, Friebe E, Galli E, *et al.* Twin study reveals non-heritable immune perturbations in multiple sclerosis. *Nature*. 2022; 603: 152–158.
- [146] Nally FK, De Santi C, McCoy CE. Nanomodulation of Macrophages in Multiple Sclerosis. *Cells*. 2019; 8: 543.
- [147] De Bondt M, Hellings N, Opendakker G, Struyf S. Neutrophils: Underestimated Players in the Pathogenesis of Multiple Sclerosis (MS). *International Journal of Molecular Sciences*. 2020; 21: 4558.
- [148] Scutera S, Musso T, Cavalla P, Piersigilli G, Spati R, Comini S, *et al.* Inhibition of Human Neutrophil Functions In Vitro by Multiple Sclerosis Disease-Modifying Therapies. *Journal of Clinical Medicine*. 2020; 9: 3542.
- [149] Lindberg RLP, Achtnichts L, Hoffmann F, Kuhle J, Kappos L. Natalizumab alters transcriptional expression profiles of blood cell subpopulations of multiple sclerosis patients. *Journal of Neuroimmunology*. 2008; 194: 153–164.
- [150] Baker D, Kang AS, Giovannoni G, Schmierer K. Neutropenia following immune-depletion, notably CD20 targeting, therapies in multiple sclerosis. *Multiple Sclerosis and Related Disorders*. 2024; 82: 105400.
- [151] Kaegi C, Wuest B, Crowley C, Boyman O. Systematic Review of Safety and Efficacy of Second- and Third-Generation CD20-Targeting Biologics in Treating Immune-Mediated Disorders. *Frontiers in Immunology*. 2022; 12: 788830.
- [152] Hosseini A, Masjedi A, Baradaran B, Hojjat-Farsangi M, Ghalamfarsa G, Anvari E, *et al.* Dimethyl fumarate: Regulatory effects on the immune system in the treatment of multiple sclerosis. *Journal of Cellular Physiology*. 2019; 234: 9943–9955.
- [153] Diebold M, Galli E, Kopf A, Sanderson NSR, Callegari I, Benkert P, *et al.* High-dimensional immune profiling identifies a biomarker to monitor dimethyl fumarate response in multiple sclerosis. *Proceedings of the National Academy of Sciences of the United States of America*. 2022; 119: e2205042119.
- [154] Mellergård J, Edström M, Vrethem M, Ernerudh J, Dahle C. Natalizumab treatment in multiple sclerosis: marked decline of chemokines and cytokines in cerebrospinal fluid. *Multiple Sclerosis*. 2010; 16: 208–217.
- [155] Zhang X, Tao Y, Chopra M, Ahn M, Marcus KL, Choudhary N, *et al.* Differential reconstitution of T cell subsets following immunodepleting treatment with alemtuzumab (anti-CD52 monoclonal antibody) in patients with relapsing-remitting multiple sclerosis. *Journal of Immunology* (Baltimore, Md.: 1950). 2013; 191: 5867–5874.
- [156] Luessi F, Kraus S, Trinschek B, Lerch S, Ploen R, Paterka M, *et al.* FTY720 (fingolimod) treatment tips the balance towards less immunogenic antigen-presenting cells in patients with multiple sclerosis. *Multiple Sclerosis*. 2015; 21: 1811–1822.
- [157] Tilly G, Cadoux M, Garcia A, Morille J, Wiertelowski S, Pecqueur C, *et al.* Teriflunomide Treatment of Multiple Sclerosis Selectively Modulates CD8 Memory T Cells. *Frontiers in Immunology*. 2021; 12: 730342.
- [158] Balasa R, Maier S, Hutanu A, Voidazan S, Andone S, Oiaga M, *et al.* Cytokine Secretion Dynamics of Isolated PBMC after Cladribine Exposure in RRMS Patients. *International Journal of Molecular Sciences*. 2022; 23: 10262.

- [159] Stępień A, Chalimoniuk M, Lubina-Dąbrowska N, Chrapusta SJ, Galbo H, Langfort J. Effects of interferon β -1a and interferon β -1b monotherapies on selected serum cytokines and nitrite levels in patients with relapsing-remitting multiple sclerosis: a 3-year longitudinal study. *Neuroimmunomodulation*. 2013; 20: 213–222.
- [160] Ganji A, Monfared ME, Shapoori S, Nourbakhsh P, Ghazavi A, Ghasami K, *et al.* Effects of interferon and glatiramer acetate on cytokine patterns in multiple sclerosis patients. *Cytokine*. 2020; 126: 154911.
- [161] Schirmer L, Schafer DP, Bartels T, Rowitch DH, Calabresi PA. Diversity and Function of Glial Cell Types in Multiple Sclerosis. *Trends in Immunology*. 2021; 42: 228–247.
- [162] Rothhammer V, Kenison JE, Tjon E, Takenaka MC, de Lima KA, Borucki DM, *et al.* Sphingosine 1-phosphate receptor modulation suppresses pathogenic astrocyte activation and chronic progressive CNS inflammation. *Proceedings of the National Academy of Sciences of the United States of America*. 2017; 114: 2012–2017.
- [163] Smith PA, Schmid C, Zurbrugg S, Jivkov M, Doelemeyer A, Theil D, *et al.* Fingolimod inhibits brain atrophy and promotes brain-derived neurotrophic factor in an animal model of multiple sclerosis. *Journal of Neuroimmunology*. 2018; 318: 103–113.
- [164] Manocha G, Ghatak A, Puig K, Combs C. Anti- α 4 β 1 Integrin Antibodies Attenuated Brain Inflammatory Changes in a Mouse Model of Alzheimer's Disease. *Current Alzheimer Research*. 2018; 15: 1123–1135.
- [165] MacMillan EL, Schubert JJ, Vavasour IM, Tam R, Rauscher A, Taylor C, *et al.* Magnetic resonance spectroscopy evidence for declining gliosis in MS patients treated with ocrelizumab versus interferon beta-1a. *Multiple Sclerosis Journal - Experimental, Translational and Clinical*. 2019; 5: 2055217319879952.
- [166] Colombo E, Bassani C, De Angelis A, Ruffini F, Ottoboni L, Comi G, *et al.* Siponimod (BAF312) Activates Nrf2 While Hampering NF κ B in Human Astrocytes, and Protects From Astrocyte-Induced Neurodegeneration. *Frontiers in Immunology*. 2020; 11: 635.
- [167] Kabiraj P, Grund EM, Clarkson BDS, Johnson RK, LaFrance-Corey RG, Lucchinetti CF, *et al.* Teriflunomide shifts the astrocytic bioenergetic profile from oxidative metabolism to glycolysis and attenuates TNF α -induced inflammatory responses. *Scientific Reports*. 2022; 12: 3049.
- [168] Rothhammer V, Mascanfroni ID, Bunse L, Takenaka MC, Kenison JE, Mayo L, *et al.* Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor. *Nature Medicine*. 2016; 22: 586–597.
- [169] Yadav SK, Ito N, Soin D, Ito K, Dhib-Jalbut S. Dimethyl Fumarate Suppresses Demyelination and Axonal Loss through Reduction in Pro-Inflammatory Macrophage-Induced Reactive Astrocytes and Complement C3 Deposition. *Journal of Clinical Medicine*. 2021; 10: 857.
- [170] Galloway DA, Williams JB, Moore CS. Effects of fumarates on inflammatory human astrocyte responses and oligodendrocyte differentiation. *Annals of Clinical and Translational Neurology*. 2017; 4: 381–391.
- [171] Brennan MS, Matos MF, Richter KE, Li B, Scannevin RH. The NRF2 transcriptional target, OSGIN1, contributes to monomethyl fumarate-mediated cytoprotection in human astrocytes. *Scientific Reports*. 2017; 7: 42054.
- [172] Baror R, Neumann B, Segel M, Chalut KJ, Fancy SPJ, Schafer DP, *et al.* Transforming growth factor-beta renders ageing microglia inhibitory to oligodendrocyte generation by CNS progenitors. *Glia*. 2019; 67: 1374–1384.
- [173] Jordão MJC, Sankowski R, Brendecke SM, Sagar, Locatelli G, Tai YH, *et al.* Single-cell profiling identifies myeloid cell subsets with distinct fates during neuroinflammation. *Science (New York, N.Y.)*. 2019; 363: eaat7554.
- [174] Kigerl KA, Gensel JC, Ankeny DP, Alexander JK, Donnelly DJ, Popovich PG. Identification of two distinct macrophage subsets with divergent effects causing either neurotoxicity or regeneration in the injured mouse spinal cord. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2009; 29: 13435–13444.
- [175] Öhrfelt A, Axelsson M, Malmström C, Novakova L, Heslegrave A, Blennow K, *et al.* Soluble TREM-2 in cerebrospinal fluid from patients with multiple sclerosis treated with natalizumab or mitoxantrone. *Multiple Sclerosis*. 2016; 22: 1587–1595.
- [176] Shang K, He J, Zou J, Qin C, Lin L, Zhou LQ, *et al.* Fingolimod promotes angiogenesis and attenuates ischemic brain damage via modulating microglial polarization. *Brain Research*. 2020; 1726: 146509.
- [177] Anthony DC, Dickens AM, Seneca N, Couch Y, Campbell S, Checa B, *et al.* Anti-CD20 inhibits T cell-mediated pathology and microgliosis in the rat brain. *Annals of Clinical and Translational Neurology*. 2014; 1: 659–669.
- [178] Montarolo F, Martire S, Marnetto F, Valentino P, Valverde S, Capobianco MA, *et al.* The Selective Agonist for Sphingosine-1-Phosphate Receptors Siponimod Increases the Expression Level of *NR4A* Genes in Microglia Cell Line. *Current Issues in Molecular Biology*. 2022; 44: 1247–1256.
- [179] Al-Otaibi KM, Alghamdi BS, Al-Ghamdi MA, Mansouri RA, Ashraf GM, Omar UM. Therapeutic effect of combination vitamin D3 and siponimod on remyelination and modulate microglia activation in cuprizone mouse model of multiple sclerosis. *Frontiers in Behavioral Neuroscience*. 2023; 16: 1068736.
- [180] Dietrich M, Hecker C, Martin E, Langui D, Gliem M, Stankoff B, *et al.* Increased Remyelination and Proregenerative Microglia Under Siponimod Therapy in Mechanistic Models. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2022; 9: e1161.
- [181] Pol S, Sveinsson M, Sudyn M, Babek N, Siebert D, Bertolino N, *et al.* Teriflunomide's Effect on Glia in Experimental Demyelinating Disease: A Neuroimaging and Histologic Study. *Journal of Neuroimaging: Official Journal of the American Society of Neuroimaging*. 2019; 29: 52–61.
- [182] Jørgensen LØ, Hyrlov KH, Elkjaer ML, Weber AB, Pedersen AE, Svenningsen ÅF, *et al.* Cladribine modifies functional properties of microglia. *Clinical and Experimental Immunology*. 2020; 201: 328–340.
- [183] González LF, Acuña E, Arellano G, Morales P, Sotomayor P, Oyarzun-Ampuero F, *et al.* Intranasal delivery of interferon- β -loaded nanoparticles induces control of neuroinflammation in a preclinical model of multiple sclerosis: A promising simple, effective, non-invasive, and low-cost therapy. *Journal of Controlled Release: Official Journal of the Controlled Release Society*. 2021; 331: 443–459.
- [184] Tastan B, Arioz BI, Tufekci KU, Tarakcioglu E, Gonul CP, Genc K, *et al.* Dimethyl Fumarate Alleviates NLRP3 Inflammation Activation in Microglia and Sickness Behavior in LPS-Challenged Mice. *Frontiers in Immunology*. 2021; 12: 737065.
- [185] Lee YS, Gupta DP, Park SH, Yang HJ, Song GJ. Anti-Inflammatory Effects of Dimethyl Fumarate in Microglia via an Autophagy Dependent Pathway. *Frontiers in Pharmacology*. 2021; 12: 612981.
- [186] Kim HJ, Ifergan I, Antel JP, Seguin R, Duddy M, Lapierre Y, *et al.* Type 2 monocyte and microglia differentiation mediated by glatiramer acetate therapy in patients with multiple sclerosis. *Journal of Immunology (Baltimore, Md.: 1950)*. 2004; 172: 7144–7153.
- [187] Jäkel S, Agirre E, Mendanha Falcão A, van Bruggen D, Lee KW, Knuesel I, *et al.* Altered human oligodendrocyte heterogeneity in multiple sclerosis. *Nature*. 2019; 566: 543–547.
- [188] Kirby L, Jin J, Cardona JG, Smith MD, Martin KA, Wang J, *et*

- al.* Oligodendrocyte precursor cells present antigen and are cytotoxic targets in inflammatory demyelination. *Nature Communications*. 2019; 10: 3887.
- [189] Jones JL, Anderson JM, Phuah CL, Fox EJ, Selmaj K, Margolin D, *et al.* Improvement in disability after alemtuzumab treatment of multiple sclerosis is associated with neuroprotective autoimmunity. *Brain: a Journal of Neurology*. 2010; 133: 2232–2247.
- [190] Miron VE, Ludwin SK, Darlington PJ, Jarjour AA, Soliven B, Kennedy TE, *et al.* Fingolimod (FTY720) enhances remyelination following demyelination of organotypic cerebellar slices. *The American Journal of Pathology*. 2010; 176: 2682–2694.
- [191] Behrangi N, Heinig L, Frintrop L, Santrau E, Kurth J, Krause B, *et al.* Siponimod ameliorates metabolic oligodendrocyte injury via the sphingosine-1 phosphate receptor 5. *Proceedings of the National Academy of Sciences of the United States of America*. 2022; 119: e2204509119.
- [192] Safaieinejad F, Asadi S, Ghafghazi S, Niknejad H. The Synergistic Anti-Apoptosis Effects of Amniotic Epithelial Stem Cell Conditioned Medium and Ponesimod on the Oligodendrocyte Cells. *Frontiers in Pharmacology*. 2021; 12: 691099.
- [193] Göttle P, Groh J, Reiche L, Gruchot J, Rychlik N, Werner L, *et al.* Teriflunomide as a therapeutic means for myelin repair. *Journal of Neuroinflammation*. 2023; 20: 7.
- [194] Feng X, Bao R, Li L, Deisenhammer F, Arnason BGW, Reder AT. Interferon- β corrects massive gene dysregulation in multiple sclerosis: Short-term and long-term effects on immune regulation and neuroprotection. *EBioMedicine*. 2019; 49: 269–283.
- [195] Guerriero C, Puliatti G, Di Marino T, Tata AM. Effects Mediated by Dimethyl Fumarate on In Vitro Oligodendrocytes: Implications in Multiple Sclerosis. *International Journal of Molecular Sciences*. 2022; 23: 3615.
- [196] Zhang Y, Jalili F, Ouamara N, Zameer A, Cosentino G, Mayne M, *et al.* Glatiramer acetate-reactive T lymphocytes regulate oligodendrocyte progenitor cell number in vitro: role of IGF-2. *Journal of Neuroimmunology*. 2010; 227: 71–79.
- [197] Akbarian F, Rossi C, Costers L, D'hooghe MB, D'haeseleer M, Nagels G, *et al.* The spectral slope as a marker of excitation/inhibition ratio and cognitive functioning in multiple sclerosis. *Human Brain Mapping*. 2023; 44: 5784–5794.
- [198] Verma M, Lizama BN, Chu CT. Excitotoxicity, calcium and mitochondria: a triad in synaptic neurodegeneration. *Translational Neurodegeneration*. 2022; 11: 3.
- [199] Lee DH, Seubert S, Huhn K, Brecht L, Rötger C, Waschbisch A, *et al.* Fingolimod effects in neuroinflammation: Regulation of astroglial glutamate transporters? *PloS One*. 2017; 12: e0171552.
- [200] Gunnarsson M, Malmström C, Axelsson M, Sundström P, Dahle C, Vrethem M, *et al.* Axonal damage in relapsing multiple sclerosis is markedly reduced by natalizumab. *Annals of Neurology*. 2011; 69: 83–89.
- [201] Simon M, Ipek R, Homola GA, Rovituso DM, Schampel A, Kleinschnitz C, *et al.* Anti-CD52 antibody treatment depletes B cell aggregates in the central nervous system in a mouse model of multiple sclerosis. *Journal of Neuroinflammation*. 2018; 15: 225.
- [202] Gurrea-Rubio M, Wang Q, Mills EA, Wu Q, Pitt D, Tsou PS, *et al.* Siponimod Attenuates Neuronal Cell Death Triggered by Neuroinflammation via NF κ B and Mitochondrial Pathways. *International Journal of Molecular Sciences*. 2024; 25: 2454.
- [203] Malla B, Liotta A, Bros H, Ulshöfer R, Paul F, Hauser AE, *et al.* Teriflunomide Preserves Neuronal Activity and Protects Mitochondria in Brain Slices Exposed to Oxidative Stress. *International Journal of Molecular Sciences*. 2022; 23: 1538.
- [204] Schroeter CB, Rolfes L, Gothan KSS, Gruchot J, Herrmann AM, Bock S, *et al.* Cladribine treatment improves cortical network functionality in a mouse model of autoimmune encephalomyelitis. *Journal of Neuroinflammation*. 2022; 19: 270.
- [205] Hirsch M, Knight J, Tobita M, Soltys J, Panitch H, Mao Draayer Y. The effect of interferon-beta on mouse neural progenitor cell survival and differentiation. *Biochemical and Biophysical Research Communications*. 2009; 388: 181–186.
- [206] Vinnenberg L, Rychlik N, Oniani T, Williams B, White JA, Kovac S, *et al.* Assessing neuroprotective effects of diroximel fumarate and siponimod via modulation of pacemaker channels in an experimental model of remyelination. *Experimental Neurology*. 2024; 371: 114572.
- [207] Makar TK, Guda PR, Ray S, Andhavarapu S, Keledjian K, Gerzanich V, *et al.* Immunomodulatory therapy with glatiramer acetate reduces endoplasmic reticulum stress and mitochondrial dysfunction in experimental autoimmune encephalomyelitis. *Scientific Reports*. 2023; 13: 5635.
- [208] Yi J, Miller AT, Archambault AS, Jones AJ, Bradstreet TR, Bandla S, *et al.* Antigen-specific depletion of CD4⁺ T cells by CAR T cells reveals distinct roles of higher- and lower-affinity TCRs during autoimmunity. *Science Immunology*. 2022; 7: eabo0777.
- [209] Krienke C, Kolb L, Diken E, Streuber M, Kirchhoff S, Bukur T, *et al.* A noninflammatory mRNA vaccine for treatment of experimental autoimmune encephalomyelitis. *Science (New York, N.Y.)*. 2021; 371: 145–153.
- [210] Zaro BW, Vinogradova EV, Lazar DC, Blewett MM, Suci RM, Takaya J, *et al.* Dimethyl Fumarate Disrupts Human Innate Immune Signaling by Targeting the IRAK4-MyD88 Complex. *Journal of Immunology (Baltimore, Md.: 1950)*. 2019; 202: 2737–2746.
- [211] Moraes AS, Boldrini VO, Dionete AC, Andrade MD, Longhini ALF, Santos I, *et al.* Decreased Neurofilament L Chain Levels in Cerebrospinal Fluid and Tolerogenic Plasmacytoid Dendritic Cells in Natalizumab-Treated Multiple Sclerosis Patients - Brief Research Report. *Frontiers in Cellular Neuroscience*. 2021; 15: 705618.
- [212] Willems E, Schepers M, Piccart E, Wolfs E, Hellings N, Ait-Tihyaty M, *et al.* The sphingosine-1-phosphate receptor 1 modulator ponesimod repairs cuprizone-induced demyelination and induces oligodendrocyte differentiation. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 2024; 38: e23413.
- [213] Martin E, Aigrot MS, Lamari F, Bachelin C, Lubetzki C, Nait Oumesmar B, *et al.* Teriflunomide Promotes Oligodendroglial 8,9-Unsaturated Sterol Accumulation and CNS Remyelination. *Neurology(R) Neuroimmunology & Neuroinflammation*. 2021; 8: e1091.
- [214] Wheeler MA, Jaronen M, Covacu R, Zandee SEJ, Scalisi G, Rothhammer V, *et al.* Environmental Control of Astrocyte Pathogenic Activities in CNS Inflammation. *Cell*. 2019; 176: 581–596.e18.
- [215] Marrie RA, Fisk JD, Fitzgerald K, Kowalec K, Maxwell C, Rotstein D, *et al.* Etiology, effects and management of comorbidities in multiple sclerosis: recent advances. *Frontiers in Immunology*. 2023; 14: 1197195.
- [216] Chouhfeh L, Kavak KS, Teter BE, Weinstock-Guttman B. Disease modifying therapies use associated with comorbid autoimmune diseases in multiple sclerosis patients. *Multiple Sclerosis and Related Disorders*. 2015; 4: 228–233.
- [217] Konen FF, Möhn N, Witte T, Schefzyk M, Wiestler M, Lovric S, *et al.* Treatment of autoimmunity: The impact of disease-modifying therapies in multiple sclerosis and comorbid autoimmune disorders. *Autoimmunity Reviews*. 2023; 22: 103312.
- [218] Kreuter A, Knierim C, Stücker M, Pawlak F, Rotterdam S, Altmeyer P, *et al.* Fumaric acid esters in necrobiosis lipoidica: results of a prospective noncontrolled study. *The British Journal of Dermatology*. 2005; 153: 802–807.
- [219] Heinz C, Heiligenhaus A. Improvement of noninfectious uveitis with fumaric acid esters: results of a pilot study. *Archives of Ophthalmology (Chicago, Ill.: 1960)*. 2007; 125: 569–571.

- [220] Patwardhan A, Spencer CH. Biologics in refractory myositis: experience in juvenile vs. adult myositis; part II: emerging biologic and other therapies on the horizon. *Pediatric Rheumatology Online Journal*. 2019; 17: 56.
- [221] Laroni A, Signori A, Maniscalco GT, Lanzillo R, Russo CV, Binello E, *et al.* Assessing association of comorbidities with treatment choice and persistence in MS: A real-life multicenter study. *Neurology*. 2017; 89: 2222–2229.
- [222] Zhang T, Tremlett H, Leung S, Zhu F, Kingwell E, Fisk JD, *et al.* Examining the effects of comorbidities on disease-modifying therapy use in multiple sclerosis. *Neurology*. 2016; 86: 1287–1295.
- [223] Parks NE, Andreou P, Marrie RA, Fisk JD, Bhan V, Kirkland SA. Comorbidity and persistence of disease-modifying therapy use in relapsing remitting multiple sclerosis. *Multiple Sclerosis and Related Disorders*. 2021; 56: 103249.
- [224] Avasarala J. FDA-approved drugs for multiple sclerosis have no efficacy or disability data in non-Caucasian patients. *CNS Spectrums*. 2019; 24: 279–280.
- [225] Williams MJ, Okai AF, Cross AH, Monson NL, Vartanian T, Thrower BW, *et al.* Demographics and baseline disease characteristics of Black and Hispanic patients with multiple sclerosis in the open-label, single-arm, multicenter, phase IV CHIMES trial. *Multiple Sclerosis and Related Disorders*. 2023; 76: 104794.
- [226] Webb EJD, Meads D, Eskyté I, Ford HL, Bekker HL, Chataway J, *et al.* Decision Making About Disease-Modifying Treatments for Relapsing-Remitting Multiple Sclerosis: Stated Preferences and Real-World Choices. *The Patient*. 2023; 16: 457–471.
- [227] Hočevár K, Ristić S, Peterlin B. Pharmacogenomics of Multiple Sclerosis: A Systematic Review. *Frontiers in Neurology*. 2019; 10: 134.
- [228] Zarzuelo Romero MJ, Pérez Ramírez C, Carrasco Campos MI, Sánchez Martín A, Calleja Hernández MÁ, Ramírez Tortosa MC, *et al.* Therapeutic Value of Single Nucleotide Polymorphisms on the Efficacy of New Therapies in Patients with Multiple Sclerosis. *Journal of Personalized Medicine*. 2021; 11: 335.
- [229] Varytė G, Arlauskienė A, Ramašauskaitė D. Pregnancy and multiple sclerosis: an update. *Current Opinion in Obstetrics & Gynecology*. 2021; 33: 378–383.
- [230] Comi G, Cook S, Giovannoni G, Rieckmann P, Sørensen PS, Vermersch P, *et al.* Effect of cladribine tablets on lymphocyte reduction and repopulation dynamics in patients with relapsing multiple sclerosis. *Multiple Sclerosis and Related Disorders*. 2019; 29: 168–174.
- [231] Baker D, Asardag AN, Quinn OA, Efimov A, Kang AS. Anti-drug antibodies to antibody-based therapeutics in multiple sclerosis. *Human Antibodies*. 2021; 29: 255–262.
- [232] Cassotta A, Mikol V, Bertrand T, Pouzieux S, Le Parc J, Ferrari P, *et al.* A single T cell epitope drives the neutralizing anti-drug antibody response to natalizumab in multiple sclerosis patients. *Nature Medicine*. 2019; 25: 1402–1407.
- [233] Baker D, Ali L, Saxena G, Pryce G, Jones M, Schmierer K, *et al.* The Irony of Humanization: Alemtuzumab, the First, But One of the Most Immunogenic, Humanized Monoclonal Antibodies. *Frontiers in Immunology*. 2020; 11: 124.
- [234] Alvarez E, Fielding J, Clement JJ, Khazhaei D, Jimenez-Moreno C, Bushnell D, *et al.* Patient and Physician Perspectives on the Wearing-off Effect in Multiple Sclerosis: Results from Structured Interviews (P1-6.015). *Neurology*. 2024; 102: 10.1212/WNL.0000000000206116.
- [235] Marian LM, Harris KA, Conway DS. The patient-reported wearing-off phenomenon with monoclonal antibody treatments for multiple sclerosis. *Multiple Sclerosis Journal - Experimental, Translational and Clinical*. 2024; 10: 20552173241251707.
- [236] Monteiro I, Nicoletta V, Fiorenza M, Novarella F, Carotenuto A, Lanzillo R, *et al.* The ocrelizumab wearing-off phenomenon is associated with reduced immunomodulatory response and increased neuroaxonal damage in multiple sclerosis. *Journal of Neurology*. 2024; 271: 5012–5024.
- [237] Li R, Tang H, Burns JC, Hopkins BT, Le Coz C, Zhang B, *et al.* BTK inhibition limits B-cell-T-cell interaction through modulation of B-cell metabolism: implications for multiple sclerosis therapy. *Acta Neuropathologica*. 2022; 143: 505–521.