

Original Communication

Fat Distribution as a Determinant of Vitamin D Status: A Cross-Sectional Study of Adults in the United States

Yu Bai^{1,*} ¹Key Laboratory of Carcinogenesis and Translational Research (Ministry of Education), Department of Pharmacy, Peking University Cancer Hospital and Institute, 100142 Beijing, China*Correspondence: yubai_pharm@163.com (Yu Bai)

Academic Editor: Torsten Bohn

Submitted: 4 February 2024 Revised: 1 September 2024 Accepted: 8 November 2024 Published: 27 December 2024

Abstract

Background: This study aimed to elucidate correlations between obesity-related indicators and vitamin D (VD) status in a nationally representative sample of U.S. adults. **Methods:** We analysed data from 9168 adults aged 20–59 years obtained from the 2011–2018 National Health and Nutrition Examination Survey. Serum 25 hydroxyvitamin D [25(OH)D] levels were measured and categorised into quartiles. Anthropometric measurements, including weight, waist circumference, and fat mass in various body regions quantified through dual-energy X-ray absorptiometry, were collected. Multiple imputation was employed to replace missing data. The importance of obesity-related indicators and serum 25(OH)D concentration was explored using multiple linear regression adjusted for demographics, lifestyle factors, dietary intake, and clinical biomarkers, and stepwise regression. **Results:** Weight, waist circumference, and fat mass across all body regions were inversely associated with serum 25(OH)D levels (all $p < 0.001$). Notable differences were observed between men and women. Stepwise regression revealed a strong inverse correlation between visceral adipose tissue and serum 25(OH)D concentration in men [β 95% CI: -13.04 (-18.10 , -7.99), $p < 0.001$], whereas in women, only weight was significantly correlated with serum 25(OH)D concentration [β 95% CI: -0.20 (-0.28 , -0.12), $p < 0.001$]. Demographic attributes, seasonal sunlight exposure, dietary VD, calcium, phosphorus, and magnesium intake, and biomarkers including alkaline phosphatase and creatinine also emerged as significant predictors. **Conclusions:** Besides conventional obesity measures, abdominal fat metrics exhibit robust associations with VD deficiency, especially in men. Public health initiatives and clinical management strategies for hypovitaminosis D in obese populations should consider nuanced aspects of adiposity distribution alongside other demographic, lifestyle, and dietary factors influencing VD.

Keywords: vitamin D; 25-hydroxyvitamin D; obesity; NHANES; adult

1. Introduction

Obesity represents a critical public health challenge, with links to a range of morbidities including type 2 diabetes, cardiovascular diseases, chronic kidney diseases, and various cancers [1–4]. These conditions drastically diminish an individual's quality of life and considerably shorten life expectancy. The prevalence of adult obesity in the United States has surged from 14% in 1980 to 42% in 2018 [5]. By 2030, nearly half of the U.S. adult population is projected to be contending with obesity [6].

A notable concurrent issue is the frequent detection of vitamin D (VD) deficiency in individuals with obesity. Evidence supporting an inverse correlation between obesity and VD levels is growing [7–10]. Although the underpinnings of this relationship are complex, factors such as limited sunlight exposure, sedentary lifestyles, and the sequestration of VD in adipose tissue are likely significant contributors [11]. Considering the pivotal role adipocytes play in VD metabolism, VD deficiency might accelerate adipogenesis, thereby exacerbating the challenge of obesity [11].

Although body mass index (BMI) is a standard metric for assessing adiposity [12,13], it does not fully capture the nuances of body fat distribution. Emerging evidence

highlights the prognostic importance of abdominal obesity for cardiovascular and metabolic disorders, often surpassing the implications of BMI [14,15]. Alternative metrics such as waist circumference (WC), waist-to-hip ratio, and waist-to-height ratio (WHtR) provide insights into abdominal obesity [16]. Notably, adipose tissue serves as a primary reservoir for VD, which, in turn, critically influences adipocyte physiology [17–20]. Moreover, studies indicate that reducing body fat can elevate serum 25(OH)D concentrations [21–23]. In light of the escalating obesity crisis and the critical role of VD, exploring their interrelationship is imperative.

This study aimed to elucidate this correlation by comparing weight, WC, and detailed indicators of body fat distribution with serum 25(OH)D levels in a comprehensive sample of adults in the United States. We hypothesised that the primary cause of lower serum VD concentrations is the accumulation of visceral fat. Our findings aim to facilitate tailored clinical and public health strategies for mitigating obesity and addressing VD deficiency.



2. Materials and Methods

2.1 Study Population

This study utilised data from the 2011–2018 National Health and Nutrition Examination Survey (NHANES), which employed a stratified, multistage, probability sampling design to obtain representative samples of the U.S. population. Detailed survey operations and procedures have been described online (<https://www.cdc.gov/nchs/nhanes>). Briefly, NHANES surveys included an in-home interview and a subsequent health examination at a mobile examination centre (MEC). Information on demographics, lifestyle behaviours, and health conditions was collected through interviews. Physical examinations, body measurements, and laboratory tests were performed at the MEC. Dietary related information was obtained through two follow-up telephone interviews to obtain a 24-hour dietary recall.

2.2 Inclusion and Exclusion Criteria

We included people who participated in NHANES from 2011 to 2018, and the exclusion criteria were as follows: we excluded patients who were only interviewed, considering the correlation between 25 (OH)D concentration and sun exposure. We included a population aged 20–59 years who participated in the sun exposure and sun protective behaviour interviews. Moreover, dietary intake and supplements can also affect 25 (OH)D levels; hence, individuals who did not participate in dietary follow-up were excluded from the analysis. To ensure the accuracy of the dual-energy X-ray absorptiometry (DXA) scan data, only complete and valid whole-body scans were included in the study. Furthermore, participants were excluded if they wore clothes or carried medical equipment during weight measurements. Participants who did not maintain an upright posture during height measurement were also excluded. Finally, any individuals with missing data on obesity-related indicators was also excluded from the analysis. The detailed screening process is shown in Fig. 1.

2.3 Laboratory Measurement of the Serum 25(OH)D Concentration

Serum concentrations of 25(OH)D₂, 25(OH)D₃, and C3-epimer-25(OH)D₃ were measured using a fully validated, standardised, high-performance liquid chromatography-tandem mass spectrometry method (Thermo Vantage mass spectrometer and Thermo Accela ultra-high-performance liquid chromatography system, ThermoElectron Corp). The total 25(OH)D concentration was defined as the sum of 25(OH)D₂ and 25(OH)D₃, excluding C3-epimer-25(OH)D₃. C3-epimer-25(OH)D₃ is an epimer of 25(OH)D₃ and was excluded to avoid overestimating VD levels. The detection limits were constant for all analytes. It should be noted that 25(OH)D₂ detection results may be lower than the limit of detection (LOD). For analytes with results below the LOD, an imputed value of 1.45 nmol/L (the LOD divided by the square root of 2) was

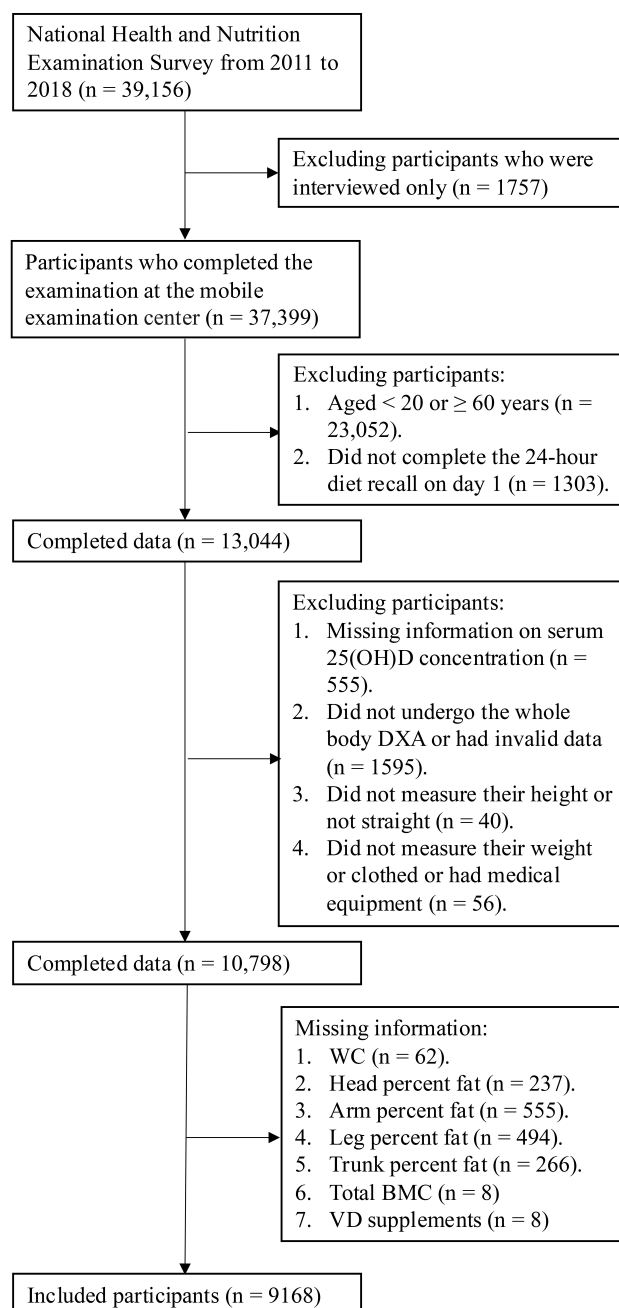


Fig. 1. Flowchart of the study population. Abbreviations: 25(OH)D, 25-hydroxyvitamin D; DXA, dual-energy X-ray absorptiometry; WC, waist circumference; BMC, bone mineral content; VD, vitamin D.

placed in the analyte results field.

2.4 Obesity-Related Indicators

BMI is an internationally recognised classification indicator for overweight and obesity, calculated by dividing weight by the square of height [24]. WC is frequently used as an indicator for evaluating abdominal obesity. Therefore, in this study, we used height, weight, and WC, along with the distribution of body fat, to evaluate the correla-

tion between obesity and 25 (OH)D serum concentration. To ensure the accuracy of our results, we excluded individuals from the NHANES data who were wearing clothes during weight measurement, carrying medical equipment, or those who failed to maintain an upright posture during height measurement, as these factors could potentially affect the results. For the measurement of human body fat, DXA is the most widely accepted method for measuring body composition due in part to its speed, ease of use, and low radiation exposure. Only data obtained from scans covering the entire body and subsequently validated were included in our analysis.

2.5 Covariates

Demographic variables included age, sex, ethnicity, education level, family income-to-poverty ratio, and examination period. Lifestyle factors comprised smoking status, alcohol consumption, time spent outdoors on weekdays and weekends, and physical activity level. Dietary intake of VD, calcium, phosphorus, and magnesium from foods and supplements were also considered as potential confounders. Additionally, biomarkers included serum calcium and phosphorus, alkaline phosphatase (ALP), alanine aminotransferase (ALT), and creatinine. Given the classic role of VD in regulating calcium and phosphorus metabolism and promoting bone development [25,26], we included dietary intake and supplementation of calcium and phosphorus, along with serum calcium and phosphorus levels, as covariates. Since ALP indicates bone formation and is involved in generating phosphorus ions and bone salt crystals [27], it was also included in the analysis as a covariate. Given that VD is metabolised in the liver and kidneys [28], serum creatinine reflects renal function [29], and ALT reflects liver function, creatinine and ALT were included as covariates.

2.6 Statistical Analysis

Serum 25(OH)D concentrations were categorised into quartiles (<49.0, 49.0–64.2, 64.3–80.3, ≥80.4 nmol/L). Demographic, lifestyle, dietary, anthropometric, and clinical characteristics across 25(OH)D quartiles were compared using Kruskal-Wallis tests for continuous variables and chi-square tests for categorical variables.

We used multiple imputation to fill in missing data. Predictive mean matching was used to impute numeric features, logistic regression to impute binary variables, and Bayesian polytomous regression to impute factor features. Simultaneously, we performed statistical analysis on both imputed and non-imputed data to evaluate the stability of the model.

We incorporated all previously mentioned covariates into the multivariable linear regression models. The models were generated to examine the associations of weight, WC, head fat, arm fat, leg fat, subcutaneous fat, and visceral adipose tissue with serum 25(OH)D levels separately. Due to

the inconsistency of evaluation index units, we standardised the data. Model 1 was adjusted for demographic characteristics, including age, sex, height, ethnicity, education level, examination period, and family income. Model 2 was further adjusted for lifestyle factors, including smoking history, alcohol consumption, sunlight exposure, and physical activity. Model 3 incorporated dietary intake and supplements (VD, calcium, magnesium, and phosphorus), bone mineral content (BMC), and biomarkers, including serum calcium and phosphorus, ALP, ALT, and creatinine. Simultaneously, we used bidirectional stepwise regression to explore which component of fat plays a key role in serum 25(OH)D levels. Stratified analysis based on sex was also performed.

All statistical analyses accounted for the complex survey design and sample weights in NHANES. Two-sided $p < 0.05$ was considered statistically significant. All analyses were performed using R version 4.2.2 (R Foundation for Statistical Computing, Vienna, Austria), with the aid of the “survey” and “mice” packages.

3. Results

3.1 Participants' Baseline Characteristics

Using imputed data from 9168 participants categorised by serum 25(OH)D levels, we stratified them into quartiles: quartile 1 (<49.0 nmol/L), quartile 2 (49.0–64.2 nmol/L), quartile 3 (64.3–80.3 nmol/L), and quartile 4 (≥80.4 nmol/L). Key demographic factors—age, sex, ethnicity, education level, and family income-to-poverty ratio—were correlated with serum 25(OH)D concentrations (all $p < 0.001$). As participants aged and as education and family income rose, 25(OH)D concentrations increased. Notably, women and non-Hispanic Whites tended to have higher 25(OH)D levels. Meanwhile, seasonal variations were conspicuous, with individuals tested in summer showing generally elevated concentrations ($p < 0.001$). Lifestyle choices influenced 25(OH)D levels: alcohol consumers had higher concentrations ($p = 0.009$), and reduced outdoor exposure corresponded with diminished 25(OH)D levels ($p < 0.001$). Anthropometric indicators, including weight, WC, and fat, sequentially decreased from quartile 1 to quartile 4 (all $p < 0.001$). Dietary patterns played a role: augmented intakes of VD, calcium, phosphorus, and magnesium correlated with heightened 25(OH)D concentrations (all $p < 0.001$). Biochemical parameters, including ALP, creatinine, and serum calcium and phosphorus, underscored pronounced differences across quartiles (all $p < 0.05$). Detailed insights from the imputed data are presented in Table 1. Preliminary findings from unimputed data largely aligned with those from imputed data (Table 2). However, while the imputed data highlighted statistical differences in ALT and physical activity across concentrations ($p < 0.05$), the unimputed dataset did not ($p \geq 0.05$).

Table 1. Baseline characteristics stratified by serum 25(OH)D quartiles using imputed NHANES data.

Characteristics	Overall	Serum 25(OH)D concentration (nmol/L)				<i>p</i> -value
		Quartile 1 (<49.0)	Quartile 2 (49.0–64.2)	Quartile 3 (64.3–80.3)	Quartile 4 (≥80.4)	
Number of participants	9168 (100.0%)	3161 (25.0%)	2397 (25.0%)	1927 (25.0%)	1683 (25.0%)	
Age (years)	38.7 (11.8)	36.2 (11.3)	37.5 (11.6)	39.4 (11.7)	41.9 (11.9)	<0.001
Male	4651 (51.7%)	1656 (54.2%)	1273 (55.6%)	1009 (54.2%)	713 (42.9%)	<0.001
Ethnicity						<0.001
Mexican American	1402 (11.6%)	581 (19.3%)	462 (15.3%)	258 (8.5%)	101 (3.3%)	
Other Hispanic	967 (7.4%)	283 (8.7%)	321 (9.3%)	230 (7.3%)	133 (4.3%)	
Non-Hispanic White	3240 (59.7%)	484 (30.1%)	808 (56.7%)	907 (69.9%)	1041 (82.7%)	
Non-Hispanic Black	1841 (10.7%)	1139 (26.9%)	360 (8.3%)	198 (4.4%)	144 (3.1%)	
Other Ethnicity	1718 (10.5%)	674 (15.0%)	446 (10.4%)	334 (9.8%)	264 (6.7%)	
Education level						<0.001
≤ high school	3626 (34.3%)	1380 (42.7%)	972 (34.9%)	716 (31.7%)	558 (28.1%)	
Above high school	5540 (65.6%)	1780 (57.2%)	1425 (65.1%)	1210 (68.3%)	1125 (71.9%)	
Unknown	2 (0.0%)	1 (0.1%)	0 (0.0%)	1 (0.0%)	0 (0.0%)	
Examination period						<0.001
November–April	4545 (45.3%)	1906 (62.2%)	1144 (45.5%)	841 (40.1%)	654 (33.2%)	
May–October	4623 (54.7%)	1255 (37.8%)	1253 (54.5%)	1086 (59.9%)	1029 (66.8%)	
Family income-to-poverty ratio	2.9 (1.7)	2.4 (1.6)	2.8 (1.7)	3.1 (1.7)	3.4 (1.6)	<0.001
Drinking	7685 (87.6%)	2574 (84.7%)	2010 (86.7%)	1636 (89.0%)	1465 (90.1%)	0.009
Smoking	3571 (40.6%)	1160 (38.1%)	907 (38.6%)	798 (43.8%)	706 (41.8%)	0.200
Stay in the shade						0.028
Always, most of the time, or sometimes	6904 (73.2%)	2468 (77.0%)	1826 (74.0%)	1395 (71.5%)	1215 (70.2%)	
Rarely or never	2261 (26.8%)	691 (23.0%)	570 (25.9%)	532 (28.5%)	468 (29.8%)	
Unknown	3 (0.0%)	2 (0.1%)	1 (0.0%)	0 (0.0%)	0 (0.0%)	
Outdoors work day (minutes)	104.1 (132.3)	91.8 (123.8)	100.2 (132.1)	118.3 (141.5)	106.3 (130.0)	<0.001
Outdoors, not work day (minutes)	155.5 (127.5)	126.3 (119.6)	147.3 (123.3)	177.9 (131.7)	170.8 (128.6)	<0.001
Physical activity (kcal/day)	1639.0 (4536.5)	1630.9 (3230.1)	1661.8 (3075.1)	1644.8 (3389.2)	1618.3 (7144.5)	0.046
Height (cm)	169.0 (9.5)	168.1 (9.7)	168.8 (9.5)	170.0 (9.7)	169.3 (9.2)	0.001
Weight (kg)	81.6 (19.8)	84.8 (22.3)	82.5 (19.1)	81.9 (19.2)	77.0 (17.5)	<0.001
BMI (kg/m ²)	28.5 (6.3)	30.0 (7.3)	28.9 (6.1)	28.3 (6.1)	26.8 (5.4)	<0.001
WC (cm)	96.9 (15.5)	99.7 (17.4)	97.9 (15.1)	96.8 (14.9)	93.3 (13.8)	<0.001
WHtR	0.57 (0.09)	0.59 (0.10)	0.58 (0.09)	0.57 (0.09)	0.55 (0.08)	<0.001
Total BMC (g)	2360.2 (441.2)	2361.8 (444.3)	2341.7 (413.8)	2404.7 (456.9)	2333.0 (445.5)	0.007
Head fat (g)	1164.9 (164.8)	1201.3 (170.7)	1178.3 (162.2)	1166.1 (165.7)	1113.4 (146.9)	<0.001
Arms fat (g)	3303.8 (1523.9)	3588.6 (1757.2)	3350.6 (1476.6)	3228.7 (1446.4)	3044.2 (1329.5)	<0.001
Legs fat (g)	9611.3 (4071.3)	10,278.2 (4592.0)	9602.9 (4000.6)	9426.7 (3889.5)	9130.9 (3653.4)	<0.001
SAT (g)	1585.6 (776.8)	1738.4 (877.1)	1625.1 (759.9)	1539.3 (746.4)	1438.1 (678.0)	<0.001
VAT (g)	489.9 (272.3)	517.3 (282.6)	511.0 (263.0)	488.5 (268.6)	442.6 (268.5)	<0.001

Table 1. Continued.

Characteristics	Overall	Serum 25(OH)D concentration (nmol/L)				<i>p</i> -value
		Quartile 1 (<49.0)	Quartile 2 (49.0–64.2)	Quartile 3 (64.3–80.3)	Quartile 4 (≥80.4)	
Dietary intake						
Vitamin D (µg/day)	4.5 (4.6)	3.8 (3.9)	4.5 (4.7)	4.9 (4.6)	4.7 (4.8)	< 0.001
Calcium (mg/day)	994.5 (531.8)	906.9 (526.0)	970.7 (501.7)	1072.0 (551.6)	1029.7 (532.5)	< 0.001
Phosphorus (mg/day)	1433.4 (617.6)	1372.8 (620.1)	1420.9 (609.4)	1504.1 (636.1)	1436.7 (597.5)	< 0.001
Magnesium (mg/day)	310.6 (139.7)	283.7 (130.5)	303.9 (137.2)	329.0 (146.8)	326.1 (139.4)	< 0.001
Dietary supplements						
Vitamin D (µg/day)	9.7 (41.6)	1.8 (13.2)	4.3 (19.2)	8.7 (32.4)	24.2 (71.1)	< 0.001
Calcium (mg/day)	92.7 (228.4)	31.8 (126.3)	61.0 (161.4)	99.4 (211.8)	179.3 (331.5)	< 0.001
Phosphorus (mg/day)	4.3 (27.3)	1.0 (9.0)	3.2 (19.6)	4.4 (27.7)	8.5 (41.5)	< 0.001
Magnesium (mg/day)	21.0 (70.4)	7.2 (44.2)	13.5 (46.8)	23.9 (68.0)	39.5 (102.5)	< 0.001
Biomarkers						
Alkaline phosphatase (U/L)	66.5 (21.5)	69.6 (23.1)	67.4 (22.9)	65.2 (19.2)	63.6 (20.1)	< 0.001
Alanine aminotransferase (U/L)	26.3 (20.2)	28.0 (22.6)	26.8 (19.8)	25.6 (20.8)	24.9 (17.3)	0.048
Creatinine (µmol/L)	76.0 (26.4)	74.7 (28.4)	74.5 (17.7)	75.8 (16.5)	78.8 (37.2)	< 0.001
Serum calcium (mmol/L)	2.35 (0.08)	2.34 (0.09)	2.34 (0.09)	2.35 (0.08)	2.35 (0.08)	< 0.001
Serum phosphorus (mmol/L)	1.21 (0.18)	1.20 (0.18)	1.21 (0.18)	1.21 (0.18)	1.22 (0.18)	0.017

Abbreviations: 25(OH)D, 25-hydroxyvitamin D; BMI, body mass index; WC, waist circumference; WHtR, waist-to-height ratio; BMC, Bone Mineral Content; SAT, subcutaneous adipose tissue; VAT, visceral adipose tissue.

Note: Continuous variables are described as means with standard deviations. Categorical variables are presented as unweighted sample sizes and weighted proportions. All estimates were adjusted to account for the complex survey design by incorporating dietary weight. Non-normally distributed data were analysed using the Kruskal-Wallis test, whereas normally distributed data were tested using the analysis of variance. Categorical variables were analysed using the Chi-square test. Family income-to-poverty ratio was calculated by dividing family (or individual) income by the poverty guidelines specific to the survey year. Bold *p*-values indicate statistical differences.

Table 2. Baseline characteristics stratified by serum 25(OH)D quartiles using unimputed NHANES data.

Characteristics	Overall	Serum 25(OH)D concentration (nmol/L)				<i>p</i> -value
		Quartile 1 (<49.0)	Quartile 2 (49.0–64.2)	Quartile 3 (64.3–80.3)	Quartile 4 (≥80.4)	
Family income-to-poverty ratio (n = 8480)	2.9 (1.7)	2.4 (1.6)	2.8 (1.7)	3.2 (1.7)	3.4 (1.6)	< 0.001
Outdoors work day (minutes) (n = 9152)	104.1 (132.3)	91.7 (123.6)	100.2 (132.1)	118.2 (141.5)	106.4 (130.0)	< 0.001
Outdoors, not work day (minutes) (n = 9144)	155.5 (127.5)	126.3 (119.5)	147.4 (123.3)	177.9 (131.7)	171.0 (128.6)	< 0.001
Physical activity (kcal/day) (n = 9135)	1633.8 (4539.7)	1630.9 (3233.1)	1652.2 (3066.9)	1636.5 (3388.7)	1615.6 (7154.8)	0.059
Alkaline phosphatase (U/L) (n = 9093)	66.5 (21.5)	69.6 (23.1)	67.5 (23.0)	65.2 (19.2)	63.6 (20.1)	< 0.001
Alanine aminotransferase (U/L) (n = 9092)	26.3 (20.2)	28.0 (22.6)	26.8 (19.8)	25.6 (20.8)	24.9 (17.3)	0.050
Creatinine (μmol/L) (n = 9095)	76.0 (26.4)	74.8 (28.5)	74.5 (17.7)	75.9 (16.5)	78.9 (37.2)	< 0.001
Calcium (mmol/L) (n = 9076)	2.35 (0.08)	2.34 (0.09)	2.34 (0.09)	2.35 (0.08)	2.35 (0.08)	< 0.001
Phosphorus (mmol/L) (n = 9094)	1.21 (0.18)	1.20 (0.18)	1.21 (0.18)	1.21 (0.18)	1.22 (0.18)	0.018

Abbreviation: 25(OH)D, 25-hydroxyvitamin D.

Note: Data that did not need to be imputed is not displayed again. Continuous variables are described as means with standard deviations. All the estimates were adjusted to account for the complex survey design by incorporating dietary weight. Non-normally distributed data were analysed using the Kruskal-Wallis test, whereas normally distributed data were analysed using analysis of variance. Family income-to-poverty ratio was calculated by dividing family (or individual) income by the poverty guidelines specific to the survey year. Bold *p*-values indicate statistical differences.

Table 3. Associations of factors with serum 25(OH)D concentrations: multiple stepwise regression analysis of imputed NHANES data.

Characteristics	Overall		Male		Female	
	β (95% CI)	<i>p</i> -value	β (95% CI)	<i>p</i> -value	β (95% CI)	<i>p</i> -value
Age (year)	0.32 (0.22, 0.42)	< 0.001	0.40 (0.29, 0.51)	< 0.001	0.26 (0.14, 0.38)	< 0.001
Ethnicity						
Mexican American			Reference			
Other Hispanic	4.07 (1.40, 6.74)	0.003	3.10 (−0.11, 6.30)	0.058	5.28 (1.58, 8.98)	0.005
Non-Hispanic White	11.08 (9.01, 13.16)	< 0.001	9.76 (7.24, 12.27)	< 0.001	13.22 (10.25, 16.20)	< 0.001
Non-Hispanic Black	−12.58 (−15.03, −10.13)	< 0.001	−16.13 (−19.03, −13.23)	< 0.001	−8.44 (−11.76, −5.12)	< 0.001
Other Race	−0.34 (−2.89, 2.21)	0.792	−1.41 (−4.49, 1.67)	0.369	1.26 (−2.02, 4.55)	0.451
Examination period						
November–April			Reference			
May–October	6.58 (4.70, 8.45)	< 0.001	5.85 (3.86, 7.84)	< 0.001	7.22 (4.71, 9.73)	< 0.001
Family income-to-poverty ratio	0.95 (0.49, 1.42)	< 0.001	0.59 (0.07, 1.11)	0.026	1.40 (0.60, 2.20)	0.001
Outdoors work day (seconds)	0.83 (0.56, 1.11)	< 0.001	0.76 (0.49, 1.04)	< 0.001	0.82 (0.15, 1.49)	0.016

Table 3. Continued.

Characteristics	Overall		Male		Female	
	β (95% CI)	<i>p</i> -value	β (95% CI)	<i>p</i> -value	β (95% CI)	<i>p</i> -value
Outdoors not work day (seconds)	0.81 (0.50, 1.11)	<0.001	0.71 (0.39, 1.03)	<0.001	0.91 (0.34, 1.49)	0.002
Vitamin D intake ($\mu\text{g/d}$)	0.33 (0.12, 0.53)	0.002	0.41 (0.19, 0.63)	<0.001	0.25 (−0.13, 0.62)	0.196
Magnesium intake ($\mu\text{g/d}$)	17.30 (8.71, 25.89)	<0.001	11.99 (4.13, 19.85)	0.003	22.89 (7.73, 38.06)	0.003
Phosphorus intake ($\mu\text{g/d}$)	−3.95 (−6.09, −1.81)	<0.001	−3.06 (−5.33, −0.80)	0.008	−4.56 (−9.93, 0.81)	0.096
Vitamin D supplement intake ($\mu\text{g/d}$)	0.13 (0.08, 0.19)	<0.001	0.21 (0.09, 0.32)	0.001	0.11 (0.06, 0.17)	<0.001
Calcium supplement intake ($\mu\text{g/d}$)	17.21 (11.99, 22.42)	<0.001	4.51 (−3.66, 12.68)	0.279	19.84 (13.57, 26.11)	<0.001
Magnesium supplement intake ($\mu\text{g/d}$)	5.07 (−9.81, 19.96)	0.504	19.23 (−13.16, 51.62)	0.244	4.20 (−13.96, 22.35)	0.651
Creatinine ($\mu\text{mol/L}$)	0.09 (0.04, 0.14)	<0.001	0.08 (0.03, 0.13)	0.002	0.14 (0.04, 0.23)	0.005
Serum Calcium (mmol/L)	20.74 (10.55, 30.93)	<0.001	23.18 (13.00, 33.36)	<0.001	19.36 (2.77, 35.95)	0.022
Total BMC (mg)	5.23 (2.61, 7.84)	<0.001	4.84 (2.25, 7.43)	<0.001	3.30 (−1.77, 8.37)	0.202
Head fat (mg)	−9.24 (−16.46, −2.03)	0.012	−4.91 (−16.28, 6.46)	0.397	−11.02 (−23.48, 1.45)	0.083
Sex						
Male			Reference			
Female	10.54 (7.66, 13.41)	<0.001	NG		NG	
Physical activity (cal/d)	0.09 (−0.04, 0.23)	0.179	0.14 (0.04, 0.23)	0.006	NG	
Subcutaneous fat (mg)	−3.40 (−4.93, −1.88)	<0.001	−4.69 (−8.76, −0.61)	0.024	NG	
Visceral adipose tissue (mg)	−6.74 (−12.66, −0.81)	0.026	−13.04 (−18.10, −7.99)	<0.001	NG	
Stay in the shade						
Always, most of the time or sometimes			Reference			
Rarely or never	1.32 (−0.43, 3.07)	0.139	NG		2.88 (0.13, 5.63)	0.040
Unknown	−2.57 (−5.83, 0.69)	0.122	NG		−1.18 (−6.20, 3.84)	0.646
Calcium intake ($\mu\text{g/d}$)	1.53 (−0.99, 4.05)	0.235	NG		3.04 (−1.82, 7.90)	0.220
Alkaline phosphatase (IU/L)	−0.06 (−0.09, −0.02)	0.001	NG		−0.12 (−0.17, −0.07)	<0.001
Height (cm)	NG		−0.13 (−0.36, 0.09)	0.246	0.24 (0.00, 0.48)	0.055
Weight (kg)	NG		0.20 (−0.02, 0.42)	0.068	−0.20 (−0.28, −0.12)	<0.001
Arms fat (mg)	NG		−1.36 (−3.26, 0.54)	0.159		
Phosphorus (mmol/L)	NG		−2.84 (−7.17, 1.49)	0.198		
Education level						
Less than or equal to high school			Reference			
Above high school	NG		NG		−1.88 (−4.69, 0.92)	0.188
Unknown	NG		NG		6.89 (2.90, 10.87)	0.001

Abbreviations: BMC, bone mineral content; SAT, subcutaneous adipose tissue; VAT, visceral adipose tissue; NG, not given.

Note: Family income-to-poverty ratio was calculated by dividing family (or individual) income by the poverty guidelines specific to the survey year. Bold *p*-values indicate statistical differences.

3.2 Correlation between Obesity Indicators and Serum 25(OH)D Concentration

Fig. 2 delineates the outcomes of multiple linear regression analyses in the general population, and the male and female subsets, which explored the correlation between distinct metrics—namely weight, WC, and fat in different parts of the body—and serum 25(OH)D levels. Adjustments were made and all indicators of the adjusted model showed statistical significance (all $p < 0.05$). In the comprehensive model using interpolated data, the relatively superior evaluation indicators include weight, WC, and subcutaneous fat [weight: β 95% CI: -5.12 ($-6.00, -4.24$), pseudo R^2 : 0.408, AIC: 75,985.36; WC: β 95% CI: -4.67 ($-5.45, -3.90$), pseudo R^2 : 0.408, AIC: 75,980.16; subcutaneous fat: β 95% CI: -4.76 ($-5.54, -3.99$), pseudo R^2 : 0.407, AIC: 75,990.86, respectively]. Among men, WC was the superior index [β 95% CI: -4.42 ($-5.44, -3.41$), pseudo R^2 : 0.400, AIC: 37,487.01], while weight was a stronger predictor in women [β 95% CI: -4.92 ($-6.19, -3.65$), pseudo R^2 : 0.425, AIC: 38,323.89]. Regarding fat distribution, visceral adipose tissue volume in men showed a stronger correlation with serum 25(OH)D levels [β 95% CI: -4.67 ($-5.70, -3.64$), pseudo R^2 : 0.403, AIC: 37,466.70], whereas subcutaneous fat volume was more closely associated with serum 25(OH)D levels in women [β 95% CI: -3.95 ($-5.06, -2.58$), pseudo R^2 : 0.422, AIC: 38,347.76]. Consistency was observed between results derived from the imputed and non-imputed data.

To comprehensively consider the effects of body fat composition on serum 25(OH)D levels, we conducted a rigorous bidirectional stepwise regression analysis on the entire population, and the male and female subpopulations. Head fat, visceral adipose tissue, and subcutaneous fat were independent determinants in the entire population [head fat: β 95% CI: -9.24 ($-16.46, -2.03$), $p = 0.012$; visceral adipose tissue: β 95% CI: -6.74 ($-12.66, -0.81$), $p = 0.026$; subcutaneous fat: β 95% CI: -3.40 ($-4.93, -1.88$), $p < 0.001$, respectively]. In men, a strong inverse correlation was observed between serum 25(OH)D concentrations and both visceral adipose tissue and subcutaneous fat, with visceral adipose tissue having a more pronounced impact [visceral adipose tissue: β 95% CI: -13.04 ($-18.10, -7.99$), $p < 0.001$; subcutaneous fat: β 95% CI: -4.69 ($-8.76, -0.16$), $p = 0.024$]. In contrast, no significant correlation was observed between fat distribution and serum 25(OH)D levels in women, although weight was identified as an important influencing factor [β 95% CI: -0.20 ($-0.28, -0.12$), $p < 0.001$]. Additionally, demographic factors like age, sex, ethnicity, examination period, and family income; lifestyle aspects including outdoor duration; dietary elements encompassing dietary intake of VD, phosphorus, and magnesium; and supplementary intake of VD and calcium are intricately linked with 25(OH)D. Notably, biomarkers such as ALP, creatinine ($\mu\text{mol/L}$), and serum calcium (mmol/L)

also share a close association with 25(OH)D. Please refer to Table 3 for a detailed overview.

4. Discussion

Our analysis of a nationally representative, contemporary sample of adults in the United States affirms an inverse association between body fat and serum 25(OH)D levels. While previous studies have highlighted the relationship between obesity and 25 (OH)D concentration using BMI [30,31], it is acknowledged that BMI does not capture nuances in fat distribution. This study expands this correlation to distinct facets of adiposity distribution. In the linear regression model adjusting for demographics, lifestyle, diet, biomarkers, and BMC, weight, WC, and regional fat depots (arms, legs, head, subcutaneous, and visceral) all demonstrate robust inverse relationships with 25(OH)D, aligning with previous evidence [32,33]. In the stepwise regression analysis, subcutaneous and visceral fat emerged as significant predictors of serum 25(OH)D levels, particularly in the male subpopulation, while weight was significantly correlated with serum 25(OH)D levels in women. These results suggest that both sex and body fat distribution play critical roles in determining the VD status.

The mechanistic basis underlying the relationship between fat and hypovitaminosis D warrants discussion. Adipose tissue serves as a major reservoir for VD and its metabolites, as evidenced by an animal study showing 75% of supplemented VD distributed to fat in pigs, with 35% as 25(OH)D, 30% in serum, 20% in muscle, and 15% elsewhere [18]. Furthermore, clinical trials have demonstrated that vitamin D3 supplementation increases 25(OH)D3 levels in both serum and subcutaneous fat [19,20]. The sequestration of VD in fat likely contributes to the comorbidity of VD deficiency and obesity [7]. However, the differential association of specific fat depots with VD remains unclear and merits further interrogation.

While adiposity loss increases 25(OH)D [21–23], whether greater fat accretion reciprocally lowers 25(OH)D remains uncertain. VD slowly mobilises from fat into circulation [34]; however, the precise mechanisms are incompletely elucidated. Notably, all VD hydroxylation enzymes are expressed in adipocytes and adipose tissue, implying potential tissue-specific regulation [17,35]. The differential expression and activity of metabolic enzymes across fat depots may thus drive the divergent associations observed here with VD. In obesity, adipose dysfunction occurs, characterised by hypertrophied adipocytes, inflammation, hypoxia, and reduced angiogenesis [36]. The nuclear VD receptor, abundant in adipocytes, mediates VD activities, influencing adipokines, energy metabolism, inflammation, oxidative stress, differentiation, and apoptosis [17]. VD deficiency could thereby disrupt adipocyte function [17,36,37]. Therefore, it is important to pay attention to VD levels in people with obesity. Our findings highlight the importance of visceral adipose tissue and subcutaneous fat, especially in men.

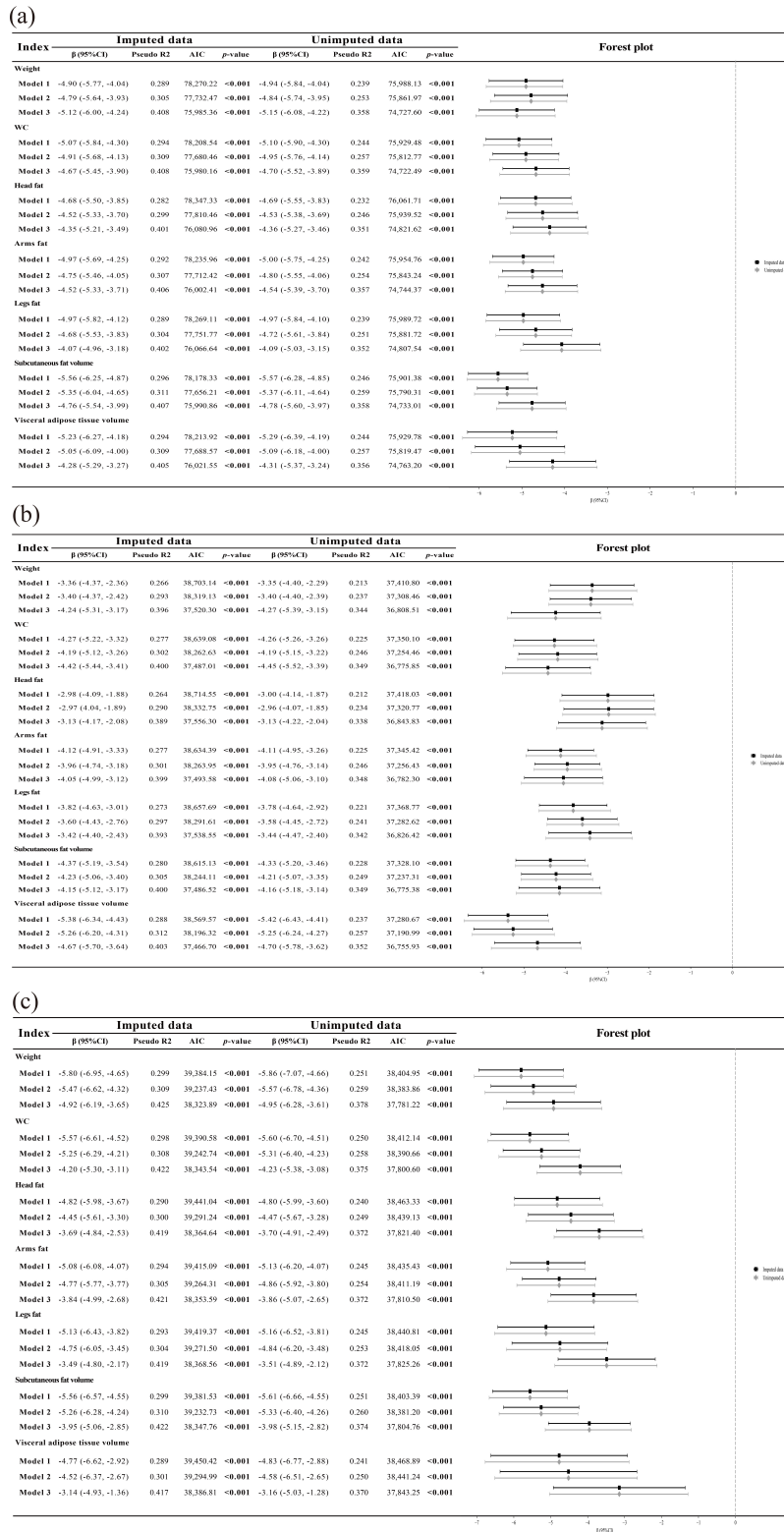


Fig. 2. Association between obesity indicators and serum 25(OH)D in total population (a), and male (b), and female subpopulations (c): Forest plot from multiple linear regression. Model 1 adjusted for demographic characteristics, including age, sex, height, ethnicity, education level, examination period and family income. Model 2 further adjusted for lifestyle factors, including smoking history, alcohol consumption, sunlight exposure, and physical activity. Model 3 incorporated dietary intake and supplements (VD, calcium, magnesium, and phosphorus), bone mineral content, and biomarkers including serum calcium and phosphorus, alkaline phosphatase, alanine aminotransferase, and creatinine.

Beyond adiposity, our analysis spotlights diverse facets intricately linked to 25(OH)D status, including demographic attributes, seasonal/light exposure, dietary patterns, and select biomarkers, which demand tailored public health and clinical considerations. Unsurprisingly, reduced outdoor exposure on weekdays and weekends was linked to lower 25(OH)D levels in our study. Surprisingly, physical activity did not emerge as an independent predictor in our stepwise regression model. Sedentary lifestyles probably overlap with limited sun exposure. Promoting outdoor activities could offer synergistic benefits by boosting activity levels, light exposure, and VD status. Moreover, dietary and supplementary intakes of VD, calcium, phosphorus, and magnesium positively influenced serum 25(OH)D. Clinicians should carefully assess intakes of VD, calcium, magnesium, and phosphorus and recommend balanced nutritional support alongside targeted supplementation.

Our data were derived from the specific national representativeness of the NHANES in the United States, imparting a degree of generalisability to the study results. At the same time, we included some obesity-related indicators to provide refined insights. Confounder adjustment using multivariable regression lent validity to the observed associations. Sensitivity analyses performed after multiple data imputation verified the robustness of outcomes. However, the cross-sectional nature of the study limits causal interpretations. Further longitudinal and mechanistic studies can build on these data, offering nuanced insights for the development of clinical and public health strategies against obesity and VD deficiency. It is crucial to acknowledge that the study focused exclusively on adults (aged 20–59 years), who participated in DXA testing and have data on body fat distribution. Our findings, viz. such as the observed increase in vitamin D concentration with age, may differ from those of studies that include broader or different populations. These differences underscore the potential variability in results due to the unique characteristics of our study cohort. The findings from the study warrant corroboration in diverse demographic groups. Besides, there are still other potential factors that affect VD that were not included in the study.

5. Conclusions

This nationally representative study offers unequivocal evidence affirming the association between fat and hypovitaminosis D among adults. Abdominal fat emerged as predictors of 25(OH)D, emphasising the importance of assessing fat distribution rather than simply replacing it with BMI or WHtR. Besides anthropometric indicators, demographic characteristics, lifestyle factors, and dietary habits also have complex effects on VD. Personalising clinical decisions and public health interventions based on these parameters could help mitigate the heavy burden of obesity and hypovitaminosis D.

Availability of Data and Materials

The data for this study were sourced from the Centers for Disease Control and Prevention's National Health and Nutrition Examination Survey, which can be obtained through the official website (<https://www.cdc.gov/nchs/nhanes/about/nhanes.htm>).

Author Contributions

YB contributed to editorial changes in the manuscript. YB read and approved the final manuscript. YB have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

The study protocol was reviewed and approved by the National Center for Health Statistics ethics review board. The protocol numbers are Continuation of Protocol #2011-17 (NHANES 2011–2017) and Protocol #2018-01 (NHANES 2017–2018). Detailed information can be found on the NHANES website. Written informed consent was obtained from all participants.

Acknowledgment

I would like to thank Editage (<https://www.editage.cn/>) for English language editing.

Funding

This research received no external funding.

Conflict of Interest

The author declares no conflict of interest.

References

- [1] Piché ME, Tchernof A, Després JP. Obesity Phenotypes, Diabetes, and Cardiovascular Diseases. *Circulation Research*. 2020; 126: 1477–1500. <https://doi.org/10.1161/CIRCRESAHA.120.316101>.
- [2] Gruber T, Pan C, Contreras RE, Wiedemann T, Morgan DA, Skowronski AA, *et al.* Obesity-associated hyperleptinemia alters the gliovascular interface of the hypothalamus to promote hypertension. *Cell Metabolism*. 2021; 33: 1155–1170.e10. <http://doi.org/10.1016/j.cmet.2021.04.007>.
- [3] Jiang Z, Wang Y, Zhao X, Cui H, Han M, Ren X, *et al.* Obesity and chronic kidney disease. *American Journal of Physiology. Endocrinology and Metabolism*. 2023; 324: E24–E41. <https://doi.org/10.1152/ajpendo.00179.2022>.
- [4] Rathmell JC. Obesity, Immunity, and Cancer. *The New England Journal of Medicine*. 2021; 384: 1160–1162. <https://doi.org/10.1056/NEJMcibr2035081>.
- [5] Mozaffarian D. Perspective: Obesity—an unexplained epidemic. *The American Journal of Clinical Nutrition*. 2022; 115: 1445–1450. <https://doi.org/10.1093/ajcn/nqac075>.
- [6] Ward ZJ, Bleich SN, Cradock AL, Barrett JL, Giles CM, Flax C, *et al.* Projected U.S. State-Level Prevalence of Adult Obesity and Severe Obesity. *The New England Journal of Medicine*. 2019; 381: 2440–2450. <https://doi.org/10.1056/NEJMsa1909301>.

- [7] Wortsman J, Matsuoka LY, Chen TC, Lu Z, Holick MF. Decreased bioavailability of vitamin D in obesity. *The American Journal of Clinical Nutrition*. 2000; 72: 690–693. <https://doi.org/10.1093/ajcn/72.3.690>.
- [8] Yuan C, Wang J, Zhang W, Yi H, Shu B, Li C, *et al.* Effects of obesity with reduced 25(OH)D levels on bone health in elderly Chinese people: a nationwide cross-sectional study. *Frontiers in Immunology*. 2023; 14: 1162175. <https://doi.org/10.3389/fimmu.2023.1162175>.
- [9] Rontoyanni VG, Avila JC, Kaul S, Wong R, Veeranki SP. Association between Obesity and Serum 25(OH)D Concentrations in Older Mexican Adults. *Nutrients*. 2017; 9: 97. <https://doi.org/10.3390/nu9020097>.
- [10] Karampela I, Sakelliou A, Vallianou N, Christodoulatos GS, Magkos F, Dalamaga M. Vitamin D and Obesity: Current Evidence and Controversies. *Current Obesity Reports*. 2021; 10: 162–180. <https://doi.org/10.1007/s13679-021-00433-1>.
- [11] Barrea L, Frias-Toral E, Pugliese G, Garcia-Velasquez E, DE Los Angeles Carignano M, Savastano S, *et al.* Vitamin D in obesity and obesity-related diseases: an overview. *Minerva Endocrinology*. 2021; 46: 177–192. <https://doi.org/10.23736/S2724-6507.20.03299-X>.
- [12] Semlitsch T, Stigler FL, Jeitler K, Horvath K, Siebenhofer A. Management of overweight and obesity in primary care-A systematic overview of international evidence-based guidelines. *Obesity Reviews: an Official Journal of the International Association for the Study of Obesity*. 2019; 20: 1218–1230. <https://doi.org/10.1111/obr.12889>.
- [13] Bray GA, Heisel WE, Afshin A, Jensen MD, Dietz WH, Long M, *et al.* The Science of Obesity Management: An Endocrine Society Scientific Statement. *Endocrine Reviews*. 2018; 39: 79–132. <https://doi.org/10.1210/er.2017-00253>.
- [14] Després JP, Lemieux I. Abdominal obesity and metabolic syndrome. *Nature*. 2006; 444: 881–887. <https://doi.org/10.1038/nature05488>.
- [15] Khan I, Chong M, Le A, Mohammadi-Shemirani P, Morton R, Brinza C, *et al.* Surrogate Adiposity Markers and Mortality. *JAMA Network Open*. 2023; 6: e2334836. <https://doi.org/10.1001/jamanetworkopen.2023.34836>.
- [16] Moltrier M, Pala L, Cosentino C, Mannucci E, Rotella CM, Cresci B. Body mass index (BMI), waist circumference (WC), waist-to-height ratio (WHR) e waist body mass index (wBMI): Which is better? *Endocrine*. 2022; 76: 578–583. <https://doi.org/10.1007/s12020-022-03030-x>.
- [17] Bennour I, Haroun N, Sicard F, Mounien L, Landrier JF. Recent insights into vitamin D, adipocyte, and adipose tissue biology. *Obesity Reviews: an Official Journal of the International Association for the Study of Obesity*. 2022; 23: e13453. <https://doi.org/10.1111/obr.13453>.
- [18] Heaney RP, Horst RL, Cullen DM, Armas LAG. Vitamin D3 distribution and status in the body. *Journal of the American College of Nutrition*. 2009; 28: 252–256. <https://doi.org/10.1080/07315724.2009.10719779>.
- [19] Best CM, Riley DV, Laha TJ, Pflaum H, Zelnick LR, Hsu S, *et al.* Vitamin D in human serum and adipose tissue after supplementation. *The American Journal of Clinical Nutrition*. 2021; 113: 83–91. <https://doi.org/10.1093/ajcn/nqaa295>.
- [20] Didriksen A, Burild A, Jakobsen J, Fuskevåg OM, Jorde R. Vitamin D3 increases in abdominal subcutaneous fat tissue after supplementation with vitamin D3. *European Journal of Endocrinology*. 2015; 172: 235–241. <https://doi.org/10.1530/EJ-14-0870>.
- [21] Gangloff A, Bergeron J, Lemieux I, Després JP. Changes in circulating vitamin D levels with loss of adipose tissue. *Current Opinion in Clinical Nutrition and Metabolic Care*. 2016; 19: 464–470. <https://doi.org/10.1097/MCO.0000000000000315>.
- [22] Gangloff A, Bergeron J, Pelletier-Beaumont E, Nazare JA, Smith J, Borel AL, *et al.* Effect of adipose tissue volume loss on circulating 25-hydroxyvitamin D levels: results from a 1-year lifestyle intervention in viscerally obese men. *International Journal of Obesity (2005)*. 2015; 39: 1638–1643. <https://doi.org/10.1038/ijo.2015.118>.
- [23] Buscemi S, Buscemi C, Corleo D, De Pergola G, Caldarella R, Meli F, *et al.* Obesity and Circulating Levels of Vitamin D before and after Weight Loss Induced by a Very Low-Calorie Ketogenic Diet. *Nutrients*. 2021; 13: 1829. <https://doi.org/10.3390/nu13061829>.
- [24] Obesity: preventing and managing the global epidemic. Report of a WHO consultation. *World Health Organization Technical Report Series*. 2000; 894: i–xii, 1–253.
- [25] DeLuca HF. The vitamin D system in the regulation of calcium and phosphorus metabolism. *Nutrition Reviews*. 1979; 37: 161–193. <https://doi.org/10.1111/j.1753-4887.1979.tb06660.x>.
- [26] Holick MF. Vitamin D deficiency. *The New England Journal of Medicine*. 2007; 357: 266–281. <https://doi.org/10.1056/NEJMr070553>.
- [27] Vimalraj S. Alkaline phosphatase: Structure, expression and its function in bone mineralization. *Gene*. 2020; 754: 144855. <https://doi.org/10.1016/j.gene.2020.144855>.
- [28] Bikle D, Christakos S. New aspects of vitamin D metabolism and action - addressing the skin as source and target. *Nature Reviews. Endocrinology*. 2020; 16: 234–252. <https://doi.org/10.1038/s41574-019-0312-5>.
- [29] Wasung ME, Chawla LS, Madero M. Biomarkers of renal function, which and when? *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 2015; 438: 350–357. <https://doi.org/10.1016/j.cca.2014.08.039>.
- [30] Khodabakhshi A, Mahmoudabadi M, Vahid F. The role of serum 25 (OH) vitamin D level in the correlation between lipid profile, body mass index (BMI), and blood pressure. *Clinical Nutrition ESPEN*. 2022; 48: 421–426. <https://doi.org/10.1016/j.clnesp.2022.01.007>.
- [31] Vimalaswaran KS, Berry DJ, Lu C, Tikkanen E, Pilz S, Hiraki LT, *et al.* Causal relationship between obesity and vitamin D status: bi-directional Mendelian randomization analysis of multiple cohorts. *PLoS Medicine*. 2013; 10: e1001383. <https://doi.org/10.1371/journal.pmed.1001383>.
- [32] Arunabh S, Pollack S, Yeh J, Aloia JF. Body fat content and 25-hydroxyvitamin D levels in healthy women. *The Journal of Clinical Endocrinology and Metabolism*. 2003; 88: 157–161. <https://doi.org/10.1210/jc.2002-020978>.
- [33] Kavarić S, Vuksanović M, Bozović D, Jovanović M, Jeremić V, Radojčić Z, *et al.* Body weight and waist circumference as predictors of vitamin D deficiency in patients with type 2 diabetes and cardiovascular disease. *Vojnosanitetski Pregled*. 2013; 70: 163–169. <https://doi.org/10.2298/vsp110713035k>.
- [34] Rosenstreich SJ, Rich C, Volwiler W. Deposition in and release of vitamin D3 from body fat: evidence for a storage site in the rat. *The Journal of Clinical Investigation*. 1971; 50: 679–687. <https://doi.org/10.1172/JCI106538>.
- [35] Ginnard OZB, Sisley S. Expression of vitamin D receptor pathway genes in subcutaneous adipose tissue of obese individuals. *Journal of the Endocrine Society*. 2021; 5: A658.
- [36] Szymczak-Pajor I, Miazek K, Selmi A, Balcerczyk A, Śliwińska A. The Action of Vitamin D in Adipose Tissue: Is There the Link between Vitamin D Deficiency and Adipose Tissue-Related Metabolic Disorders? *International Journal of Molecular Sciences*. 2022; 23: 956. <https://doi.org/10.3390/ijms23020956>.
- [37] Abbas MA. Physiological functions of Vitamin D in adipose tissue. *The Journal of Steroid Biochemistry and Molecular Biology*. 2017; 165: 369–381. <https://doi.org/10.1016/j.jsbmb.2016.08.004>.