

Review

Unveiling Methods to Stimulate Plant Resistance against Pathogens

Roohallah Saberi Riseh^{1,*}, Mozhgan Gholizadeh Vazvani¹

¹Department of Plant Protection, Faculty of Agriculture, Vali-e-Asr University of Rafsanjan, 7718897111 Rafsanjan, Iran

Academic Editor: Jen-Tsung Chen

Submitted: 27 January 2024 Revised: 5 April 2024 Accepted: 17 April 2024 Published: 14 May 2024

Abstract

Plant diseases caused by pathogens pose significant threats to agricultural productivity and food security worldwide. The traditional approach of relying on chemical pesticides for disease management has proven to be unsustainable, emphasizing the urgent need for sustainable and environmentally friendly alternatives. One promising strategy is to enhance plant resistance against pathogens through various methods. This review aims to unveil and explore effective methods for stimulating plant resistance, transforming vulnerable plants into vigilant defenders against pathogens. We discuss both conventional and innovative approaches, including genetic engineering, induced systemic resistance (ISR), priming, and the use of natural compounds. Furthermore, we analyze the underlying mechanisms involved in these methods, highlighting their potential advantages and limitations. Through an understanding of these methods, scientists and agronomists can develop novel strategies to combat plant diseases effectively while minimizing the environmental impact. Ultimately, this research offers valuable insights into harnessing the plant's innate defense mechanisms and paves the way for sustainable disease management practices in agriculture.

Keywords: stimulating plant resistance; vigilant defenders; priming; ISR; genetic engineering; sustainable disease management

1. Introduction

Plants play a crucial role in sustaining life on Earth, providing us with food, fiber, fuel, and numerous other resources [1]. However, their productivity and survival are constantly challenged by various pathogens, including bacteria, fungi, viruses, and pests [2–4]. These pathogens can cause devastating diseases that lead to significant crop losses and threaten global food security [5-7]. In response to these challenges, plants have developed intricate defense mechanisms to protect themselves against pathogens. The concept of plant resistance refers to the ability of plants to recognize and respond to pathogen attacks [8-11]. Understanding and harnessing plant resistance is of paramount importance for sustainable agriculture and crop protection. Traditional approaches to disease management often rely on the use of chemical pesticides, which can have detrimental effects on the environment and human health. Moreover, pathogens can develop resistance to these chemicals over time, rendering them less effective [12-16]. Transitioning from a model of vulnerability to one of vigilance, where plants are empowered to defend themselves against pathogens, offers a more sustainable and environmentally friendly approach to disease management. By boosting plant resistance, we can reduce reliance on synthetic pesticides, minimize the environmental impact, and promote healthier ecosystems [17–19].

Various methods and techniques have been developed to stimulate and enhance plant resistance against pathogens. These methods can be broadly categorized into two main approaches: conventional breeding and biotechnological

interventions. (1) Conventional breeding approach involves selectively breeding plants with desirable resistance traits through classical breeding techniques. It relies on natural genetic variations within plant populations and aims to develop new cultivars with improved resistance. Conventional breeding methods often take advantage of techniques such as marker-assisted selection and quantitative trait loci (QTL) mapping to identify and incorporate resistance genes into breeding programs [20-24]. (2) Biotechnology offers powerful tools to engineer or augment plant resistance traits. Genetic engineering techniques, such as transgenic approaches, enable the introduction of specific genes into plants to confer resistance against pathogens [25–27]. For example, plants can be engineered to produce antimicrobial peptides or proteins that directly inhibit pathogen growth [28,29]. Additionally, genome editing techniques, such as clustered regularly interspaced short palindromic repeats-CRISPR-associated (CRISPR-Cas9), allow precise modifications of the plant's own genetic material to enhance resistance [30-32].

Other biotechnological strategies include the use of beneficial microbes, such as rhizobacteria and mycorrhiza fungi, which can establish symbiotic relationships with plants and stimulate their immune responses [33–35]. Moreover, elicitors derived from pathogen-associated molecular patterns (PAMPs) or plant-derived compounds can be applied to induce systemic acquired resistance (SAR) or induced systemic resistance (ISR), respectively [36–38]. Also viral cross protection in plants is known as an acquired immunity phenomenon, that a mild virus isolate/strain can protect plants against economic damage

^{*}Correspondence: r.saberi@vru.ac.ir (Roohallah Saberi Riseh)

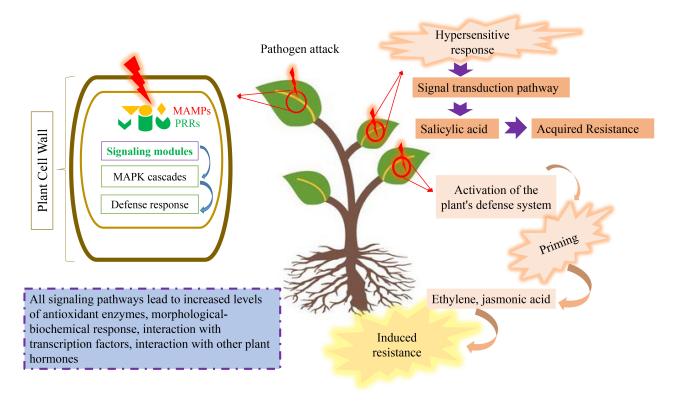


Fig. 1. The different stages of the signaling pathway. MAMPs, Microbe-Associated Molecular Patterns; PRR, Pattern Recognition Receptor; MAPK, Mitogen-Activated Protein Kinase.

caused by a severe challenge strain or isolate of the same virus [39–41]. These methods to stimulate plant resistance are not mutually exclusive and can be combined to achieve synergistic effects in enhancing plant defenses against pathogens [42–44].

In summary, understanding and harnessing plant resistance is crucial for sustainable agriculture. By transitioning from vulnerability to vigilance and utilizing various breeding and biotechnological methods, we can empower plants to defend themselves against pathogens, reduce reliance on chemical pesticides, and promote more sustainable crop protection practices.

2. Understanding Plant-Pathogen Interactions

Plant-pathogen interactions form a complex and dynamic field of study within the realm of plant pathology [45–47]. Plants have evolved intricate defense mechanisms to protect themselves against invading pathogens, while pathogens have developed various strategies to overcome these defenses and establish successful infections [48,49]. Understanding the intricacies of these interactions is crucial for developing effective strategies to manage plant diseases and ensure global food security. This section will delve into the key components of plant-pathogen interactions, including the recognition process, signaling pathways, and defense mechanisms employed by both the plant and the pathogen.

2.1 Recognition Process

This process consists of two main parts: (1) Pathogen-associated Molecular Patterns (PAMPs) and Pattern Recognition Receptors (PRRs); (2) Effector-triggered Immunity (ETI). The recognition process in plant-pathogen interactions begins with the detection of conserved microbial molecules known as pathogen-associated molecular patterns (PAMPs) by the plant's pattern recognition receptors (PRRs) [50–52]. PAMPs can be structural components of pathogens such as chitin or flagellin, which are recognized by specific PRRs present on the surface of plant cells [53,54]. The binding of PAMPs to PRRs triggers a series of events leading to the activation of defense responses [55,56].

Some pathogens, known as biotrophic pathogens, secrete effector molecules into the plant cells to suppress host defenses and establish successful infections [57–59]. However, plants have evolved Resistance (R) proteins that recognize specific effector molecules and trigger a strong immune response called Effector-triggered Immunity (ETI) [60,61]. ETI often leads to localized hypersensitive responses, where the infected plant cells undergo programmed cell death to limit the pathogen's spread [62–64].

2.2 Signaling Pathways

Fig. 1 shows the different stages of the signaling pathway. Signaling pathway include three steps, that we explain these steps: (1) Mitogen-Activated Protein Kinase



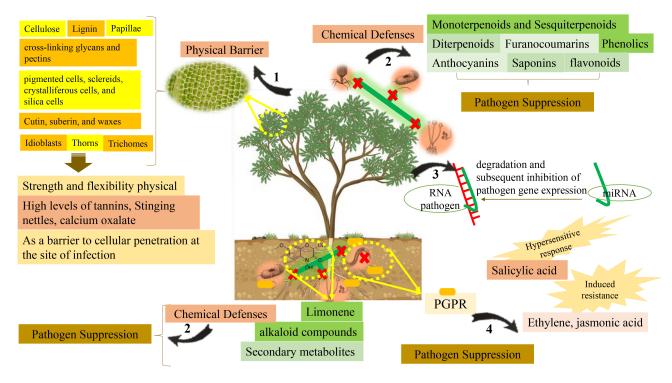


Fig. 2. Plant defense mechanisms against pathogens. PGPR, plant growth-promoting rhizobacteria; miRNA, microRNA.

(MAPK) Pathway: Upon PAMP perception or ETI activation, plants activate signaling pathways to transmit the defense signals throughout the plant [65–67]. One of the central signaling pathways involved in plant immunity is the Mitogen-Activated Protein Kinase (MAPK) pathway [68,69]. MAPK cascades are activated upon PRR or R protein activation and regulate various defense responses, including the production of antimicrobial compounds, reinforcement of cell walls, and induction of defense-related genes [70–72].

(2) Salicylic Acid (SA)-Mediated Signaling Pathway: The salicylic acid (SA)-mediated signaling pathway is a crucial component of plant defense against biotrophic pathogens [73–75]. SA acts as a signaling molecule and regulates several defense-related processes, including the expression of pathogenesis-related (PR) genes, accumulation of reactive oxygen species (ROS), and activation of SAR [76,77]. The SA pathway often functions in parallel with other signaling pathways to provide robust and effective defense against pathogens [78,79].

(3) Jasmonic Acid (JA)-Mediated Signaling Pathway: The jasmonic acid (JA)-mediated signaling pathway plays a significant role in defense against necrotrophic pathogens and herbivorous insects [79–81]. JA and its derivatives, known as Jasmonate, regulate the expression of numerous defense-related genes, leading to the production of toxic metabolites, proteinase inhibitors, and other defensive compounds [82,83]. The JA pathway also interacts with other signaling pathways, such as the SA pathway, to fine-tune plant defense responses [84,85].

2.3 Defense Mechanisms

Plant defense mechanisms against pathogens are extremely complex. Different ways of protecting themselves against external factors are used by plants. In the event that a defense barrier against the pathogen attack is not effective, the plant uses a different method to suppress the pathogen attack. Plant defense mechanisms against pathogens are summarized in Fig. 2. A further description of each of these mechanisms will be provided.

2.3.1 Physical Barriers

Plants possess physical barriers that act as the first line of defense against pathogens. The plant cell wall provides a physical barrier that can prevent pathogen entry. Additionally, plants produce specialized structures like trichomes, thorns, and waxes that can impede pathogen movement and colonization. Furthermore, stomatal closure, induced by pathogen detection, restricts pathogen entry through leaf pores [86–94].

2.3.2 Chemical Defenses

The biosynthesis of secondary metabolites from plants is related to induced chemical defense generated for protection against different phytopathogenic attack [95]. Plants produce an array of chemical compounds that can inhibit pathogen growth, deter herbivores, and attract beneficial organisms. These chemical defenses include antimicrobial compounds (phytoalexins), toxic secondary metabolites (alkaloids, terpenoids), and volatile organic compounds (VOCs) that can act as signaling molecules to attract



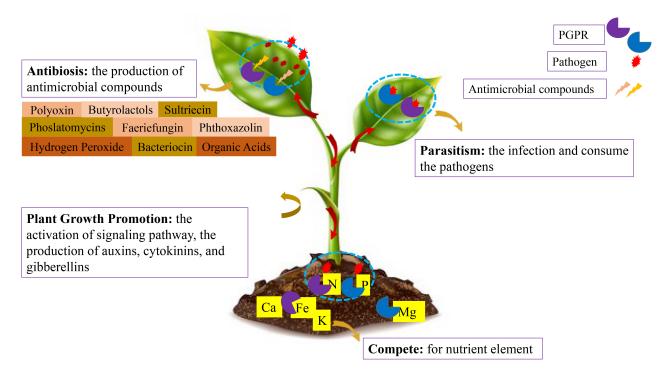


Fig. 3. The role of beneficial microorganisms in plant defense.

natural enemies of the pathogen [96–102]. For example in Calceolaria spp. (a notorious weed and as a popular ornamental garden plant) is synthesized a number of iridoids, flavonoids, naphthoguinones and phenylpropanoids that have been shown to possess biological activities. These metabolites have activities insecticides, fungicides, and Bactericidal [95]. In the Gramineae, the cyclic hydroxamic 2,4-dihydroxy-1,4-benzoxazin-3-one 2,4-dihydroxy-7-methoxy-1,4-benzoxazin-3-one and (DIMBOA) form part of the defense against insects and microbial pathogens [103]. Benzoxazinoids are a class of indole-derived plant chemical defenses comprising compounds. These phytochemicals are widespread in grasses, including important cereal crops such as maize, wheat, and rye, as well as a few dicot species, and display a wide range of antifeedant, insecticidal, antimicrobial, and allelopathic activities [104]. Also DIMBOA in Maize seedlings is directly associated with multiple insect-resistance against insect pests such as Asian corn borer and corn leaf aphids [105].

2.3.3 RNA Silencing

RNA silencing is an important defense mechanism employed by plants to regulate gene expression and combat invading pathogens. Small RNA molecules, known as small interfering RNAs (siRNAs) or microRNAs (miRNAs), can target pathogen-derived nucleic acids, leading to their degradation and subsequent inhibition of pathogen gene expression. RNA silencing also plays a role in regulating plant developmental processes and stress responses [106–117].

2.3.4 Induced Systemic Resistance (ISR)

ISR is a defense mechanism in which local infection by certain pathogens triggers enhanced resistance throughout the entire plant. ISR is mediated by beneficial microbes present in the rhizosphere, such as plant growth-promoting rhizobacteria (PGPR). These microbes induce systemic defense responses in the plant, priming it to mount stronger defenses against future infections [38,118–124]. The use of beneficial microorganisms as biocontrol agents has gained significant attention in recent years due to their potential to reduce chemical pesticide dependence, mitigate environmental risks associated with synthetic pesticides, and contribute to sustainable agricultural practices. These biocontrol agents can act directly or indirectly in plant defense by antagonizing pathogenic microbes through competition, antibiosis, parasitism, induced resistance, or by enhancing the plant's own defense mechanisms [125,126]. Various groups of microorganisms have been identified as inducers of systemic resistance in plants. (1) PGPR: PGPR are a diverse group of bacteria that colonize the rhizosphere and promote plant growth through various mechanisms. Many PGPR strains are capable of inducing systemic resistance in plants. Examples include strains of Pseudomonas, Bacillus, and Serratia, which have been shown to effectively protect crops against diseases caused by fungal, bacterial, and viral pathogens [127-130]. (2) Mycorrhizal Fungi: Mycorrhizal fungi form mutualistic associations with plant roots, facilitating nutrient uptake and providing other benefits to the host plant. Certain species of mycorrhizal fungi, such as Trichoderma and Glomus, have been found to induce systemic resistance in plants. These fungi can colonize the



plant roots and trigger a range of defense responses that confer protection against pathogens [131–133]. Table 1 (Ref. [134–156]) shows the types of defense mechanisms in plants against pathogens and defense pathways in plantpathogen interaction.

- 2.3.4.1 Role of Beneficial Microorganisms in Plant Defense. Beneficial microorganisms lead to the induction of defense reactions and ISR through different processes (Fig. 3). These processes include:
- (1) Competition: Certain beneficial microorganisms compete with plant pathogens for resources such as space, nutrients, and moisture, thereby limiting the growth and establishment of pathogenic populations. This competitive exclusion can effectively suppress pathogen colonization and subsequent disease development [157–160].
- (2) Antibiosis: Some biocontrol agents produce antimicrobial compounds, such as antibiotics or antifungal metabolites that inhibit the growth and activity of plant pathogens. These compounds can directly kill or inhibit the pathogens, preventing them from causing diseases in plants [161–164].
- (3) Parasitism: Certain beneficial microorganisms are parasites of plant pathogens. They can infect and consume the pathogens, leading to their decline and control [165]. Examples include mycoparasitic fungi that attack other fungi [166,167], bacteriophages that infect and destroy bacteria [168–170], and nematodes that feed on plant-parasitic nematodes [171–173].
- (4) Plant Growth Promotion: In addition to their role in plant defense, beneficial microorganisms can promote plant growth and development. They can produce plant growth-promoting substances like auxins, cytokinins, and gibberellins, which enhance root and shoot growth, increase nutrient availability, and improve stress tolerance in plants.
- 2.3.4.2 Application Methods and Considerations for Using Biocontrol Agents. The successful implementation of biocontrol agents relies on appropriate application methods and considerations to ensure their efficacy and sustainability. The following aspects need to be taken into account when using biocontrol agents:
- (1) Selection of Effective Strains: The first step in utilizing biocontrol agents is selecting appropriate strains with demonstrated efficacy against specific plant pathogens. These strains should have a proven track record of disease suppression, compatibility with target crops, and environmental adaptability [174–176]. In choosing a biocontrol strain, it should be noted that not all biocontrol agents have the same biocontrol activity and the best type of strain should be identified with laboratory screenings [176,177]. Some strains may induce plant resistance under field conditions, providing effective suppression of disease and that there may be some benefit to the integration of rhizosphereapplied strains and foliar-applied biological control agents

- [178]. The effectiveness of plant growth—promoting bacteria is variable under different biotic and abiotic conditions. Abiotic factors may negatively affect the beneficial properties and efficiency of the introduced PGPR inoculants [179]. Also has been reported the combination of microorganisms gives better results probably due to the different mechanisms used [180,181].
- (2) Formulation Development: Biocontrol agents are typically formulated into products that facilitate their storage, handling, and application. Formulations can include granules, powders, liquids, or carrier-based systems. The choice of formulation depends on factors such as target pathogens, crop type, application method, and economic feasibility [182–196]. The application of delivery system based on encapsulation technology shows a promising technique to store and deliver biocontrol agents [197,198]. Generally encapsulation improves the viability, stability, and controlled release of bacteria [199]. The encapsulation process consists of incorporation of an active core into a matrix structure through mechanical action, followed by stabilization by chemical or physicochemical means [200].

Of course, in encapsulation, attention should also be paid to the coating in which the biocontrol agents are enclosed. Today, much attention is paid to polymer and protein coatings such as chitosan, alginate, and whey, which have biocompatibility properties [192,194,195,199, 201–207]. For example in a research in order to biological control of Damping-off disease due to Pythium aphanidermatum was synthesis a suitable formulation for a Streptomyces fulvissimus Uts22 strain based on alginate-Arabic gum and nanoparticles (SiO₂ and TiO₂). The S. fulvissimus suspension was 1010 CFU/mL, that maximum release of bacteria from the capsule was recorded on the 35th day of storage, about 109 CFU/gr. Encapsulated bacteria resulted in a 95% reduction in damping-off disease of cucumber [193]. In other research the survivability and controlled release of Pseudomonas fluorescens encapsulated with alginate (Alg) combined with whey protein concentrate (WPC), carboxymethyl cellulose (CMC), and peanut butter (PB) were evaluated. The number of cells in all mixtures was 2×10^{11} CFU/mL. The Alg-WPC microcapsules provided a better controlled release over two months and they were able to suppress the pathogen up to 90% [208]. In most of the studies, the bacterial population in microcapsule is reported to be 10^{10} – 10^{11} CFU/mL which was able to control various plant diseases [209–211].

(3) Application Timing: Timely application of biocontrol agents is crucial for effective disease management [212,213]. It is essential to identify the critical periods during which pathogen establishment and infection are most likely, and apply biocontrol agents accordingly. Early preventive application is often employed to establish a protective layer around the plant roots or foliage before pathogens become established.

Table 1. Types of defense mechanisms in plants against pathogens and signaling pathways related.

Defense mechanisms	Pathogen	Host	Signaling pathway	Reference
Physical Barriers	Fusarium oxysporum f. sp. pisi	Pea	cell wall strengthening, formation of papilla-like structures at penetration sites and ac-	[134]
			cumulation of different substances within and between cells	
Physical Barriers	Botrytis bunch rot	Grapes	Increase in cuticle thickness	[135]
Physical Barriers	Alternaria alternata	Tomato	enhancement of alkaloids and terpenoids along with increase in cell wall hemicellulose content (cellulose, lignin and pectin)	[136]
Physical Barriers	Pseudomonas solanacearum	Tomato	In resistant cultivars, tyloses occluded the colonized vessels and the contiguous ones, limiting bacterial spread	[137]
Chemical Defenses	Rhizoctonia solani	rice	alkaloid and terpenoid compounds in the leaf extracts of Zizyphus jujube: increase in chitinase, β -1,3-glucanase and peroxidase	[138]
Chemical Defenses	Puccinia graminis f. sp. tritici	Wheat	high expression of metabolites from thiamine and glyoxylate, such as flavonoid gly- cosides: defense response strategy by wheat varieties and biochemical changes in the expression of wheat metabolites	[139]
Chemical Defenses	Blumeria graminis	Wheat	tryptophan and pyrimidine metabolism, soquinoline alkaloid biosynthesis, phenolic acids, flavonoid carboglycosides, indole alkaloids, phenolic amines, amino acids	[140]
Chemical Defenses	Fusarium crown rot	Wheat	higher content of benzoxazolin-2-one (BOA) and 6-methoxy-benzoxazolin-2-one (MBOA): resistance to Fusarium crown rot	[141]
Chemical Defenses	streak mosaic virus	Wheat	increase in amino acid metabolism, lipid metabolism and alkaloids pathways	[142]
Chemical Defenses	late blight	Potato	increase in benzylisoquinoline alkaloids (BIAs) in response to pathogen infection and cell wall reinforcement	[143]
Chemical Defenses	Alternaria solani	Tomato	Activation metabolic (steroidal glycol-alkaloid, lignin, and flavonoid biosynthetic pathways)	[144]
RNA Silencing	Verticillium genus	Arabidopsis	RNA silencing: role in defence against bacterial plant pathogens through modulating host defence responses	[145]
RNA Silencing	Fusarium graminearum	Barley	targeting key components of the fungal RNAi machinery via spray-induced gene silencing: protect barley leaves from Fusarium infection	[146]
RNA Silencing	Hyaloperonospora parasitica	Arabidopsis	The RPP5 (for recognition of Peronospora parasitica 5) locus is poised to respond to pathogens that disturb RNA silencing	[147]
RNA Silencing	barley stripe mosaic virus	Wheat	Virus-induced gene silencing: a powerful tool for dissecting the genetic pathways of disease resistance in hexaploid wheat	[148]
RNA Silencing	Puccinia striiformis f. sp. tritici	Wheat	RNA interference: efficiently and durably control, hyphal development strongly restricted; necrosis of plant cells as resistance responses	[149]
Induced Systemic Resistance by different strains bacterial (from the rhizosphere, endosphere, and phyllosphere)	Fusarium root and crown rot	Wheat	Pseudomonas mediterranea: by inducing antioxidant enzyme activity and resistance to disease	[150]
Induced Systemic Resistance by Pseudomonas fluorescens	Take-all disease	Wheat	Increased the activities of β -1-3-glucanase, β -1,4-glucanase and phenolic compounds in the wheat	[151]
Induced Systemic Resistance by Tricho- derma pubescens	Rhizoctonia solani	Tomato	Increase in antioxidant enzyme production	[152]
Induced Systemic Resistance by Trichoderma hamatum	tobacco mosaic virus	Tomato	Increases in the transcriptional levels of polyphenol genes and pathogenesis-related proteins	[153]
Induced Systemic Resistance by Trichoderma	powdery mildew	Grapevines	Increase in chitinase and β -1, 3-glucanase peroxidase and polyphenol oxidase activities	[154]
Induced Systemic Resistance by Bacillus	Sarocladium oryzae	Rice	Increase in defense enzymes and reactive oxygen species	[155]
Induced Systemic Resistance by arbuscular mycorrhizal fungus Glomus mosseae	Phytophthora parasitic	Tomato	Increase in phenolics and plant cell defense responses	[156]

- (4) Application Methods: Several application methods can be used to introduce biocontrol agents into the plant system. These methods include seed treatment, soil drenching, foliar spraying, root dip, and soil incorporation. The choice of application method depends on factors such as the target pathogen's location, stage of crop growth, formulation type, and available equipment [214,215]. The delivery of a high number of viable cells to the plant is a prerequisite to reach a satisfactory colonization rate, which in turn enhances the desired effect in the field. A product has to display a sufficiently long shelf life, which describes the stability throughout the production process, packaging, storage and transport conditions. During subsequent application on the field, the inoculant is confronted with additional factors that are detrimental to its viability such as UV radiation, temperature and pH [216-218]. A suitable formulations is nesasory to enhance biocontrol agents efficiency at the target site and to facilitate the practical use by farmers [218-222].
- (5) Environmental Factors: Environmental conditions can influence the success of biocontrol agent applications. Factors such as temperature, humidity, pH, and soil moisture should be considered to ensure optimal survival, establishment, and activity of the biocontrol agents. Compatibility with existing agricultural practices and pesticides should also be assessed to avoid potential interactions or negative effects [223,224]. Selection of strains with multiple capabilities in different biological conditions can help in the sustainable management of diseases. For example in research a number of bacterial strains (Bacillus, Pseudomonas, Stenotrophomonas and Peribacillus) were examined under waterlogged conditions. Waterlogging shown to induce shifts in the bacterial strains potential, as well as on the structure of a population exhibiting a particular trait. The abundances of populations promoting nutrient availability to plants were found to increase after waterlogging, biocontrol protease activity proved to be the trait most sensitive to waterlogging. Enterobacter and Pantoea species were identified as main players in biofilm formation under waterlogging conditions. Waterlogging also appears to trigger the emergence of extremophilic and acidophilic bacteria, aspect important for potential adaptation of soil bacteria to future climate projections, with Bacillus and Pseudomonas showed best potential [225]. The efficiency of Bacillus biocontrol agents, however, is negatively influenced by ultraviolet (UV) radiation. But has been reported that preventive and repair molecular mechanisms of DNA caused by UV radiation are in B. subtilis. B. subtilis spores use recombination repair (in repairing damage resulting from UV-A), nucleotide excision repair (to repair UV-B and UV-C), and spore photoproduct lyase (to repairing UV-B and UV-C) [226]. Biological control agents have different growth in different temperature conditions [227-232]. Also, PGPR can compete with other bacteria by colonizing rapidly and accumulating a greater supply of
- nutrients, preventing other organisms from growing. PGPR has different strategies to colonize, of which each is related to a particular host [233–235]. It is necessary to mention researchers have explored sustainable tools for plant protection against pathogens, including the use of biological control agents (BCAs), which have the potential to complement or replace chemical fungicides. The complex interactions among a target pathogen, a host plant, and the BCA population in a changing environment can be studied by processbased, weather-driven mathematical models, able to interpret the combined effects on BCA efficacy of: (1) BCA mechanism of action, (2) timing of BCA application with respect to timing of pathogen infection (preventative vs. curative), (3) temperature and moisture requirements for both pathogen and BCA growth, and (4) BCA survival capability [236]. The environmental responses of BCAs should be considered during their selection, BCA survival capability should be considered during both selection and formulation, and weather conditions and forecasts should be considered at the time of BCA application in the field [236].
- (6) Integrated Disease Management (IDM): The integration of biocontrol agents into an IDM program is essential for sustainable disease management [237,238]. Biocontrol agents can be combined with other cultural, physical, or chemical control measures in a complementary manner to maximize their effectiveness while minimizing reliance on synthetic pesticides [239–241].
- (7) Monitoring and Evaluation: Regular monitoring and evaluation of biocontrol agent efficacy are crucial for assessing their performance and making necessary adjustments. This may involve monitoring disease incidence, severity, crop health, and yield parameters. By monitoring the effectiveness of biocontrol agents, growers can make informed decisions regarding their continued use or the need for alternative control strategies [242,243].
- (8) Host genetic: The success of microbial biocontrol depends on the outcome of complex interactions among the plant host, beneficial microflora, pathogens, and environment [244,245]. The host effect on biocontrol is mediated by different mechanisms. The discovery of a genetic basis in the host for interactions with a biocontrol agent suggests new opportunities to exploit natural genetic variation in host species to enhance understanding of beneficial plantmicrobe interactions and develop ecologically sound strategies for disease control in agriculture [246,247]. Various reports show that significant potential there is to exploit genetic variation in the host through breeding to enhance beneficial interactions with microorganisms. Plant breeding uses tremendous genetic variation in plant germplasm to increase crop productivity and enhance the plant's ability to endure pathogens, pests, and abiotic stresses [246,248,249]. For example research to assess the role of the plant host in disease suppression, used a genetic mapping population of tomato to evaluate the efficacy of the biocontrol agent Bacillus cereus against the seed pathogen Pythium torulo-

sum. This research detected significant phenotypic variation among recombinant inbred lines (RIL) that comprise the mapping population for resistance to *P. torulosum*, disease suppression by *B. cereus*, and growth of *B. cereus* on the seed. Genetic analysis revealed that three QTL associated with disease suppression by B. cereus explained 38% of the phenotypic variation among the recombinant inbred lines. In two cases, QTL for disease suppression by B. cereus map to the same locations as QTL for other traits, suggesting that the host effect on biocontrol is mediated by different mechanisms [246].

- 2.3.4.3 Application of ISR in Agriculture. The concept of induced systemic resistance has significant implications for sustainable agriculture and disease management. Harnessing the potential of beneficial microorganisms to enhance plant defense offers an environmentally friendly alternative to synthetic pesticides and fungicides. Some key applications of ISR in agriculture include:
- (1) Biological control of plant diseases: Beneficial microorganisms that induce systemic resistance can be used as biological control agents to protect crops against pathogens. By applying these microorganisms to the soil or seed treatments, farmers can reduce reliance on chemical pesticides and promote sustainable disease management [250–252].
- (2) Enhanced crop productivity: Induced systemic resistance not only improves disease resistance but also has positive effects on plant growth and productivity. Plants treated with beneficial microorganisms often exhibit increased vigor, nutrient uptake efficiency, and tolerance to abiotic stresses, leading to higher yields and improved crop quality [253,254].
- (3) Reduction of chemical inputs: Utilizing induced systemic resistance can help reduce the dependence on synthetic chemicals in agriculture. By promoting natural defense mechanisms in plants, the need for chemical fungicides and pesticides can be minimized, thus reducing environmental pollution and potential health risks [255,256].
- (4) Crop rotation and intercropping strategies: Incorporating crops with different induced systemic resistance profiles into rotation systems or intercropping arrangements can help manage diseases more effectively. This approach disrupts the buildup of specific pathogens in the soil and promotes a diverse microbial community that enhances overall disease suppression [257–259].
- 2.3.4.4 Mechanisms of ISR. The mechanisms by which beneficial microorganisms induce systemic resistance in plants are complex and multifaceted. Several key processes have been identified:
- (1) Priming: Beneficial microorganisms can prime the plant's immune system, preparing it to respond more rapidly and effectively to pathogen attacks. This priming involves the activation of various signaling pathways and the up regulation of defense-related genes [260–262].

- (2) Production of antimicrobial compounds: Some beneficial microorganisms produce antimicrobial compounds, such as antibiotics and antifungal metabolites that directly inhibit the growth and development of plant pathogens. These compounds can help protect the plant from infection and reduce disease severity [263–265].
- (3) Induction of systemic acquired resistance (SAR): SAR is a long-lasting immune response that occurs throughout the entire plant following localized pathogen attack. Beneficial microorganisms can induce SAR in plants, leading to the production of antimicrobial proteins, phytoalexins, and other defense molecules that confer resistance against a wide range of pathogens [36,266].
- (4) Hormonal signaling: Beneficial microorganisms can modulate the plant's hormonal balance, particularly the levels of salicylic acid (SA), jasmonic acid (JA), and ethylene. These hormones play critical roles in regulating plant defense responses. By manipulating hormone levels, beneficial microorganisms can enhance the plant's ability to defend against pathogens [267–269].
- (5) Systemic signaling: Beneficial microorganisms can also elicit systemic signaling within the plant, where chemical signals are transmitted from the site of microbial colonization to distant parts of the plant. This systemic signaling helps activate defense responses in uninfected tissues, providing protection against future pathogen attacks [266].

In conclusion, understanding plant-pathogen interactions is essential for developing effective disease management strategies and ensuring global food security. The recognition process, signaling pathways, and defense mechanisms discussed in this section highlight the intricate mechanisms involved in plant immunity and pathogen virulence. Further research in this field is crucial to unravel the complexities of plant-pathogen interactions and discover new targets for disease control. One area of research that holds immense potential is the identification and characterization of effector proteins. Effector proteins are produced by pathogens and play a crucial role in suppressing plant defenses and promoting infection [270,271]. Understanding the function and mode of action of these effectors can provide valuable insights into how pathogens manipulate the plant's immune system. By targeting specific effectors, scientists can develop novel strategies to disrupt pathogen virulence and enhance plant resistance. Advancements in molecular biology and genomics have revolutionized the study of plant-pathogen interactions. Techniques such as next-generation sequencing and high-throughput functional genomics enable comprehensive analysis of pathogen genomes and the identification of key genes involved in virulence and host adaptation [272–279]. This information can be utilized to develop diagnostic tools for early detection of pathogens and design crop breeding programs for enhanced disease resistance. Furthermore, exploring the role of beneficial microbes in plant immunity has gained



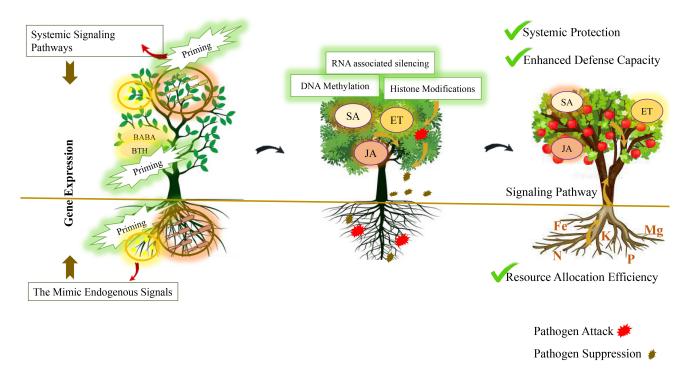


Fig. 4. Priming mechanisms, sources of priming and its benefits. BABA, β -aminobutyric acid; BTH, benzothiadiazole; SA, salicylic acid; ET, ethylene; JA, Jasmonic acid.

significant attention in recent years. Certain microorganisms can form symbiotic relationships with plants, providing protection against pathogens through mechanisms such as induced systemic resistance. Harnessing the potential of these beneficial microbes could offer sustainable and ecofriendly alternatives to conventional chemical-based disease management strategies [38,280-283]. Collaboration between researchers, plant breeders, farmers, and policymakers is vital to translate scientific discoveries into practical solutions. By bridging the gap between academia and agriculture, innovative disease management strategies can be implemented on a larger scale, benefiting both smallscale farmers and large-scale agricultural systems. In conclusion, ongoing research into plant-pathogen interactions is instrumental in developing effective disease management strategies, reducing crop losses, and ensuring global food security. Continued investment in this field will lead to the discovery of novel approaches for disease control, ultimately contributing to sustainable and resilient agricultural systems.

3. Priming Plant Defense Responses

Priming plant defense responses is a strategy employed by plants to enhance their ability to respond rapidly and effectively to potential threats such as pathogens, pests, and environmental stresses. This process involves the activation of various defense mechanisms, which are prepped and ready to be deployed when a threat is detected. Primed plants exhibit an enhanced defense response upon subsequent exposure to stressors, allowing them to mount a

quicker and more efficient defense compared to non-primed plants [284–287]. Priming mechanisms, sources of priming and its benefits are shown in Fig. 4. We will continue to explain these things.

3.1 Inducers of Priming

The factors that induce priming can be mentioned as follows:

- (1) Beneficial Microbes: Certain beneficial microbes, such as rhizobacteria and mycorrhiza fungi, have been shown to prime plant defense responses. These microbes establish symbiotic relationships with plants and activate systemic signaling pathways that enhance plant defenses. They can induce hormonal changes, trigger defense gene expression, and enhance nutrient uptake, all of which contribute to priming [34,43,288–293].
- (2) Plant Extracts and Elicitors: Application of specific plant extracts or elicitors derived from microbial pathogens can also prime plant defense responses. These extracts contain bioactive compounds that mimic signals generated during pathogen invasion, triggering a defense response in the absence of actual threat. This pre-emptive activation prepares the plant for a rapid and effective response when encountering real pathogens or pests [294–296].
- (3) Priming Agents: Synthetic compounds known as priming agents can be used to induce priming in plants. These compounds are designed to mimic endogenous signals or modulate hormone levels, thereby enhancing plant defense responses [297]. Examples of priming agents in-



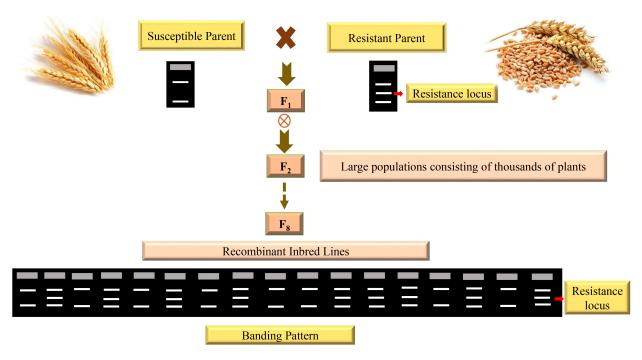


Fig. 5. Marker assisted selection.

clude chemical inducers like β -aminobutyric acid (BABA) [298–300], pipecolic acid [301,302], and benzothiadiazole (BTH) [286,303].

3.2 Mechanisms of Priming

The mechanisms of priming are include: (1) Hormonal Signaling: Plant hormones play a crucial role in priming plant defense responses. Jasmonic acid (JA), salicylic acid (SA), ethylene (ET), and abscisic acid (ABA) are among the key signaling molecules involved. These hormones regulate gene expression and metabolic processes that contribute to defense responses. Priming promotes the accumulation of these hormones or sensitizes the plant to their action, enabling a faster and stronger defense response upon attack [304–315]. (2) Reactive Oxygen Species (ROS): Priming can induce the production of ROS, including hydrogen peroxide (H₂O₂) and superoxide radicals (O^{2-}) . Controlled levels of ROS act as signaling molecules that activate defense-related genes and strengthen cell walls. ROS also participate in triggering SAR, which confers broad-spectrum resistance throughout the plant [316–322]. (3) Defense Gene Activation: Priming leads to the up regulation of defense-related genes encoding antimicrobial proteins, pathogenesis-related (PR) proteins, and enzymes involved in secondary metabolite synthesis. These genes are transcriptionally activated and ready to be expressed upon encountering a stressor. The increased expression of defense genes primes the plant for a rapid and robust defense response [263,292,323-326]. (4) Epigenetic Modifications: Priming can induce epigenetic modifications, such as DNA methylation and histone modifications, which alter the accessibility of genes involved in defense responses. These modifications can result in the heritable enhancement of defense-related gene expression, enabling a more efficient response to subsequent stressors [327–330].

3.3 Benefits of Priming

Priming has many advantages, which can be mentioned as follows: (1) Enhanced Defense Capacity: Primed plants possess an increased capacity to defend against various stresses, including biotic and abiotic challenges. The priming process allows them to rapidly activate defense mechanisms, resulting in faster pathogen recognition and containment, reduced damage, and improved survival rates [331,332]. (2) Resource Allocation Efficiency: Priming enables plants to allocate resources more efficiently by priming only when necessary without constitutive activation of defense responses. This strategy minimizes energy expenditure under normal conditions while ensuring swift and effective responses in times of stress. (3) Systemic Protection: Priming can induce SAR or ISR, where the entire plant becomes more resistant to subsequent attacks by pathogens or pests. This systemic protection extends beyond the initially primed tissue, providing a broad-spectrum defense response throughout the plant [333–335].

In conclusion, priming plant defense responses is a sophisticated and dynamic process that prepares plants for potential threats. By priming various defense mechanisms, plants can mount faster and stronger responses when faced with pathogens, pests, or environmental stresses, ultimately improving their chances of survival and reducing damage. Understanding and harnessing the mechanisms of priming could have significant implications for sustainable agriculture and crop protection strategies in the future.



4. Genetic Approaches to Enhance Plant Resistance

4.1 Breeding for Resistance Traits

Breeding for resistance traits has long been a traditional approach to enhance plant resistance against pathogens. This method involves selecting and crossing plants with desirable resistance traits to develop new varieties that are resistant to specific pathogens. Through careful selection and breeding, plant breeders can introduce and amplify genes that confer resistance, resulting in improved disease resistance in subsequent generations. Advancements in genetic research and technology have aided breeders in identifying and incorporating specific resistance genes into commercial crop varieties more efficiently. The use of molecular markers and marker-assisted selection (MAS) techniques enables breeders to identify and track the presence of desired genes during the breeding process (Fig. 5). This targeted approach expedites the development of resistant varieties, saving time and resources compared to conventional breeding methods [336–344].

4.2 Genetic Modification Techniques for Pathogen Resistance

Genetic modification techniques, such as genetic engineering or transgenic approaches, offer another avenue to enhance plant resistance to pathogens. This method involves introducing genes from other organisms into the plant's genome to confer resistance against specific pathogens. By inserting genes encoding pathogen-derived resistance proteins or proteins involved in defense signaling pathways, scientists can create genetically modified (GM) crops with enhanced resistance [338,345,346]. One common genetic modification technique is the introduction of Bacillus thuringiensis (Bt) genes into crop plants to confer resistance against insect pests. Bt genes encode proteins toxic to specific insect pests, providing an effective means of pest control without the need for chemical insecticides [347–350]. Similarly, researchers have successfully engineered plants with increased tolerance to viral, fungal, and bacterial infections by introducing genes encoding antiviral proteins, antimicrobial peptides, or enzymes involved in defense mechanisms [351–357].

4.3 Case Studies Highlighting Successful Genetic Approaches

- (1) Bt cotton: Bt cotton is one of the most prominent examples of genetically modified crops exhibiting enhanced resistance. By introducing Bt genes into cotton plants, scientists have successfully conferred resistance against bollworms and other devastating insect pests. This has led to increased crop yield, reduced pesticide use, and improved economic outcomes for farmers [358,359].
- (2) Rainbow papaya: The rainbow papaya is an example of a genetically modified fruit that achieved resistance against the papaya ringspot virus (PRSV) [360–362]. By

introducing a gene from the PRSV into the papaya genome, researchers developed transgenic papaya varieties that exhibited strong resistance to the virus [363–365]. This genetic approach saved the Hawaiian papaya industry from severe losses caused by PRSV infection.

(3) Late blight-resistant potato: Late blight is a devastating fungal disease that affects potato plants worldwide. Researchers have utilized genetic modification techniques to introduce genes from wild potato relatives that confer resistance against late blight. By incorporating these resistance genes into commercial potato varieties, scientists have successfully developed genetically modified potatoes with enhanced resistance to late blight, reducing crop losses and the need for fungicide applications [366–369].

These case studies demonstrate the effectiveness of genetic approaches in enhancing plant resistance to pathogens. Genetic techniques offer targeted and precise solutions to combat specific diseases, providing sustainable options for improving crop productivity and reducing the reliance on chemical inputs. Nonetheless, it is crucial to consider the potential environmental and socioeconomic impacts associated with the deployment of genetically modified crops while ensuring rigorous safety assessments and regulatory frameworks are in place.

5. Cultural Practices for Pathogen Management

Cultural practices play a crucial role in managing and mitigating the impact of pathogens on crops. These practices involve various techniques and approaches that farmers can adopt to prevent, control, and reduce the occurrence of plant diseases. Cultural practices offer sustainable and environmentally friendly alternatives that promote long-term resilience in agricultural systems [370–373]. One of the primary benefits of cultural practices is their ability to create an unfavorable environment for pathogens. By modifying growing conditions, farmers can discourage pathogenic organisms from thriving and spreading. This approach reduces the reliance on chemical inputs and minimizes the risks associated with pesticide resistance and environmental contamination [374].

5.1 Crop Rotation

Crop rotation is a widely employed cultural practice that involves systematically changing the type of crops grown in a particular field over successive seasons. This practice disrupts the life cycles of many pathogens, as different plants have varying susceptibility to specific diseases. By alternating crops, farmers can break the cycle of pathogens that rely on a single host species for survival, reducing their population densities and preventing the buildup of soil-borne pathogens. Additionally, crop rotation can improve soil health, nutrient availability, and overall plant vigor, further enhancing the ability of crops to resist diseases [375–379].



Integrated Disease Management

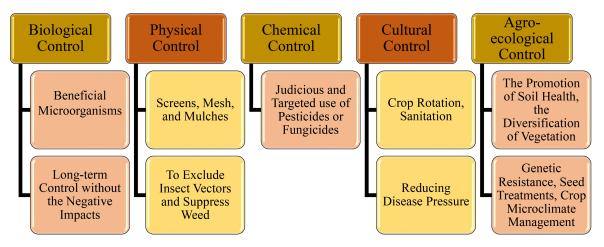


Fig. 6. Integrated Disease Management.

5.2 Sanitation

Sanitation practices focus on maintaining clean and pathogen-free farming environments. This includes removing infected plant debris, weeds, and diseased or dead plant material promptly. Infected plant residues left on the field provide a breeding ground for pathogens to survive and proliferate. Farmers should also regularly clean and disinfect tools, equipment, and machinery to prevent the transmission of pathogens between plants and fields. Proper sanitation measures minimize the chances of pathogen introduction and reduce inoculum levels, consequently lowering disease incidence and severity [380,381].

5.3 Proper Planting Techniques

Adopting appropriate planting techniques is another essential cultural practice for managing plant diseases. These techniques involve careful consideration of factors such as plant spacing, depth of planting, and timing of sowing [382–384]. By optimizing these parameters, farmers can promote better air circulation and light penetration within the crop canopy, reducing humidity and creating an unfavorable environment for many foliar pathogens. Additionally, proper planting techniques ensure optimal seedbed [385–389].

5.4 Irrigation Management

Water management practices have a significant impact on disease development in crops. Excessive or poorly timed irrigation can create favorable conditions for pathogens, especially those that thrive in moist environments [390–394]. On the other hand, deficit irrigation, as appropriate for specific crops, can induce stress responses in plants, making them more susceptible to certain diseases. By adopting pre-

cise and targeted irrigation methods, farmers can regulate moisture levels, minimize disease-friendly environments, and promote plant health and resilience [395–399].

6. Integrated Disease Management (IDM) Strategies

Integrated Disease Management (IDM) is an approach that combines various cultural, biological, physical, and chemical control methods to manage plant diseases effectively (Fig. 6) [400–403].

IDM strategies aim to integrate multiple approaches into a holistic management plan that maximizes disease control while minimizing negative environmental impacts and reducing reliance on any single control method [404–406]. The following are key components of IDM:

- (1) Cultural Control: As discussed earlier, cultural practices form the foundation of IDM. Crop rotation, sanitation, and other cultural practices play a vital role in preventing the establishment and spread of pathogens [407,408]. By adopting a comprehensive cultural control program, farmers can create an environment that promotes plant health and reduces disease pressure.
- (2) Biological Control: Biological control involves the use of beneficial organisms to suppress or control plant pathogens. This includes the introduction of natural enemies such as predatory insects, parasitic nematodes, and microorganisms that antagonize pathogens [409–412]. Biocontrol agents can help reduce pathogen populations, disrupt their life cycles, and provide long-term control without the negative impacts associated with chemical interventions.
- (3) Physical Control: Physical control methods focus on creating physical barriers or modifying the environment



to prevent pathogen entry or spread. This may involve the use of barriers like screens or mesh to exclude insect vectors [413,414], employing mulches to suppress weed growth [415–418], or implementing measures to improve air circulation and light penetration within plant canopies [419].

(4) Chemical Control: While the emphasis of IDM is on minimizing chemical inputs, judicious and targeted use of pesticides or fungicides can be part of an integrated approach. However, it is crucial to prioritize the use of low-risk or bio-based products and employ them only when other control methods are insufficient to manage disease outbreaks effectively. Integrated pest management (IPM) principles should guide the application of chemicals to minimize their impact on non-target organisms and the environment.

(5) Agro-ecological control: the objective of agroecological control is to promote healthy crops. The principles of agro ecological crop protection are identical to the principles of agro ecology from which they originate and they are inspired by certain principles of crop protection used in organic agriculture [403]. Agro ecological crop protection is a concept of crop protection based on cropping systems whose aim is to improve the sustainability of agroecosystems by taking into account their ecological functioning. Agro ecological crop protection aims to promote the ecological health of agroecosystems by directly or indirectly optimizing interactions between living (plant, animal, microbial) communities both below and above the ground. Agro ecological crop protection considers both biodiversity and soil health, to make agroecosystems less susceptible to biotic stresses [403,420]. Agricultural management of diseases implies a comprehensive approach, in which farmers first consider the characteristics of the type of agriculture and its socio-economic context. Plant pathogens can be understood as a community or assemblage of organisms that includes a large genetics. Their functional diversity can be determined by using functional groups that are in accordance with their relationship with the design and management of system agriculture. These have different relationships with major agricultural management strategies, such as vegetation diversity, soil health promotion and landscape heterogeneity. Therefore, there is a need to complete general strategies including seed disinfection and selection of genotypes and cultivars with high genetic capability (resistant to living and non-living stresses) [403].

In conclusion, cultural practices form a cornerstone in the management of plant diseases. By implementing techniques such as crop rotation, sanitation, and adopting integrated disease management (IDM) strategies, farmers can create an environment that suppresses pathogens, enhances plant resistance, and promotes sustainable agricultural systems. Therefore, paying attention to all aspects of disease resistance, especially genetic resistance, and paying attention to the ecosystem is one of the basic criteria in agro ecological control.

7. Chemical Elicitors as Plant Resistance Inducers

7.1 Definition and Characteristics of Chemical Elicitors

Chemical elicitors are compounds that stimulate the defense responses in plants, leading to enhanced resistance against pathogens, pests, and environmental stresses. These elicitors can be naturally occurring or synthetic compounds that mimic the signals produced by pathogens during an attack. When applied to plants, chemical elicitors trigger a cascade of biochemical and physiological responses that ultimately result in the activation of various defense mechanisms [421-424]. Chemical elicitors possess certain characteristics that make them effective in inducing plant resistance. Firstly, they have a high degree of specificity, meaning they can selectively activate specific defense pathways without causing harm to the plant. This specificity allows plants to allocate their resources efficiently towards combating the particular stress they are facing [423,425,426]. Secondly, chemical elicitors typically have low phytotoxicity, meaning they do not cause significant damage to plant tissues when applied at recommended concentrations. This is crucial for the practical application of these elicitors, as excessive phytotoxicity could offset the benefits gained from enhanced resistance [427,428].

Furthermore, effective chemical elicitors are stable and easily applicable. They should remain active under a range of environmental conditions and be compatible with common application methods such as foliar sprays, seed treatments, or soil drenches. This ensures that the elicitor can be conveniently integrated into existing agricultural practices.

7.2 Modes of Action and Effectiveness of Chemical Elicitors

Chemical elicitors exert their effects through various modes of action, targeting different stages of the plant's defense response. One common mode of action involves the recognition of the elicitor by specific receptors on the plant's cell surface. This recognition triggers a signaling cascade that leads to the activation of defense-related genes and the production of defense compounds [36,429]. Another mode of action involves the stimulation of the plant's SAR pathway. SAR is a long-lasting defense response that occurs throughout the entire plant following an initial localized attack. Chemical elicitors can induce SAR by activating the production of signaling molecules, such as salicylic acid, that initiate the defense response in distal parts of the plant [429-431]. Chemical elicitors can also enhance the activity of certain enzymes involved in defense responses, such as chitinases or glucanases, which degrade the cell walls of pathogens. By increasing the activity of these enzymes, chemical elicitors contribute to a more effective defense against invading pathogens [432–436]. The effectiveness of chemical elicitors varies depending on several factors. Firstly, the specific elicitor and its mode of action play



a crucial role. Different pathogens may trigger different defense pathways in plants, so an effective elicitor should be capable of activating the appropriate defense mechanisms for the given stress. Additionally, the timing and method of elicitor application are important. Applying the elicitor before the pathogen attack or at the early stages of infection often yields better results.

Furthermore, environmental conditions can influence the efficacy of chemical elicitors. Factors such as temperature, humidity, and light intensity can affect the responsiveness of plants to elicitors, highlighting the need for tailored application strategies based on the local environmental conditions.

7.3 Considerations and Challenges in the Use of Chemical Elicitors

While chemical elicitors hold significant potential as plant resistance inducers, there are several considerations and challenges associated with their use. One key consideration is the cost-effectiveness of incorporating elicitors into agricultural practices. Currently, many commercially available elicitors are relatively expensive, limiting their widespread adoption, particularly among small-scale farmers. Therefore, efforts are underway to develop costeffective formulations and delivery methods to make these products more accessible [437-443]. Another challenge is the potential development of resistance in pathogens towards the elicitors. Continuous and extensive use of specific elicitors can exert selective pressure on the target organisms, potentially leading to the emergence of resistant strains. To mitigate this risk, it is important to rotate or combine different elicitors with distinct modes of action, making it more difficult for pathogens to develop resistance [444.445].

Formulation and stability of chemical elicitors also pose challenges. Elicitors can degrade rapidly under certain conditions, reducing their shelf life and effectiveness. Developing stable formulations that ensure the long-term viability of the active compounds is crucial for practical application. Furthermore, the potential impact of chemical elicitors on non-target organisms and the environment must be carefully evaluated. While efforts are made to select elicitors with low toxicity and environmental persistence, comprehensive studies are needed to assess any potential risks associated with their use. Lastly, understanding the complex signaling networks and interactions between elicitors and plants is an ongoing challenge. Elucidating these mechanisms would enable the design of more specific and efficient elicitors and their optimal application strategies. In conclusion, chemical elicitors have emerged as valuable tools for boosting plant resistance against pathogens, pests, and environmental stresses. Their specificity, low phytotoxicity, and ease of application make them attractive options for integrated pest management and sustainable agriculture. However, addressing the considerations and challenges associated with their use is essential to maximize their efficacy and minimize potential risks. Continued research and development in this field hold promise for harnessing the full potential of chemical elicitors in promoting plant health and crop productivity.

8. Future Directions and Challenges

8.1 Emerging Research Trends in Plant Resistance Stimulation

As the field of plant resistance continues to evolve, several emerging research trends are shaping the future direction of this area. Researchers are increasingly focusing on understanding the molecular mechanisms underlying plant resistance and exploring innovative approaches to stimulate resistance in plants. One key trend is the exploration of plant immune system components, such as pattern recognition receptors (PRRs) and effector-triggered immunity (ETI), to develop effective strategies for enhancing resistance. Additionally, there is growing interest in studying the role of plant hormones, such as salicylic acid and jasmonic acid, in modulating plant resistance responses. Furthermore, researchers are investigating the potential of microbiome manipulation to enhance plant resistance through beneficial interactions between plants and associated microorganisms.

8.2 Potential Applications of New Technologies (e.g., CRISPR) in Enhancing Plant Resistance

The advent of new technologies, particularly CRISPR-Cas9 gene editing, offers exciting opportunities for enhancing plant resistance [446]. CRISPR enables precise and targeted modifications of plant genomes, allowing researchers to manipulate genes involved in plant defense pathways. By engineering specific genes associated with resistance, it may be possible to develop crops with enhanced protection against pathogens, pests, and environmental stresses. CRISPR technology also holds potential for accelerating the breeding process by introducing desirable traits into crop plants, ultimately leading to improved resistance profiles [447–449]. However, ethical considerations, regulatory frameworks, and public acceptance will need to be addressed to ensure responsible and sustainable use of these technologies.

8.3 Addressing Challenges and Limitations in the Field

Despite significant advancements, several challenges and limitations persist in the field of plant resistance. One major challenge is the rapid evolution of pathogens and pests, which can render resistance mechanisms ineffective over time [450–452]. Developing durable resistance strategies that can withstand evolving threats remains a key priority [453–456]. Another challenge lies in translating laboratory-based research findings into practical applications in agriculture [457]. Scaling up resistance-enhancing techniques and ensuring their efficacy under field conditions is crucial for widespread adoption.



Additionally, addressing the potential trade-offs between plant growth and resistance is essential to avoid compromising crop productivity [458,459]. Finally, there is a need to consider the ecological and environmental impacts of resistance-enhancing technologies to ensure they do not disrupt natural ecosystems or harm beneficial organisms.

In conclusion, future directions in plant resistance research involve exploring emerging trends such as understanding plant immune system components, studying plant hormones and microbiome interactions, and leveraging new technologies like CRISPR for enhancing resistance. However, addressing challenges related to pathogen evolution, translating research into practical applications, managing trade-offs, and considering ecological implications will be critical in advancing the field and maximizing its positive impact on agriculture and food security.

9. Conclusion

In conclusion, this paper has explored various methods to stimulate plant resistance against pathogens and highlighted the importance of adopting integrated approaches for sustainable disease management. Throughout the discussion, several key methods have been identified as effective means to enhance plant resistance against pathogens. These methods include:

- (1) Genetic approaches: Breeding resistant plant varieties through traditional breeding techniques or genetic engineering can provide plants with inherent resistance against specific pathogens.
- (2) Cultural practices: Implementing proper sanitation measures, crop rotation, and intercropping can disrupt pathogen life cycles and reduce disease incidence.
- (3) Chemical control: The judicious use of fungicides and other chemical agents can help manage diseases in agricultural settings. However, it is important to use these chemicals responsibly to minimize negative impacts on the environment and human health.
- (4) Biological control: Utilizing beneficial organisms such as biocontrol agents, antagonistic microbes, and predatory insects can effectively suppress pathogen populations and protect plants from diseases.

It is crucial to recognize that no single method alone can provide comprehensive protection against plant pathogens. Integrated approaches that combine multiple strategies are essential for sustainable disease management. By integrating genetic, cultural, chemical, and biological control methods, farmers and researchers can develop holistic and long-term solutions that mitigate the risk of disease outbreaks. Integrated disease management not only enhances plant resistance but also minimizes the reliance on chemical inputs, reduces environmental pollution, preserves natural resources, and promotes the overall health and sustainability of agricultural systems. While significant progress has been made in understanding plant-pathogen interactions and developing resistance strategies, there is

still much to learn. Further research is needed to identify new resistance mechanisms, improve the efficacy of existing methods, and develop innovative approaches for disease management.

Additionally, it is essential to bridge the gap between scientific knowledge and practical implementation. Farmers, extension services, and policymakers should be encouraged to adopt and promote sustainable disease management practices. This can be achieved through educational programs, outreach initiatives, and policy support at local, national, and international levels.

In conclusion, addressing plant diseases requires a collaborative effort from researchers, farmers, industry stakeholders, and policymakers. By investing in research, implementing effective strategies, and promoting sustainable practices, we can better protect our crops, enhance food security, and ensure the long-term resilience of agricultural systems in the face of evolving pathogen threats.

Author Contributions

RSR designed the research study. RSR performed the research. MGV provided help and advice on figures. RSR and MGV wrote the manuscript. Both authors contributed to editorial changes in the manuscript. Both authors read and approved the final manuscript. Both authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

The authors would like to acknowledge Vali-e-Asr University of Rafsanjan.

Funding

This research received no external funding.

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Ramawat K, Ahuja M. Fiber plants: an overview. Fiber Plants: Biology, Biotechnology and Applications. 2016; 3–15.
- [2] Thakur M, Bhattacharya S, Khosla PK, Puri S. Improving production of plant secondary metabolites through biotic and abiotic elicitation. Journal of Applied Research on Medicinal and Aromatic Plants. 2019; 12: 1–12.
- [3] Tsedaley B. Review on seed health tests and detection methods of seed-borne diseases. Journal of Biology, Agriculture and Healthcare. 2015; 5.
- [4] Saberi Riseh R, Vatankhah M, Hassanisaadi M, Barka EA. Unveiling the Role of Hydrolytic Enzymes from Soil Biocontrol Bacteria in Sustainable Phytopathogen Management. Frontiers in Bioscience (Landmark Edition). 2024; 29: 105.



- [5] Brooks DR, Hoberg EP, Boeger WA, Trivellone V. Emerging infectious disease: An underappreciated area of strategic concern for food security. Transboundary and Emerging Diseases. 2022; 69: 254–267.
- [6] McDonald BA, Stukenbrock EH. Rapid emergence of pathogens in agro-ecosystems: global threats to agricultural sustainability and food security. Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences. 2016; 371: 20160026.
- [7] Saberi Riseh R, Vatankhah M, Hassanisaadi M, Kennedy JF. Chitosan-based nanocomposites as coatings and packaging materials for the postharvest improvement of agricultural product: A review. Carbohydrate Polymers. 2023; 309: 120666.
- [8] Garcia-Ruiz H, Szurek B, Van den Ackerveken G. Stop helping pathogens: engineering plant susceptibility genes for durable resistance. Current Opinion in Biotechnology, 2021; 70: 187–195.
- [9] Kumar D. Plant immune response strategies against pathogens. Plant Archives (09725210). 2020; 20.
- [10] Shittu HO, Aisagbonhi E, Onyemaechi HO. PLANTS'INNATE DEFENCE MECHANISMS AGAINST PHYTOPATHOGENS. The Journal of Microbiology, Biotechnology and Food Sciences. 2019: 9: 314.
- [11] Song XP, Verma KK, Tian DD, Zhang XQ, Liang YJ, Huang X, *et al.* Exploration of silicon functions to integrate with biotic stress tolerance and crop improvement. Biological Research. 2021; 54: 19.
- [12] Mishra S, Singh A, Keswani C, Saxena A, Sarma B, Singh H. Harnessing plant-microbe interactions for enhanced protection against phytopathogens. In Arora NK (ed.) Plant microbes symbiosis: applied facets (pp. 111–125). Springer: New Delhi. 2015.
- [13] Barea J. Future challenges and perspectives for applying microbial biotechnology in sustainable agriculture based on a better understanding of plant-microbiome interactions. Journal of Soil Science and Plant Nutrition. 2015; 15: 261–282.
- [14] Lamichhane JR, Dachbrodt-Saaydeh S, Kudsk P, Messéan A. Toward a Reduced Reliance on Conventional Pesticides in European Agriculture. Plant Disease. 2016; 100: 10–24.
- [15] Saberi Riseh R. Advancing agriculture through bioresource technology: The role of cellulose-based biodegradable mulches. International Journal of Biological Macromolecules. 2024; 255: 128006
- [16] Hassanisaadi M, Saberi Riseh R, Rabiei A, Varma RS, Kennedy JF. Nano/micro-cellulose-based materials as remarkable sorbents for the remediation of agricultural resources from chemical pollutants. International Journal of Biological Macromolecules. 2023; 246: 125763.
- [17] He DC, Zhan JS, Xie LH. Problems, challenges and future of plant disease management: from an ecological point of view. Journal of Integrative Agriculture. 2016; 15: 705-715.
- [18] Brzozowski L, Mazourek M. A sustainable agricultural future relies on the transition to organic agroecological pest management. Sustainability. 2018; 10: 2023.
- [19] Gomiero T, Pimentel D, Paoletti MG. Environmental impact of different agricultural management practices: conventional vs. organic agriculture. Critical Reviews in Plant Sciences. 2011; 30: 95–124.
- [20] Miah G, Rafii MY, Ismail MR, Puteh AB, Rahim HA, Asfaliza R, et al. Blast resistance in rice: a review of conventional breeding to molecular approaches. Molecular Biology Reports. 2013; 40: 2369–2388.
- [21] Yadav RK, Tripathi MK, Tiwari S, Tripathi N, Asati R, Patel V, et al. Breeding and Genomic Approaches towards Development of Fusarium Wilt Resistance in Chickpea. Life. 2023; 13: 988.
- [22] Acquaah G. Conventional plant breeding principles and techniques. In Al-Khayri JM, Jain SM, Johnson DV (eds.) Advances in plant breeding strategies: Breeding, biotechnology

- and molecular tools (pp. 115-158). Springer: Cham. 2015.
- [23] Oladosu Y, Rafii MY, Abdullah N, Hussin G, Ramli A, Rahim HA, *et al.* Principle and application of plant mutagenesis in crop improvement: a review. Biotechnology & Biotechnological Equipment. 2016; 30: 1–16.
- [24] Wakchaure R, Ganguly S, Praveen P, Kumar A, Sharma S, Mahajan T. Marker assisted selection (MAS) in animal breeding: a review. Journal of Drug Metabolism and Toxicology. 2015; 6: e127.
- [25] Mall T, Gupta M, Dhadialla TS, Rodrigo S. Overview of Biotechnology-Derived Herbicide Tolerance and Insect Resistance Traits in Plant Agriculture. Methods in Molecular Biology. 2019; 1864: 313–342.
- [26] Ricroch AE, Hénard-Damave MC. Next biotech plants: new traits, crops, developers and technologies for addressing global challenges. Critical Reviews in Biotechnology. 2016; 36: 675– 690.
- [27] Dong OX, Ronald PC. Genetic Engineering for Disease Resistance in Plants: Recent Progress and Future Perspectives. Plant Physiology. 2019; 180: 26–38.
- [28] Slavokhotova AA, Naumann TA, Price NPJ, Rogozhin EA, Andreev YA, Vassilevski AA, et al. Novel mode of action of plant defense peptides hevein-like antimicrobial peptides from wheat inhibit fungal metalloproteases. The FEBS Journal. 2014; 281: 4754–4764.
- [29] Castro MS, Fontes W. Plant defense and antimicrobial peptides. Protein and Peptide Letters. 2005; 12: 13–18.
- [30] Deb S, Choudhury A, Kharbyngar B, Satyawada RR. Applications of CRISPR/Cas9 technology for modification of the plant genome. Genetica. 2022; 150: 1–12.
- [31] Boubakri H. Recent progress in CRISPR/Cas9-based genome editing for enhancing plant disease resistance. Gene. 2023; 866: 147334.
- [32] Arora L, Narula A. Gene Editing and Crop Improvement Using CRISPR-Cas9 System. Frontiers in Plant Science. 2017; 8: 1932.
- [33] Igiehon NO, Babalola OO. Below-ground-above-ground Plant-microbial Interactions: Focusing on Soybean, Rhizobacteria and Mycorrhizal Fungi. The Open Microbiology Journal. 2018; 12: 261–279.
- [34] Pozo MJ, Jung SC, Martínez-Medina A, López-Ráez JA, Azcón-Aguilar C, Barea J-M. Root allies: arbuscular mycorrhizal fungi help plants to cope with biotic stresses. In Aroca R (ed.) Symbiotic endophytes (pp. 289–307). Springer: Berlin, Heidelberg. 2012
- [35] Cameron DD, Neal AL, van Wees SCM, Ton J. Mycorrhizainduced resistance: more than the sum of its parts? Trends in Plant Science. 2013; 18: 539–545.
- [36] Meena M, Yadav G, Sonigra P, Nagda A, Mehta T, Swapnil P, et al. Role of elicitors to initiate the induction of systemic resistance in plants to biotic stress. Plant Stress. 2022; 5: 100103.
- [37] Burketova L, Trda L, Ott PG, Valentova O. Bio-based resistance inducers for sustainable plant protection against pathogens. Biotechnology Advances. 2015; 33: 994–1004.
- [38] Pieterse CMJ, Zamioudis C, Berendsen RL, Weller DM, Van Wees SCM, Bakker PAHM. Induced systemic resistance by beneficial microbes. Annual Review of Phytopathology. 2014; 52: 347–375.
- [39] Zhou C, Zhou Y. Strategies for viral cross protection in plants. Methods in Molecular Biology. 2012; 894: 69–81.
- [40] Zhang XF, Qu F. Cross protection of plant viruses: recent developments and mechanistic implications. In Wang AM, Zhou XP (eds.) Current research topics in plant virology (pp. 241–250). Springer: Cham. 2016.
- [41] Ryals J, Uknes S, Ward E. Systemic Acquired Resistance. Plant Physiology. 1994; 104: 1109–1112.



- [42] Pérez-de-Luque A, Tille S, Johnson I, Pascual-Pardo D, Ton J, Cameron DD. The interactive effects of arbuscular mycorrhiza and plant growth-promoting rhizobacteria synergistically enhance host plant defences against pathogens. Scientific Reports. 2017; 7: 16409.
- [43] Pozo MJ, Verhage A, García-Andrade J, García JM, Azcón-Aguilar C. Priming plant defence against pathogens by arbuscular mycorrhizal fungi. In Azcón-Aguilar C, Barea JM, Gianinazzi S, Gianinazzi-Pearson V (eds.) Mycorrhizas-functional processes and ecological impact (pp. 123–135). Springer: Berlin, Heidelberg. 2009.
- [44] Mishra V, Gupta A, Kaur P, Singh S, Singh N, Gehlot P, et al. Synergistic effects of Arbuscular mycorrhizal fungi and plant growth promoting rhizobacteria in bioremediation of iron contaminated soils. International Journal of Phytoremediation. 2016; 18: 697–703.
- [45] Mahlein AK, Heim RHJ, Brugger A, Gold K, Li Y, Bashir AK, *et al.* Digital plant pathology for precision agriculture. Journal of Plant Diseases and Protection. 2022; 129: 455–456.
- [46] Ampt EA, van Ruijven J, Raaijmakers JM, Termorshuizen AJ, Mommer L. Linking ecology and plant pathology to unravel the importance of soil-borne fungal pathogens in species-rich grasslands. European Journal of Plant Pathology. 2019; 154: 141– 156.
- [47] Velásquez AC, Castroverde CDM, He SY. Plant-Pathogen Warfare under Changing Climate Conditions. Current Biology. 2018; 28: R619–R634.
- [48] Daayf F, El Hadrami A, El-Bebany AF, Henriquez MA, Yao Z, Derksen H, *et al.* Phenolic compounds in plant defense and pathogen counter-defense mechanisms. Recent Advances in Polyphenol Research. 2012; 3: 191–208.
- [49] Faulkner C, Robatzek S. Plants and pathogens: putting infection strategies and defence mechanisms on the map. Current Opinion in Plant Biology. 2012; 15: 699–707.
- [50] Lacombe S, Rougon-Cardoso A, Sherwood E, Peeters N, Dahlbeck D, van Esse HP, et al. Interfamily transfer of a plant pattern-recognition receptor confers broad-spectrum bacterial resistance. Nature Biotechnology. 2010; 28: 365–369.
- [51] Boller T, Felix G. A renaissance of elicitors: perception of microbe-associated molecular patterns and danger signals by pattern-recognition receptors. Annual Review of Plant Biology. 2009; 60: 379–406.
- [52] Abramovitch RB, Anderson JC, Martin GB. Bacterial elicitation and evasion of plant innate immunity. Nature Reviews. Molecular Cell Biology. 2006; 7: 601–611.
- [53] Song W, Forderer A, Yu D, Chai J. Structural biology of plant defence. The New Phytologist. 2021; 229: 692–711.
- [54] Teixeira RM, Ferreira MA, Raimundo GAS, Loriato VAP, Reis PAB, Fontes EPB. Virus perception at the cell surface: revisiting the roles of receptor-like kinases as viral pattern recognition receptors. Molecular Plant Pathology. 2019; 20: 1196–1202.
- [55] Rahman A, Sinha KV, Sopory SK, Sanan-Mishra N. Influence of virus-host interactions on plant response to abiotic stress. Plant Cell Reports. 2021; 40: 2225–2245.
- [56] Sun L, Zhang J. Regulatory role of receptor-like cytoplasmic kinases in early immune signaling events in plants. FEMS Microbiology Reviews. 2020; 44: 845–856.
- [57] Fei W, Liu Y. Biotrophic Fungal Pathogens: a Critical Overview. Applied Biochemistry and Biotechnology. 2023; 195: 1–16.
- [58] Mapuranga J, Zhang N, Zhang L, Chang J, Yang W. Infection Strategies and Pathogenicity of Biotrophic Plant Fungal Pathogens. Frontiers in Microbiology. 2022; 13: 799396.
- [59] Jaswal R, Kiran K, Rajarammohan S, Dubey H, Singh PK, Sharma Y, et al. Effector Biology of Biotrophic Plant Fungal Pathogens: Current Advances and Future Prospects. Microbiological Research. 2020; 241: 126567.

- [60] Vo KTX, Yi Q, Jeon JS. Engineering effector-triggered immunity in rice: Obstacles and perspectives. Plant, Cell & Environment. 2023; 46: 1143–1156.
- [61] Remick BC, Gaidt MM, Vance RE. Effector-Triggered Immunity. Annual Review of Immunology. 2023; 41: 453–481.
- [62] Leonetti P, Stuttmann J, Pantaleo V. Regulation of plant antiviral defense genes via host RNA-silencing mechanisms. Virology Journal. 2021; 18: 194.
- [63] Saur IML, Hückelhoven R. Recognition and defence of plantinfecting fungal pathogens. Journal of Plant Physiology. 2021; 256: 153324.
- [64] Lukan T, Županič A, Mahkovec Povalej T, Brunkard JO, Kmetič M, Juteršek M, et al. Chloroplast redox state changes mark cell-to-cell signaling in the hypersensitive response. The New Phytologist. 2023; 237: 548–562.
- [65] Ding LN, Li YT, Wu YZ, Li T, Geng R, Cao J, et al. Plant Disease Resistance-Related Signaling Pathways: Recent Progress and Future Prospects. International Journal of Molecular Sciences. 2022; 23: 16200.
- [66] Kumar J, Ramlal A, Kumar K, Rani A, Mishra V. Signaling Pathways and Downstream Effectors of Host Innate Immunity in Plants. International Journal of Molecular Sciences. 2021; 22: 9022
- [67] Naveed ZA, Wei X, Chen J, Mubeen H, Ali GS. The PTI to ETI Continuum in *Phytophthora*-Plant Interactions. Frontiers in Plant Science. 2020; 11: 593905.
- [68] Mo S, Qian Y, Zhang W, Qian L, Wang Y, Cailin G, et al. Mitogen-activated protein kinase action in plant response to high-temperature stress: a mini review. Protoplasma. 2021; 258: 477–482.
- [69] Jiang M, Zhang Y, Li P, Jian J, Zhao C, Wen G. Mitogen-Activated Protein Kinase and Substrate Identification in Plant Growth and Development. International Journal of Molecular Sciences. 2022; 23: 2744.
- [70] Swaminathan S, Lionetti V, Zabotina OA. Plant Cell Wall Integrity Perturbations and Priming for Defense. Plants. 2022; 11: 3539.
- [71] Abbas A, Mubeen M, Zheng H, Sohail MA, Shakeel Q, Solanki MK, et al. Trichoderma spp. Genes Involved in the Biocontrol Activity Against Rhizoctonia solani. Frontiers in Microbiology. 2022; 13: 884469.
- [72] Wu J, Zhao Q, Yang Q, Liu H, Li Q, Yi X, et al. Comparative transcriptomic analysis uncovers the complex genetic network for resistance to Sclerotinia sclerotiorum in Brassica napus. Scientific Reports. 2016; 6: 19007.
- [73] An C, Mou Z. Salicylic acid and its function in plant immunity. Journal of Integrative Plant Biology. 2011; 53: 412–428.
- [74] Li N, Han X, Feng D, Yuan D, Huang LJ. Signaling Crosstalk between Salicylic Acid and Ethylene/Jasmonate in Plant Defense: Do We Understand What They Are Whispering? International Journal of Molecular Sciences. 2019; 20: 671.
- [75] Tanaka S, Han X, Kahmann R. Microbial effectors target multiple steps in the salicylic acid production and signaling pathway. Frontiers in Plant Science. 2015; 6: 349.
- [76] Niu D, Wang X, Wang Y, Song X, Wang J, Guo J, et al. Bacillus cereus AR156 activates PAMP-triggered immunity and induces a systemic acquired resistance through a NPR1-and SA-dependent signaling pathway. Biochemical and Biophysical Research Communications. 2016; 469: 120–125.
- [77] Shine MB, Xiao X, Kachroo P, Kachroo A. Signaling mechanisms underlying systemic acquired resistance to microbial pathogens. Plant Science. 2019; 279: 81–86.
- [78] Wang L, Liu S, Gao M, Wang L, Wang L, Wang Y, et al. The Crosstalk of the Salicylic Acid and Jasmonic Acid Signaling Pathways Contributed to Different Resistance to Phytoplasma Infection Between the Two Genotypes in Chinese Jujube. Fron-



- tiers in Microbiology. 2022; 13: 800762.
- [79] Vlot AC, Sales JH, Lenk M, Bauer K, Brambilla A, Sommer A, et al. Systemic propagation of immunity in plants. The New Phytologist. 2021; 229: 1234–1250.
- [80] Carvalhais LC, Schenk PM, Dennis PG. Jasmonic acid signalling and the plant holobiont. Current Opinion in Microbiology. 2017; 37: 42–47.
- [81] Costarelli A, Bianchet C, Ederli L, Salerno G, Piersanti S, Rebora M, et al. Salicylic acid induced by herbivore feeding antagonizes jasmonic acid mediated plant defenses against insect attack. Plant Signaling & Behavior. 2020; 15: 1704517.
- [82] Ali MS, Baek KH. Jasmonic Acid Signaling Pathway in Response to Abiotic Stresses in Plants. International Journal of Molecular Sciences. 2020; 21: 621.
- [83] Marquis V, Smirnova E, Graindorge S, Delcros P, Villette C, Zumsteg J, et al. Broad-spectrum stress tolerance conferred by suppressing jasmonate signaling attenuation in Arabidopsis JASMONIC ACID OXIDASE mutants. The Plant Journal: for Cell and Molecular Biology. 2022; 109: 856–872.
- [84] Lorenzo O, Solano R. Molecular players regulating the jasmonate signalling network. Current Opinion in Plant Biology. 2005: 8: 532–540.
- [85] Leon-Reyes A, Du Y, Koornneef A, Proietti S, Körbes AP, Memelink J, et al. Ethylene signaling renders the jasmonate response of Arabidopsis insensitive to future suppression by salicylic Acid. Molecular Plant-Microbe Interactions. 2010; 23: 187–197.
- [86] Sitarek P, Merecz-Sadowska A, Kowalczyk T, Wieczfinska J, Zajdel R, Śliwiński T. Potential Synergistic Action of Bioactive Compounds from Plant Extracts against Skin Infecting Microorganisms. International Journal of Molecular Sciences. 2020; 21: 5105
- [87] Dora S, Terrett OM, Sánchez-Rodríguez C. Plant-microbe interactions in the apoplast: Communication at the plant cell wall. The Plant Cell. 2022; 34: 1532–1550.
- [88] Ha CM, Rao X, Saxena G, Dixon RA. Growth-defense tradeoffs and yield loss in plants with engineered cell walls. The New Phytologist. 2021; 231: 60–74.
- [89] Kariyat RR, Gaffoor I, Sattar S, Dixon CW, Frock N, Moen J, et al. Sorghum 3-Deoxyanthocyanidin Flavonoids Confer Resistance against Corn Leaf Aphid. Journal of Chemical Ecology. 2019; 45: 502–514.
- [90] Kaur S. Estimating the Functional Consequences of Intra-Specific Trichome Variation of Tomato, Solanum lycopersicum on the Specialist Herbivore, Manduca sexta [Doctoral of thesis]. The University of Texas Rio Grande Valley. 2023.
- [91] Meena SS, Mankoti M, Rout PR, Mohanty A. Plant-microbe interactions and its effect on crop productivity. In Nayak SK, Baliyarsingh B, Singh A, Mannazzu I, Mishra BB (eds.) Advances in Agricultural and Industrial Microbiology: Volume-2: Applications of Microbes for Sustainable Agriculture and in-silico Strategies (pp. 29–60). Springer: Singapore. 2022.
- [92] Dutton C, Hôrak H, Hepworth C, Mitchell A, Ton J, Hunt L, et al. Bacterial infection systemically suppresses stomatal density. Plant, Cell & Environment. 2019; 42: 2411–2421.
- [93] Bharath P, Gahir S, Raghavendra AS. Abscisic Acid-Induced Stomatal Closure: An Important Component of Plant Defense Against Abiotic and Biotic Stress. Frontiers in Plant Science. 2021; 12: 615114.
- [94] Xiang Q, Lott AA, Assmann SM, Chen S. Advances and perspectives in the metabolomics of stomatal movement and the disease triangle. Plant Science. 2021; 302: 110697.
- [95] Cespedes CL, Aqueveque PM, Avila JG, Alarcon J, Kubo I. New advances in chemical defenses of plants: researches in calceolariaceae. Phytochemistry Reviews. 2015; 14: 367–380.
- [96] Xu Y, Xu L, He H. Chemicals mediate the interaction between

- plant-associated fungi and insects. Journal of Systematics and Evolution. 2023; 61: 506-517.
- [97] Chowdhary VA, Tank JG. Biomolecules Regulating Defense Mechanism in Plants. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences. 2023; 93: 17– 25
- [98] Ninkuu V, Zhang L, Yan J, Fu Z, Yang T, Zeng H. Biochemistry of Terpenes and Recent Advances in Plant Protection. International Journal of Molecular Sciences. 2021; 22: 5710.
- [99] Slavković F, Bendahmane A. Floral Phytochemistry: Impact of Volatile Organic Compounds and Nectar Secondary Metabolites on Pollinator Behavior and Health. Chemistry & Biodiversity. 2023; 20: e202201139.
- [100] Tiku AR. Antimicrobial compounds and their role in plant defense. In Singh A, Singh IK (eds.) Molecular aspects of plant-pathogen interaction (pp. 283–307). Springer: Singapore. 2018.
- [101] Baetz U, Martinoia E. Root exudates: the hidden part of plant defense. Trends in Plant Science. 2014; 19: 90–98.
- [102] Al-Khayri JM, Rashmi R, Toppo V, Chole PB, Banadka A, Sudheer WN, et al. Plant Secondary Metabolites: The Weapons for Biotic Stress Management. Metabolites. 2023; 13: 716.
- [103] Frey M, Chomet P, Glawischnig E, Stettner C, Grün S, Winklmair A, *et al.* Analysis of a chemical plant defense mechanism in grasses. Science. 1997; 277: 696–699.
- [104] Wouters FC, Blanchette B, Gershenzon J, Vassão DG. Plant defense and herbivore counter-defense: benzoxazinoids and insect herbivores. Phytochemistry Reviews: Proceedings of the Phytochemical Society of Europe. 2016; 15: 1127–1151.
- [105] Niu Y, Zhao X, Chao W, Lu P, Bai X, Mao T. Genetic Variation, DIMBOA Accumulation, and Candidate Gene Identification in Maize Multiple Insect-Resistance. International Journal of Molecular Sciences. 2023; 24: 2138.
- [106] Alvarado V, Scholthof HB. Plant responses against invasive nucleic acids: RNA silencing and its suppression by plant viral pathogens. Seminars in Cell & Developmental Biology. 2009; 20: 1032–1040.
- [107] Kumar K, Mandal SN, Neelam K, de Los Reyes BG. MicroRNA-mediated host defense mechanisms against pathogens and herbivores in rice: balancing gains from genetic resistance with trade-offs to productivity potential. BMC Plant Biology. 2022; 22: 351.
- [108] Boualem A, Dogimont C, Bendahmane A. The battle for survival between viruses and their host plants. Current Opinion in Virology. 2016; 17: 32–38.
- [109] Simón-Mateo C, García JA. Antiviral strategies in plants based on RNA silencing. Biochimica et Biophysica Acta. 2011; 1809: 722–731.
- [110] Duan CG, Wang CH, Guo HS. Application of RNA silencing to plant disease resistance. Silence. 2012; 3: 5.
- [111] Gago-Zachert S, Schuck J, Weinholdt C, Knoblich M, Pantaleo V, Grosse I, *et al.* Highly efficacious antiviral protection of plants by small interfering RNAs identified in vitro. Nucleic Acids Research. 2019; 47: 9343–9357.
- [112] Sioud M. Recent advances in small interfering RNA sensing by the immune system. New Biotechnology. 2010; 27: 236–242.
- [113] Katiyar-Agarwal S, Jin H. Role of small RNAs in host-microbe interactions. Annual Review of Phytopathology. 2010; 48: 225– 246.
- [114] Rodrigues M, Fan J, Lyon C, Wan M, Hu Y. Role of Extracellular Vesicles in Viral and Bacterial Infections: Pathogenesis, Diagnostics, and Therapeutics. Theranostics. 2018; 8: 2709–2721.
- [115] Zhu H, Duan CG, Hou WN, Du QS, Lv DQ, Fang RX, et al. Satellite RNA-derived small interfering RNA satsiR-12 targeting the 3' untranslated region of Cucumber mosaic virus triggers viral RNAs for degradation. Journal of Virology. 2011; 85: 13384–13397.



- [116] Ramesh SV, Yogindran S, Gnanasekaran P, Chakraborty S, Winter S, Pappu HR. Virus and Viroid-Derived Small RNAs as Modulators of Host Gene Expression: Molecular Insights Into Pathogenesis. Frontiers in Microbiology. 2021; 11: 614231.
- [117] Tabassum B, Nasir IA, Aslam U, Husnain T. How RNA interference combat viruses in plants. In Meroni G, Petrera F (eds.) Functional Genomics (pp. 113–130). InTech: Rijeka. 2012.
- [118] Chen W, Ye T, Sun Q, Niu T, Zhang J. Arbuscular mycorrhizal fungus alleviates anthracnose disease in tea seedlings. Frontiers in Plant Science. 2023; 13: 1058092.
- [119] Naqvi SAH, Wang J, Malik MT, Umar UUD, Hasnain A, Sohail MA, et al. Citrus canker—distribution, taxonomy, epidemiology, disease cycle, pathogen biology, detection, and management: A critical review and future research agenda. Agronomy. 2022; 12: 1075.
- [120] Mashabela MD, Piater LA, Dubery IA, Tugizimana F, Mhlongo MI. Rhizosphere Tripartite Interactions and PGPR-Mediated Metabolic Reprogramming towards ISR and Plant Priming: A Metabolomics Review. Biology. 2022; 11: 346.
- [121] Bukhat S, Imran A, Javaid S, Shahid M, Majeed A, Naqqash T. Communication of plants with microbial world: Exploring the regulatory networks for PGPR mediated defense signaling. Microbiological Research. 2020; 238: 126486.
- [122] Salwan R, Sharma M, Sharma A, Sharma V. Insights into plant beneficial microorganism-triggered induced systemic resistance. Plant Stress. 2023; 7: 100140.
- [123] Mhlongo MI, Piater LA, Madala NE, Labuschagne N, Dubery IA. The Chemistry of Plant-Microbe Interactions in the Rhizosphere and the Potential for Metabolomics to Reveal Signaling Related to Defense Priming and Induced Systemic Resistance. Frontiers in Plant Science. 2018; 9: 112.
- [124] Meena M, Swapnil P, Divyanshu K, Kumar S, Harish, Tripathi YN, et al. PGPR-mediated induction of systemic resistance and physiochemical alterations in plants against the pathogens: Current perspectives. Journal of Basic Microbiology. 2020; 60: 828–861.
- [125] Raymaekers K, Ponet L, Holtappels D, Berckmans B, Cammue BP. Screening for novel biocontrol agents applicable in plant disease management—a review. Biological Control. 2020; 144: 104240.
- [126] Guzmán-Guzmán P, Kumar A, de Los Santos-Villalobos S, Parra-Cota FI, Orozco-Mosqueda MDC, Fadiji AE, et al. Trichoderma Species: Our Best Fungal Allies in the Biocontrol of Plant Diseases-A Review. Plants. 2023; 12: 432.
- [127] Sivasakthi S, Usharani G, Saranraj P. Biocontrol potentiality of plant growth promoting bacteria (PGPR)-Pseudomonas fluorescens and Bacillus subtilis: A review. African Journal of Agricultural Research. 2014; 9: 1265–1277.
- [128] Kour D, Rana KL, Yadav N, Yadav AN, Kumar A, Meena VS, et al. Rhizospheric microbiomes: biodiversity, mechanisms of plant growth promotion, and biotechnological applications for sustainable agriculture. In Kumar A, Meena VS (eds.) Plant growth promoting rhizobacteria for agricultural sustainability: from theory to practices (pp. 19–65). Springer: Singapore 2019.
- [129] Annapurna K, Kumar A, Kumar LV, Govindasamy V, Bose P, Ramadoss D. PGPR-induced systemic resistance (ISR) in plant disease management. In Maheshwari DK (ed.) Bacteria in agrobiology: disease management (pp. 405–425). Springer: Berlin, Heidelberg 2013.
- [130] Vaghela N, Gohel S. Medicinal plant-associated rhizobacteria enhance the production of pharmaceutically important bioactive compounds under abiotic stress conditions. Journal of Basic Microbiology. 2023; 63: 308–325.
- [131] Díaz-Urbano M, Goicoechea N, Velasco P, Poveda J. Development of agricultural bio-inoculants based on mycorrhizal fungi and endophytic filamentous fungi: co-inoculants for improve

- plant-physiological responses in sustainable agriculture. Biological Control. 2023; 182: 105223.
- [132] Sellappan R, Thangavel K, Uthandi S. Bioprotective potential of maize apoplastic fluid bacterium (Bacillus amyloliquefaciens) and arbuscular mycorrhizal fungi (Glomus intraradices) against Spodoptera frugiperda infestation in maize. Physiological and Molecular Plant Pathology. 2023; 127: 102050.
- [133] Martínez-Medina A, Pascual JA, Pérez-Alfocea F, Albacete A, Roldán A. Trichoderma harzianum and Glomus intraradices modify the hormone disruption induced by Fusarium oxysporum infection in melon plants. Phytopathology. 2010; 100: 682–688.
- [134] Bani M, Pérez-De-Luque A, Rubiales D, Rispail N. Physical and Chemical Barriers in Root Tissues Contribute to Quantitative Resistance to *Fusarium oxysporum* f. sp. *pisi* in Pea. Frontiers in Plant Science. 2018; 9: 199.
- [135] Herzog K, Schwander F, Kassemeyer HH, Bieler E, Dürrenberger M, Trapp O, et al. Towards Sensor-Based Phenotyping of Physical Barriers of Grapes to Improve Resilience to Botrytis Bunch Rot. Frontiers in Plant Science. 2022; 12: 808365.
- [136] Ahmad A, Shafique S, Shafique S. Cytological and physiological basis for tomato varietal resistance against Alternaria alternata. Journal of the Science of Food and Agriculture. 2013; 93: 2315–2322.
- [137] Grimault V, Gélie B, Lemattre M, Prior P, Schmit J. Comparative histology of resistant and susceptible tomato cultivars infected by Pseudomonas solanacearum. Physiological and Molecular Plant Pathology. 1994; 44: 105–123.
- [138] Kagale S, Marimuthu T, Kagale J, Thayumanavan B, Samiyappan R. Induction of systemic resistance in rice by leaf extracts of Zizyphus jujuba and Ipomoea carnea against Rhizoctonia solani. Plant Signaling & Behavior. 2011; 6: 919–923.
- [139] Maserumule M, Rauwane M, Madala NE, Ncube E, Figlan S. Defence-related metabolic changes in wheat (*Triticum aestivum* L.) seedlings in response to infection by *Puccinia graminis* f. sp. *tritici*. Frontiers in Plant Science. 2023; 14: 1166813.
- [140] Zhang X, Huang T, Wang Q, Guo Y, Li L, Liu J, et al. Metabonomic Analysis of Iron Shell Wheat (Triticum aestivum ssp. yunnanense King) Reveals Mechanisms of Resistance against Powdery Mildew. Russian Journal of Plant Physiology. 2023; 70: 89.
- [141] Duan S, Jin J, Gao Y, Jin C, Mu J, Zhen W, et al. Integrated transcriptome and metabolite profiling highlights the role of benzoxazinoids in wheat resistance against Fusarium crown rot. The Crop Journal. 2022; 10: 407–417.
- [142] Farahbakhsh F, Hamzehzarghani H, Massah A, Tortosa M, Yassaie M, Rodriguez VM. Comparative metabolomics of temperature sensitive resistance to wheat streak mosaic virus (WSMV) in resistant and susceptible wheat cultivars. Journal of Plant Physiology. 2019; 237: 30–42.
- [143] Yogendra KN, Dhokane D, Kushalappa AC, Sarmiento F, Rodriguez E, Mosquera T. StWRKY8 transcription factor regulates benzylisoquinoline alkaloid pathway in potato conferring resistance to late blight. Plant Science. 2017; 256: 208–216.
- [144] Shinde BA, Dholakia BB, Hussain K, Panda S, Meir S, Rogachev I, *et al.* Dynamic metabolic reprogramming of steroidal glycol-alkaloid and phenylpropanoid biosynthesis may impart early blight resistance in wild tomato (Solanum arcanum Peralta). Plant Molecular Biology. 2017; 95: 411–423.
- [145] Ellendorff U, Fradin EF, de Jonge R, Thomma BPHJ. RNA silencing is required for Arabidopsis defence against Verticillium wilt disease. Journal of Experimental Botany. 2009; 60: 591– 602
- [146] Werner BT, Gaffar FY, Schuemann J, Biedenkopf D, Koch AM. RNA-Spray-Mediated Silencing of Fusarium graminearum AGO and DCL Genes Improve Barley Disease Resistance. Frontiers in Plant Science. 2020; 11: 476.



- [147] Yi H, Richards EJ. A cluster of disease resistance genes in Arabidopsis is coordinately regulated by transcriptional activation and RNA silencing. The Plant Cell. 2007; 19: 2929–2939.
- [148] Scofield SR, Huang L, Brandt AS, Gill BS. Development of a virus-induced gene-silencing system for hexaploid wheat and its use in functional analysis of the Lr21-mediated leaf rust resistance pathway. Plant Physiology. 2005; 138: 2165–2173.
- [149] Zhu X, Qi T, Yang Q, He F, Tan C, Ma W, et al. Host-Induced Gene Silencing of the MAPKK Gene PsFUZ7 Confers Stable Resistance to Wheat Stripe Rust. Plant Physiology. 2017; 175: 1853–1863.
- [150] Ullah H, Yasmin H, Mumtaz S, Jabeen Z, Naz R, Nosheen A, et al. Multitrait Pseudomonas spp. Isolated from Monocropped Wheat (*Triticum aestivum*) Suppress Fusarium Root and Crown Rot. Phytopathology. 2020; 110: 582–592.
- [151] Sari E, Etebarian HR, Aminian H. Effects of Pseudomonas fluorescens CHA0 on the resistance of wheat seedling roots to the take-all fungus Gaeumannomyces graminis var. tritici. Plant Production Science. 2008; 11: 298–306.
- [152] Behiry S, Soliman SA, Massoud MA, Abdelbary M, Kordy AM, Abdelkhalek A, et al. Trichoderma pubescens Elicit Induced Systemic Resistance in Tomato Challenged by Rhizoctonia solani. Journal of Fungi. 2023; 9: 167.
- [153] Abdelkhalek A, Al-Askar AA, Arishi AA, Behiry SI. Trichoderma hamatum Strain Th23 Promotes Tomato Growth and Induces Systemic Resistance against Tobacco Mosaic Virus. Journal of Fungi. 2022; 8: 228.
- [154] Sawant IS, Wadkar PN, Ghule SB, Salunkhe VP, Chavan V, Sawant SD. Induction of systemic resistance in grapevines against powdery mildew by Trichoderma asperelloides strains. Australasian Plant Pathology. 2020; 49: 107–117.
- [155] Sawant SB, Prabhukarthikeyan S, Mishra MK, Parameswaran C, Keerthana U, Senapati AK. Induction of defense-related enzymes and enhanced disease resistance in rice against Sarocladium oryzae by Bacillus cereus RBS-57. Physiological and Molecular Plant Pathology. 2023; 128: 102168.
- [156] Cordier C, Pozo MJ, Barea J-M, Gianinazzi S, Gianinazzi-Pearson V. Cell defense responses associated with localized and systemic resistance to Phytophthora parasitica induced in tomato by an arbuscular mycorrhizal fungus. Molecular Plant-Microbe Interactions. 1998; 11: 1017–1028.
- [157] Fravel D, Olivain C, Alabouvette C. Fusarium oxysporum and its biocontrol. The New Phytologist. 2003; 157: 493–502.
- [158] Weng W, Yan J, Zhou M, Yao X, Gao A, Ma C, et al. Roles of Arbuscular mycorrhizal Fungi as a Biocontrol Agent in the Control of Plant Diseases. Microorganisms. 2022; 10: 1266.
- [159] Sarrocco S, Mauro A, Battilani P. Use of Competitive Filamentous Fungi as an Alternative Approach for Mycotoxin Risk Reduction in Staple Cereals: State of Art and Future Perspectives. Toxins. 2019; 11: 701.
- [160] Hodge A, Fitter AH. Microbial mediation of plant competition and community structure. Functional Ecology. 2013; 27: 865– 875.
- [161] Haggag WM, Mohamed H. Biotechnological aspects of microorganisms used in plant biological control. American-Eurasian Journal of Sustainable Agriculture. 2007; 1: 7–12.
- [162] Arguelles-Arias A, Ongena M, Halimi B, Lara Y, Brans A, Joris B, et al. Bacillus amyloliquefaciens GA1 as a source of potent antibiotics and other secondary metabolites for biocontrol of plant pathogens. Microbial Cell Factories. 2009; 8: 63.
- [163] Etesami H, Jeong BR, Glick BR. Biocontrol of plant diseases by Bacillus spp. Physiological and Molecular Plant Pathology. 2023; 126: 102048.
- [164] Amaning Danquah C, Minkah PAB, Osei Duah Junior I, Amankwah KB, Somuah SO. Antimicrobial Compounds from Microorganisms. Antibiotics. 2022; 11: 285.

- [165] Poveda J, Abril-Urias P, Escobar C. Biological Control of Plant-Parasitic Nematodes by Filamentous Fungi Inducers of Resistance: *Trichoderma*, Mycorrhizal and Endophytic Fungi. Frontiers in Microbiology. 2020; 11: 992.
- [166] Sugiyama A, Sano CM, Yazaki K, Sano H. Caffeine fostering of mycoparasitic fungi against phytopathogens. Plant Signaling & Behavior. 2016; 11: e1113362.
- [167] Sinkala N. Efficacy of mycoparasitic and entomopathogenic fungi against fungal agents of plant diseases [Bachelor's thesis]. University of South Bohemia České Budějovice. 2023. (In Czech)
- [168] Jamal M, Bukhari SMAUS, Andleeb S, Ali M, Raza S, Nawaz MA, et al. Bacteriophages: an overview of the control strategies against multiple bacterial infections in different fields. Journal of Basic Microbiology. 2019; 59: 123–133.
- [169] Born Y, Fieseler L, Thöny V, Leimer N, Duffy B, Loessner MJ. Engineering of Bacteriophages Y2::dpoL1-C and Y2::luxAB for Efficient Control and Rapid Detection of the Fire Blight Pathogen, Erwinia amylovora. Applied and Environmental Microbiology. 2017; 83: e00341-17.
- [170] Yamada T, Kawasaki T, Nagata S, Fujiwara A, Usami S, Fujie M. New bacteriophages that infect the phytopathogen Ralstonia solanacearum. Microbiology. 2007; 153: 2630–2639.
- [171] Coyne DL, Cortada L, Dalzell JJ, Claudius-Cole AO, Haukeland S, Luambano N, et al. Plant-Parasitic Nematodes and Food Security in Sub-Saharan Africa. Annual Review of Phytopathology. 2018; 56: 381–403.
- [172] Saikai KK, Oduori C, Situma E, Njoroge S, Murunde R, Kimenju JW, et al. Biocontrol-based strategies for improving soil health and managing plant-parasitic nematodes in coffee production. Frontiers in Plant Science. 2023; 14: 1196171.
- [173] Sharma N, Khanna K, Jasrotia S, Kumar D, Bhardwaj R, Ohri P. Metabolites and chemical agents in the plant roots: An overview of their use in plant-parasitic nematode management. Nematology. 2023; 25: 243–257.
- [174] Pliego C, Ramos C, de Vicente A, Cazorla FM. Screening for candidate bacterial biocontrol agents against soilborne fungal plant pathogens. Plant and Soil. 2011; 340: 505–520.
- [175] Xue QY, Chen Y, Li SM, Chen LF, Ding GC, Guo DW, et al. Evaluation of the strains of Acinetobacter and Enterobacter as potential biocontrol agents against Ralstonia wilt of tomato. Biological Control. 2009; 48: 252–258.
- [176] Lagzian A, Saberi Riseh R, Khodaygan P, Sedaghati E, Dashti H. Introduced Pseudomonas fluorescens VUPf5 as an important biocontrol agent for controlling Gaeumannomyces graminis var. tritici the causal agent of take-all disease in wheat. Archives of Phytopathology and Plant Protection. 2013; 46: 2104–2116.
- [177] Wang X, Mavrodi DV, Ke L, Mavrodi OV, Yang M, Thomashow LS, *et al.* Biocontrol and plant growth-promoting activity of rhizobacteria from Chinese fields with contaminated soils. Microbial Biotechnology. 2015; 8: 404–418.
- [178] Ji P, Campbell H, Kloepper J, Jones J, Suslow T, Wilson M. Integrated biological control of bacterial speck and spot of tomato under field conditions using foliar biological control agents and plant growth-promoting rhizobacteria. Biological Control. 2006; 36: 358–367.
- [179] Egamberdieva D, Davranov K, Wirth S, Hashem A, Abd Allah EF. Impact of soil salinity on the plant-growth promoting and biological control abilities of root associated bacteria. Saudi Journal of Biological Sciences. 2017; 24: 1601–1608.
- [180] Domenech J, Reddy M, Kloepper J, Ramos B, Gutierrez-Mañero J. Combined application of the biological product LS213 with Bacillus, Pseudomonas or Chryseobacterium for growth promotion and biological control of soil-borne diseases in pepper and tomato. BioControl. 2006; 51: 245–258.
- [181] Sun G, Yao T, Feng C, Chen L, Li J, Wang L. Identification



- and biocontrol potential of antagonistic bacteria strains against Sclerotinia sclerotiorum and their growth-promoting effects on Brassica napus. Biological Control. 2017; 104: 35–43.
- [182] Lacey LA, Grzywacz D, Shapiro-Ilan DI, Frutos R, Brown-bridge M, Goettel MS. Insect pathogens as biological control agents: Back to the future. Journal of Invertebrate Pathology. 2015; 132: 1–41.
- [183] Kagimu N, Ferreira T, Malan A. The attributes of survival in the formulation of entomopathogenic nematodes utilised as insect biocontrol agents. African Entomology. 2017; 25: 275–291.
- [184] Saldaña-Mendoza SA, Pacios-Michelena S, Palacios-Ponce AS, Chávez-González ML, Aguilar CN. Trichoderma as a biological control agent: mechanisms of action, benefits for crops and development of formulations. World Journal of Microbiology & Biotechnology. 2023; 39: 269.
- [185] Sharma P. Biocontrol strategies—retrospect and prospects. Indian Phytopathology. 2023; 76: 47–59.
- [186] Zottele M, Mayrhofer M, Embleton H, Enkerli J, Eigner H, Tarasco E, et al. Integrated Biological Control of the Sugar Beet Weevil Asproparthenis punctiventris with the Fungus Metarhizium brunneum: New Application Approaches. Pathogens. 2023; 12: 99.
- [187] Limdolthamand S, Songkumarn P, Suwannarat S, Jantasorn A, Dethoup T. Biocontrol efficacy of endophytic Trichoderma spp. in fresh and dry powder formulations in controlling northern corn leaf blight in sweet corn. Biological Control. 2023; 181: 105217.
- [188] Rathore G, Kaushal R, Sharma V, Lalkhumliana F. Quantifying the effect of fermented liquid bio formulations and organic amendments on yield and quality of eggplant (Solanum melongena L.). Journal of Plant Nutrition. 2023; 46: 2276–2288.
- [189] Behle R, Birthisel T. Formulations of entomopathogens as bioinsecticides. In Morales-Ramos JA, Rojas MG, Shapiro-Ilan DI (eds.) Mass production of beneficial organisms (pp. 407– 429). 2nd edn. Elsevier: Amsterdam, The Netherlands. 2023.
- [190] Mitra D, De Los Santos-Villalobos S, Parra-Cota FI, Montelongo AMG, Blanco EL, Olatunbosun AN, *et al.* Rice (Oryza sativa L.) plant protection using dual biological control and plant growth-promoting agents: Current scenarios and future prospects. Pedosphere. 2023; 33: 268–286.
- [191] Saberi Riseh R, Vatankhah M, Hassanisaadi M, Kennedy JF. Chitosan/silica: A hybrid formulation to mitigate phytopathogens. International Journal of Biological Macromolecules. 2023; 239: 124192.
- [192] Saberi Riseh R, Hassanisaadi M, Vatankhah M, Kennedy JF. Encapsulating biocontrol bacteria with starch as a safe and edible biopolymer to alleviate plant diseases: A review. Carbohydrate Polymers. 2023; 302: 120384.
- [193] Saberi Riseh R, Moradi Pour M, Ait Barka E. A Novel route for double-layered encapsulation of Streptomyces fulvissimus Uts22 by alginate—Arabic gum for controlling of Pythium aphanidermatum in Cucumber. Agronomy. 2022; 12: 655.
- [194] Saberi Riseh R, Gholizadeh Vazvani M, Hassanisaadi M, Thakur VK, Kennedy JF. Use of whey protein as a natural polymer for the encapsulation of plant biocontrol bacteria: A review. International Journal of Biological Macromolecules. 2023; 234: 123708.
- [195] Moradi Pour M, Hassanisaadi M, Kennedy JF, Saberi Riseh R. A novel biopolymer technique for encapsulation of Bacillus velezensis BV9 into double coating biopolymer made by in alginate and natural gums to biocontrol of wheat take-all disease. International Journal of Biological Macromolecules. 2024; 257: 128526.
- [196] Hassanisaadi M, Kennedy JF, Rabiei A, Riseh RS, Taheri A. Nature's coatings: Sodium alginate as a novel coating in safeguarding plants from frost damages. International Journal of Bi-

- ological Macromolecules. 2024; 267: 131203.
- [197] Vejan P, Khadiran T, Abdullah R, Ismail S, Dadrasnia A. Encapsulation of plant growth promoting Rhizobacteria—prospects and potential in agricultural sector: a review. Journal of Plant Nutrition. 2019; 42: 2600–2623.
- [198] Riseh RS, Tamanadar E, Pour MM, Thakur VK. Novel approaches for encapsulation of plant probiotic bacteria with sustainable polymer gums: application in the management of pests and diseases. Advances in Polymer Technology. 2022; 2022: 1–10
- [199] Riseh RS, Tamanadar E, Hajabdollahi N, Vatankhah M, Thakur VK, Skorik YA. Chitosan microencapsulation of rhizobacteria for biological control of plant pests and diseases: Recent advances and applications. Rhizosphere. 2022; 23: 100565.
- [200] Soukoulis C, Bohn T. A comprehensive overview on the microand nano-technological encapsulation advances for enhancing the chemical stability and bioavailability of carotenoids. Critical Reviews in Food Science and Nutrition. 2018; 58: 1–36.
- [201] Saberi Riseh R, Skorik YA, Thakur VK, Moradi Pour M, Tamanadar E, Noghabi SS. Encapsulation of Plant Biocontrol Bacteria with Alginate as a Main Polymer Material. International Journal of Molecular Sciences. 2021; 22: 11165.
- [202] Saberi Riseh R, Hassanisaadi M, Vatankhah M, Soroush F, Varma RS. Nano/microencapsulation of plant biocontrol agents by chitosan, alginate, and other important biopolymers as a novel strategy for alleviating plant biotic stresses. International Journal of Biological Macromolecules. 2022; 222: 1589–1604.
- [203] Saberi-Riseh R, Moradi-Pour M, Mohammadinejad R, Thakur VK. Biopolymers for Biological Control of Plant Pathogens: Advances in Microencapsulation of Beneficial Microorganisms. Polymers. 2021; 13: 1938.
- [204] Saberi Riseh R, Gholizadeh Vazvani M, Hassanisaadi M, Skorik YA. Micro-/Nano-Carboxymethyl Cellulose as a Promising Biopolymer with Prospects in the Agriculture Sector: A Review. Polymers. 2023; 15: 440.
- [205] Saberi Riseh R, Hassanisaadi M, Vatankhah M, Varma RS, Thakur VK. Nano/Micro-Structural Supramolecular Biopolymers: Innovative Networks with the Boundless Potential in Sustainable Agriculture. Nano-Micro Letters. 2024; 16: 147.
- [206] Saberi Riseh R, Vatankhah M, Hassanisaadi M, Varma RS. A review of chitosan nanoparticles: Nature's gift for transforming agriculture through smart and effective delivery mechanisms. International Journal of Biological Macromolecules. 2024; 260: 129522.
- [207] Saberi Riseh R, Vatankhah M, Hassanisaadi M, Shafiei-Hematabad Z, Kennedy JF. Advancements in coating technologies: Unveiling the potential of chitosan for the preservation of fruits and vegetables. International Journal of Biological Macromolecules. 2024; 254: 127677.
- [208] Fathi F, Saberi-Riseh R, Khodaygan P. Survivability and controlled release of alginate-microencapsulated Pseudomonas fluorescens VUPF506 and their effects on biocontrol of Rhizoctonia solani on potato. International Journal of Biological Macromolecules. 2021; 183: 627–634.
- [209] Moradi Pour M, Saberi Riseh R, Skorik YA. Sodium Alginate-Gelatin Nanoformulations for Encapsulation of *Bacillus velezensis* and Their Use for Biological Control of Pistachio Gummosis. Materials. 2022; 15: 2114.
- [210] Fathi F, Saberi Riseh R, Khodaygan P, Hosseini S, Skorik YA. Microencapsulation of a *Pseudomonas* Strain (VUPF506) in Alginate-Whey Protein-Carbon Nanotubes and Next-Generation Sequencing Identification of This Strain. Polymers. 2021; 13: 4269.
- [211] Szczech M, Maciorowski R. Microencapsulation technique with organic additives for biocontrol agents. Journal of Horticultural Research. 2016; 24: 111–122.



- [212] Saeed Ben Youssef A. Modern trends for the application of biological control and modern technologies in agricultural projects. International Journal of Modern Agriculture and Environment. 2023; 1: 26-53.
- [213] Sare AR, Jijakli MH, Massart S. Microbial ecology to support integrative efficacy improvement of biocontrol agents for postharvest diseases management. Postharvest Biology and Technology. 2021; 179: 111572.
- [214] Jambhulkar PP, Sharma P, Yadav R. Delivery systems for introduction of microbial inoculants in the field. In Singh DP, Singh HB, Prabha R (eds.) Microbial Inoculants in Sustainable Agricultural Productivity: Vol. 2: Functional Applications (pp. 199–218). Springer: New Delhi. 2016.
- [215] Elad Y, Freeman S. Biological control of fungal plant pathogens. In Kempken F (ed.) Agricultural applications (pp. 93–109). Springer: Berlin, Heidelberg. 2002.
- [216] Maheshwari DK. Plant growth and health promoting bacteria. Springer Science & Business Media: Berlin, Germany, 2010.
- [217] Zohar-Perez C, Chernin L, Chet I, Nussinovitch A. Structure of dried cellular alginate matrix containing fillers provides extra protection for microorganisms against UVC radiation. Radiation Research. 2003; 160: 198–204.
- [218] Berninger T, González López Ó, Bejarano A, Preininger C, Sessitsch A. Maintenance and assessment of cell viability in formulation of non-sporulating bacterial inoculants. Microbial Biotechnology. 2018; 11: 277–301.
- [219] Malusá E, Sas-Paszt L, Ciesielska J. Technologies for beneficial microorganisms inocula used as biofertilizers. The Scientific World Journal. 2012; 2012: 491206.
- [220] Herrmann L, Lesueur D. Challenges of formulation and quality of biofertilizers for successful inoculation. Applied Microbiology and Biotechnology. 2013; 97: 8859–8873.
- [221] Bashan Y, de-Bashan LE, Prabhu S, Hernandez JP. Advances in plant growth-promoting bacterial inoculant technology: formulations and practical perspectives (1998–2013). Plant and Soil. 2014; 378: 1–33.
- [222] O'Callaghan M. Microbial inoculation of seed for improved crop performance: issues and opportunities. Applied Microbiology and Biotechnology. 2016; 100: 5729–5746.
- [223] Kessler P, Matzke H, Keller S. The effect of application time and soil factors on the occurrence of Beauveria brongniartii applied as a biological control agent in soil. Journal of Invertebrate Pathology. 2003; 84: 15–23.
- [224] Bardin M, Nicot PC. Microorganisms as Biocontrol Products. In Fauvergue X, Rusch A, Barret M, Bardin M, Jacquin-Joly E, Malausa T, Lannou C (eds.) Extended Biocontrol (pp. 127–136). Springer: Dordrecht. 2022.
- [225] Senko H, Kajić S, Hud A, Palijan G, Petek M, Rajnović I, et al. Will the beneficial properties of plant-growth promoting bacteria be affected by waterlogging predicted in the wake of climate change: A model study. Applied Soil Ecology. 2024; 198: 105379.
- [226] Idris AL, Li W, Huang F, Lin F, Guan X, Huang T. Impacts of UV radiation on Bacillus biocontrol agents and their resistance mechanisms. World Journal of Microbiology & Biotechnology. 2024; 40: 58.
- [227] Costa E, Usall J, Teixidó N, Delgado J, Viñas I. Water activity, temperature, and pH effects on growth of the biocontrol agent Pantoea agglomerans CPA-2. Canadian Journal of Microbiology. 2002; 48: 1082–1088.
- [228] Fedele G, Brischetto C, Rossi V. Biocontrol of *Botrytis cinerea* on Grape Berries as Influenced by Temperature and Humidity. Frontiers in Plant Science. 2020; 11: 1232.
- [229] Larach A, Vega-Celedón P, Castillo-Novales D, Tapia L, Cuneo I, Cádiz F, et al. Diplodia seriata Biocontrol Is Altered via Temperature and the Control of Bacteria. Microorganisms.

- 2024; 12: 350.
- [230] Gotor-Vila A, Teixidó N, Sisquella M, Torres R, Usall J. Biological Characterization of the Biocontrol Agent Bacillus amyloliquefaciens CPA-8: The Effect of Temperature, pH and Water Activity on Growth, Susceptibility to Antibiotics and Detection of Enterotoxic Genes. Current Microbiology. 2017; 74: 1089–1099.
- [231] Abbas Z, Authman S, Al-Ezee A. Temperature effects on growth of the biocontrol agent Pantoea agglomerans (an oval isolate from Iraqi soils). Journal of Advanced Laboratory Research in Biology. 2017; 8: 85–88.
- [232] Humair B, González N, Mossialos D, Reimmann C, Haas D. Temperature-responsive sensing regulates biocontrol factor expression in Pseudomonas fluorescens CHA0. The ISME Journal. 2009; 3: 955–965.
- [233] Abd El-Mageed TA, Rady MM, Taha RS, Abd El Azeam S, Simpson CR, Semida WM. Effects of integrated use of residual sulfur-enhanced biochar with effective microorganisms on soil properties, plant growth and short-term productivity of Capsicum annuum under salt stress. Scientia Horticulturae. 2020; 261: 108930.
- [234] Choudhary DK, Sharma KP, Gaur RK. Biotechnological perspectives of microbes in agro-ecosystems. Biotechnology Letters. 2011; 33: 1905–1910.
- [235] El-Saadony MT, Saad AM, Soliman SM, Salem HM, Ahmed AI, Mahmood M, et al. Plant growth-promoting microorganisms as biocontrol agents of plant diseases: Mechanisms, challenges and future perspectives. Frontiers in Plant Science. 2022; 13: 923880.
- [236] Fedele G, Bove F, González-Domínguez E, Rossi V. Modelling biocontrol agents as plant protection tools. Biology and Life Sciences Forum, MDPI. 2020; 4: 75.
- [237] Jacobsen BJ, Zidack NK, Larson BJ. The role of bacillus-based biological control agents in integrated pest management systems: plant diseases. Phytopathology. 2004; 94: 1272–1275.
- [238] Bothikar PA, Pagire KS. Integrated Pest Management (IPM)–A new dimension in sustainable agriculture development. Journal of Advanced Education and Sciences. 2022; 2: 7–11.
- [239] Rodríguez D, Coy-Barrera E. Overview of Updated Control Tactics for Western Flower Thrips. Insects. 2023; 14: 649.
- [240] Baker BP, Green TA, Loker AJ. Biological control and integrated pest management in organic and conventional systems. Biological Control. 2020; 140: 104095.
- [241] Hassanisaadi M, Saberi Riseh R, Rabiei A, Varma RS, Kennedy JF. Corrigendum to "Nano/micro-cellulose-based materials as adsorbents for the remediation of chemical pollutants from agricultural resources" [Int. J. Biol. Macromol. 246 (2023) published online/125763]. International Journal of Biological Macromolecules. 2023; 250: 126128.
- [242] Van Driesche RG, Bellows TS, Van Driesche RG, Bellows TS. Natural enemy monitoring and evaluation. Biological Control (pp. 259–295). Springer, Boston, MA. 1996.
- [243] Ahmed S, Ibrahim M, El-Sawi S, Motawe H. Monitoring and evaluation of some Egyptian wild plants grown in the Eastern Desert of Egypt. Journal of Materials and Environmental Sciences. 2018; 9: 1692–1699.
- [244] Syed Ab Rahman SF, Singh E, Pieterse CMJ, Schenk PM. Emerging microbial biocontrol strategies for plant pathogens. Plant Science. 2018; 267: 102–111.
- [245] Heydari A, Pessarakli M. A review on biological control of fungal plant pathogens using microbial antagonists. Journal of Biological Sciences. 2010; 10: 273–290.
- [246] Smith KP, Handelsman J, Goodman RM. Genetic basis in plants for interactions with disease-suppressive bacteria. Proceedings of the National Academy of Sciences of the United States of America. 1999; 96: 4786–4790.



- [247] Pearson DE, Callaway RM. Indirect effects of host-specific biological control agents. Trends in Ecology & Evolution. 2003; 18: 456–461.
- [248] Salgotra RK, Chauhan BS. Genetic Diversity, Conservation, and Utilization of Plant Genetic Resources. Genes. 2023; 14: 174
- [249] Bharadwaj DN. Sustainable agriculture and plant breeding. In Al-Khayri JM, Jain SM, Johnson DV (eds.) Advances in plant breeding strategies: agronomic, abiotic and biotic stress traits (pp. 3–34). Springer: Cham. 2016.
- [250] Wang X, Zhao D, Shen L, Jing C, Zhang C. Application and mechanisms of Bacillus subtilis in biological control of plant disease. In Meena VS (ed.) Role of rhizospheric microbes in soil: Volume 1: Stress management and agricultural sustainability (pp. 225–250). Springer: Singapore. 2018.
- [251] Nguvo KJ, Gao X. Weapons hidden underneath: bio-control agents and their potentials to activate plant induced systemic resistance in controlling crop Fusarium diseases. Journal of Plant Diseases and Protection. 2019; 126: 177–190.
- [252] Lagzian A, Riseh RS, Sarikhan S, Ghorbani A, Khodaygan P, Borriss R, et al. Genome mining conformance to metabolite profile of Bacillus strains to control potato pathogens. Scientific Reports. 2023; 13: 19095.
- [253] Kumar A, Verma JP. The role of microbes to improve crop productivity and soil health. In Achal V, Mukherjee A (eds.) Ecological wisdom inspired restoration engineering (pp. 249–265). Springer: Singapore. 2019.
- [254] Li J, Wang J, Liu H, Macdonald CA, Singh BK. Application of microbial inoculants significantly enhances crop productivity: a meta-analysis of studies from 2010 to 2020. Journal of Sustainable Agriculture and Environment. 2022; 1: 216–225.
- [255] Mekonnen H, Kibret M. The roles of plant growth promoting rhizobacteria in sustainable vegetable production in Ethiopia. Chemical and Biological Technologies in Agriculture. 2021; 8: 1–11
- [256] Baar J. From production to application of arbuscular mycorrhizal fungi in agricultural systems: requirements and needs. In Varma A (ed.) Mycorrhiza: state of the art, genetics and molecular biology, eco-function, biotechnology, eco-physiology, structure and systematics (pp. 361–373). Springer: Berlin, Heidelberg. 2008.
- [257] Ehrmann J, Ritz K. Plant: soil interactions in temperate multicropping production systems. Plant and Soil. 2014; 376: 1–29.
- [258] Yang H, Zhang W, Li L. Intercropping: Feed more people and build more sustainable agroecosystems. Frontiers of Agricultural Science and Engineering. 2021; 8: 373–386.
- [259] Litsinger J, Moody K. Integrated pest management in multiple cropping systems. Multiple Cropping. 1976; 27: 293–316.
- [260] Mashabela MD, Tugizimana F, Steenkamp PA, Piater LA, Dubery IA, Terefe T, et al. Metabolomic evaluation of PGPR defence priming in wheat (*Triticum aestivum* L.) cultivars infected with *Puccinia striiformis* f. sp. tritici (stripe rust). Frontiers in Plant Science. 2023; 14: 1103413.
- [261] García-Montelongo AM, Montoya-Martínez AC, Morales-Sandoval PH, Parra-Cota FI, de los Santos-Villalobos S. Beneficial Microorganisms as a Sustainable Alternative for Mitigating Biotic Stresses in Crops. Stresses. 2023; 3: 210–228.
- [262] Chakraborti S, Bera K, Sadhukhan S, Dutta P. Bio-priming of seeds: Plant stress management and its underlying cellular, biochemical and molecular mechanisms. Plant Stress. 2022; 3: 100052.
- [263] Dimkić I, Janakiev T, Petrović M, Degrassi G, Fira D. Plantassociated Bacillus and Pseudomonas antimicrobial activities in plant disease suppression via biological control mechanisms-A review. Physiological and Molecular Plant Pathology. 2022; 117: 101754.

- [264] Manzar N, Kashyap AS, Goutam RS, Rajawat MVS, Sharma PK, Sharma SK, et al. Trichoderma: Advent of versatile biocontrol agent, its secrets and insights into mechanism of biocontrol potential. Sustainability. 2022; 14: 12786.
- [265] Kenawy A, Dailin DJ, Abo-Zaid GA, Malek RA, Ambehabati KK, Zakaria KHN, et al. Biosynthesis of antibiotics by PGPR and their roles in biocontrol of plant diseases. In Sayyed RZ (ed.) Plant Growth Promoting Rhizobacteria for Sustainable Stress Management: Volume 2: Rhizobacteria in Biotic Stress Management (pp. 1–35). Springer: Singapore. 2019.
- [266] Kaur S, Samota MK, Choudhary M, Choudhary M, Pandey AK, Sharma A, et al. How do plants defend themselves against pathogens-Biochemical mechanisms and genetic interventions. Physiology and Molecular Biology of Plants. 2022; 28: 485–504.
- [267] Yuan M, Huang Y, Ge W, Jia Z, Song S, Zhang L, et al. Involvement of jasmonic acid, ethylene and salicylic acid signaling pathways behind the systemic resistance induced by Trichoderma longibrachiatum H9 in cucumber. BMC Genomics. 2019; 20: 144.
- [268] Boamah S, Ojangba T, Zhang S, Zhu N, Osei R, John Tiika R, et al. Evaluation of salicylic acid (SA) signaling pathways and molecular markers in Trichoderma-treated plants under salinity and Fusarium stresses. A Review. European Journal of Plant Pathology. 2023; 166: 259–274.
- [269] Abid SA, Karim SH. Salicylic Acid, Jasmonic Acid and Ethylene Involved in The Resistance Induced By The Rhizobacterium Pta-Ct2 In Arabidopsis thaliana Against Botrytis cinerea. Journal of Plant Production. 2021; 12: 361–364.
- [270] Rajput NA, Zhang M, Ru Y, Liu T, Xu J, Liu L, et al. Phytophthora sojae effector PsCRN70 suppresses plant defenses in Nicotiana benthamiana. PLoS ONE. 2014; 9: e98114.
- [271] Buonaurio R. Infection and plant defense responses during plant-bacterial interaction. Plant-Microbe interactions. 2008; 169–197.
- [272] Varshney RK, Pandey MK, Chitikineni A. Plant Genetics and Molecular Biology: An Introduction. Advances in Biochemical Engineering/Biotechnology. 2018; 164: 1–9.
- [273] Rowland BE, Henriquez MA, Nilsen KT, Subramaniam R, Walkowiak S. Unraveling Plant-Pathogen Interactions in Cereals Using RNA-seq. Methods in Molecular Biology. 2023; 2659: 103–118.
- [274] Van Borm S, Belák S, Freimanis G, Fusaro A, Granberg F, Höper D, et al. Next-generation sequencing in veterinary medicine: how can the massive amount of information arising from high-throughput technologies improve diagnosis, control, and management of infectious diseases? Methods in Molecular Biology. 2015; 1247: 415–436.
- [275] Hu B, Xie G, Lo CC, Starkenburg SR, Chain PSG. Pathogen comparative genomics in the next-generation sequencing era: genome alignments, pangenomics and metagenomics. Briefings in Functional Genomics. 2011; 10: 322–333.
- [276] Winkler KR, Mizrahi V, Warner DF, De Wet TJ. Highthroughput functional genomics: A (myco)bacterial perspective. Molecular Microbiology. 2023; 120: 141–158.
- [277] Dracatos PM, Lu J, Sánchez-Martín J, Wulff BBH. Resistance that stacks up: engineering rust and mildew disease control in the cereal crops wheat and barley. Plant Biotechnology Journal. 2023; 21: 1938–1951.
- [278] Larena I, Espeso EA, Veloso J. Editorial: Impact of novel omic technologies on biological control against plant pathogens. Frontiers in Microbiology. 2023; 14: 1162422.
- [279] Tuo D, Yao Y, Yan P, Chen X, Qu F, Xue W, et al. Development of cassava common mosaic virus-based vector for protein expression and gene editing in cassava. Plant Methods. 2023; 19: 78.



- [280] Rani K, Kumar K, Navpute G, Panwar A, Kaur A, Kaushik P. Microbiome Interactions Affect Plant Physiology and Immunity to Environmental Stresses. 2023. (preprint)
- [281] Patil R, Satpute R, Nalage D. Plant microbiomes and their role in plant health. Microenvironment & Microecology Research. 2023: 5: 2.
- [282] Yu Y, Gui Y, Li Z, Jiang C, Guo J, Niu D. Induced Systemic Resistance for Improving Plant Immunity by Beneficial Microbes. Plants. 2022; 11: 386.
- [283] Hashem A, Tabassum B, Fathi Abd Allah E. Bacillus subtilis: A plant-growth promoting rhizobacterium that also impacts biotic stress. Saudi Journal of Biological Sciences. 2019; 26: 1291–1297.
- [284] Kim J, Felton GW. Priming of antiherbivore defensive responses in plants. Insect Science. 2013; 20: 273–285.
- [285] Martinez-Medina A, Flors V, Heil M, Mauch-Mani B, Pieterse CMJ, Pozo MJ, *et al.* Recognizing Plant Defense Priming. Trends in Plant Science. 2016; 21: 818–822.
- [286] Hönig M, Roeber VM, Schmülling T, Cortleven A. Chemical priming of plant defense responses to pathogen attacks. Frontiers in Plant Science. 2023; 14: 1146577.
- [287] Dubey SK, Mansoori A, Mohan M, Sharma K, Kumar A. Hedychium coronarium flower extract modulates rice plant photosystem II and antioxidant enzyme activity to induce resistance against bacterial blight. Physiological and Molecular Plant Pathology. 2023; 127: 102114.
- [288] Yuan T, Ren W, Wang Z, Fry EL, Tang S, Yin J, *et al.* How does the pattern of root metabolites regulating beneficial microorganisms change with different grazing pressures? Frontiers in Plant Science. 2023; 14: 1180576.
- [289] Molinari S, Leonetti P. Bio-control agents activate plant immune response and prime susceptible tomato against root-knot nematodes. PLoS ONE. 2019; 14: e0213230.
- [290] Russo A, Pollastri S, Ruocco M, Monti MM, Loreto F. Volatile organic compounds in the interaction between plants and beneficial microorganisms. Journal of Plant Interactions. 2022; 17: 840–852.
- [291] Jung SC, Martinez-Medina A, Lopez-Raez JA, Pozo MJ. Mycorrhiza-induced resistance and priming of plant defenses. Journal of Chemical Ecology. 2012; 38: 651–664.
- [292] Kurth F, Mailänder S, Bönn M, Feldhahn L, Herrmann S, Große I, et al. Streptomyces-induced resistance against oak powdery mildew involves host plant responses in defense, photosynthesis, and secondary metabolism pathways. Molecular Plant-Microbe Interactions. 2014; 27: 891–900.
- [293] Llorens E, González-Hernández AI, Scalschi L, Fernández-Crespo E, Camañes G, Vicedo B, *et al.* Priming mediated stress and cross-stress tolerance in plants: Concepts and opportunities. Priming-mediated stress and cross-stress tolerance in crop plants (pp. 1–20). Elsevier: Amsterdam, The Netherlands. 2020.
- [294] Margaritopoulou T, Toufexi E, Kizis D, Balayiannis G, Anagnostopoulos C, Theocharis A, et al. Reynoutria sachalinensis extract elicits SA-dependent defense responses in courgette genotypes against powdery mildew caused by Podosphaera xanthii. Scientific Reports. 2020; 10: 3354.
- [295] Ramkissoon A, Ramsubhag A, Jayaraman J. Phytoelicitor activity of three Caribbean seaweed species on suppression of pathogenic infections in tomato plants. Journal of Applied Phycology. 2017; 29: 3235–3244.
- [296] Agarwal PK, Dangariya M, Agarwal P. Seaweed extracts: Potential biodegradable, environmentally friendly resources for regulating plant defence. Algal Research. 2021; 58: 102363.
- [297] Buswell W, Schwarzenbacher RE, Luna E, Sellwood M, Chen B, Flors V, *et al.* Chemical priming of immunity without costs to plant growth. The New Phytologist. 2018; 218: 1205–1216.
- [298] Thevenet D, Pastor V, Baccelli I, Balmer A, Vallat A, Neier

- R, *et al*. The priming molecule β -aminobutyric acid is naturally present in plants and is induced by stress. The New Phytologist. 2017; 213: 552–559.
- [299] Luna E, van Hulten M, Zhang Y, Berkowitz O, López A, Pétriacq P, et al. Plant perception of β-aminobutyric acid is mediated by an aspartyl-tRNA synthetase. Nature Chemical Biology. 2014; 10: 450–456.
- [300] Virág E, Kiniczky M, Kutasy B, Nagy Á, Pallos JP, Laczkó L, et al. Supplementation of the Plant Conditioner ELICE Vakcina[®] Product with β-Aminobutyric Acid and Salicylic Acid May Lead to *Trans*-Priming Signaling in Barley (*Hordeum vulgare*). Plants. 2023; 12: 2308.
- [301] Döring AC. Exploring the role of pipecolic acid in defence priming and inducible immunity in Arabidopsis thaliana (L.) Heynh [Doctoral of thesis]. Düsseldorf, Heinrich-Heine-Universität, Diss. 2015.
- [302] Balmer A, Pastor V, Glauser G, Mauch-Mani B. Tricarboxylates Induce Defense Priming Against Bacteria in *Arabidopsis* thaliana. Frontiers in Plant Science. 2018; 9: 1221.
- [303] Yi HS, Yang JW, Ryu CM. ISR meets SAR outside: additive action of the endophyte Bacillus pumilus INR7 and the chemical inducer, benzothiadiazole, on induced resistance against bacterial spot in field-grown pepper. Frontiers in Plant Science. 2013; 4: 122.
- [304] Vos IA, Pieterse CM, Van Wees SC. Costs and benefits of hormone-regulated plant defences. Plant Pathology. 2013; 62: 43– 55.
- [305] Vidhyasekaran P. Plant hormone signaling systems in plant innate immunity. Springer: Dordrecht. 2015.
- [306] Martínez-Medina A, Fernández I, Sánchez-Guzmán MJ, Jung SC, Pascual JA, Pozo MJ. Deciphering the hormonal signalling network behind the systemic resistance induced by Trichoderma harzianum in tomato. Frontiers in Plant Science. 2013; 4: 206.
- [307] Bari R, Jones JDG. Role of plant hormones in plant defence responses. Plant Molecular Biology. 2009; 69: 473–488.
- [308] de Torres Zabala M, Bennett MH, Truman WH, Grant MR. Antagonism between salicylic and abscisic acid reflects early host-pathogen conflict and moulds plant defence responses. The Plant Journal: for Cell and Molecular Biology. 2009; 59: 375–386.
- [309] Robert-Seilaniantz A, Navarro L, Bari R, Jones JDG. Pathological hormone imbalances. Current Opinion in Plant Biology. 2007; 10: 372–379.
- [310] Benjamin G, Pandharikar G, Frendo P. Salicylic Acid in Plant Symbioses: Beyond Plant Pathogen Interactions. Biology. 2022; 11: 861.
- [311] Lv ZY, Sun WJ, Jiang R, Chen JF, Ying X, Zhang L, et al. Phytohormones jasmonic acid, salicylic acid, gibberellins, and abscisic acid are key mediators of plant secondary metabolites. World Journal of Traditional Chinese Medicine. 2021; 7: 307–325.
- [312] Wang J, Song L, Gong X, Xu J, Li M. Functions of Jasmonic Acid in Plant Regulation and Response to Abiotic Stress. International Journal of Molecular Sciences. 2020; 21: 1446.
- [313] Jaiswal AK, Alkan N, Elad Y, Sela N, Philosoph AM, Graber ER, *et al.* Molecular insights into biochar-mediated plant growth promotion and systemic resistance in tomato against Fusarium crown and root rot disease. Scientific Reports. 2020; 10: 13934.
- [314] Bian C, Duan Y, Wang J, Xiu Q, Wang J, Hou Y, et al. Validamycin A Induces Broad-Spectrum Resistance Involving Salicylic Acid and Jasmonic Acid/Ethylene Signaling Pathways. Molecular Plant-Microbe Interactions. 2020; 33: 1424–1437.
- [315] Li L, Zhu T, Song Y, Feng L, Kear PJ, Riseh RS, et al. Salicylic acid fights against Fusarium wilt by inhibiting target of rapamycin signaling pathway in Fusarium oxysporum. Journal of Advanced Research. 2022; 39: 1–13.
- [316] Noorbakhsh Z, Taheri P. Nitric oxide: a signaling molecule



- which activates cell wall-associated defense of tomato against Rhizoctonia solani. European Journal of Plant Pathology. 2016; 144: 551–568.
- [317] Kohli SK, Khanna K, Bhardwaj R, Abd Allah EF, Ahmad P, Corpas FJ. Assessment of Subcellular ROS and NO Metabolism in Higher Plants: Multifunctional Signaling Molecules. Antioxidants. 2019; 8: 641.
- [318] Saed-Moucheshi A, Pakniyat H, Pirasteh-Anosheh H, Azooz M. Role of ROS as signaling molecules in plants. In Ahmad P (ed.) Oxidative damage to plants (pp. 585–620). Elsevier: Amsterdam, The Netherlands. 2014.
- [319] Zaninotto F, La Camera S, Polverari A, Delledonne M. Cross talk between reactive nitrogen and oxygen species during the hypersensitive disease resistance response. Plant Physiology. 2006; 141: 379–383.
- [320] Anand A, Kumari A, Thakur M, Koul A. Hydrogen peroxide signaling integrates with phytohormones during the germination of magnetoprimed tomato seeds. Scientific Reports. 2019; 9: 8814.
- [321] Hossain MA, Bhattacharjee S, Armin SM, Qian P, Xin W, Li HY, *et al.* Hydrogen peroxide priming modulates abiotic oxidative stress tolerance: insights from ROS detoxification and scavenging. Frontiers in Plant Science. 2015; 6: 420.
- [322] Hemalatha G, Renugadevi J, Eevera T. Studies on seed priming with hydrogen peroxide for mitigating salt stress in rice. International Journal of Current Microbiology and Applied Sciences. 2017; 6: 691–695.
- [323] Akbudak MA, Yildiz S, Filiz E. Pathogenesis related protein-1 (PR-1) genes in tomato (Solanum lycopersicum L.): Bioinformatics analyses and expression profiles in response to drought stress. Genomics. 2020; 112: 4089–4099.
- [324] Yu XY, Bi Y, Yan L, Liu X, Wang Y, Shen KP, et al. Activation of phenylpropanoid pathway and PR of potato tuber against Fusarium sulphureum by fungal elicitor from Trichothecium roseum. World Journal of Microbiology & Biotechnology. 2016; 32: 142.
- [325] Li T, Fan P, Yun Z, Jiang G, Zhang Z, Jiang Y. β-Aminobutyric Acid Priming Acquisition and Defense Response of Mango Fruit to *Colletotrichum gloeosporioides* Infection Based on Quantitative Proteomics. Cells. 2019; 8: 1029.
- [326] Yang C, Liang Y, Qiu D, Zeng H, Yuan J, Yang X. Lignin metabolism involves Botrytis cinerea BcGs1- induced defense response in tomato. BMC Plant Biology. 2018; 18: 103.
- [327] Joseph JT, Shah JM. Biotic stress-induced epigenetic changes and transgenerational memory in plants. Biologia. 2022; 77: 2007–2021.
- [328] Louis N, Dhankher OP, Puthur JT. Seed priming can enhance and retain stress tolerance in ensuing generations by inducing epigenetic changes and trans-generational memory. Physiologia Plantarum. 2023; 175: e13881.
- [329] Sheikh AH, Nawaz K, Tabassum N, Almeida-Trapp M, Mariappan KG, Alhoraibi H, *et al.* Linker histone H1 modulates defense priming and immunity in plants. Nucleic Acids Research. 2023; 51: 4252–4265.
- [330] Kotkar H, Giri A. Plant epigenetics and the 'intelligent' priming system to combat biotic stress. In Kabelitz D, Bhat J (eds.) Epigenetics of the immune system (pp. 25–38). Elsevier: Amsterdam, The Netherlands. 2020.
- [331] Ellouzi H, Rabhi M, Khedher S, Debez A, Abdelly C, Zorrig W. Silicon seed priming enhances salt tolerance of barley seedlings through early ROS detoxification and stimulation of antioxidant defence. Silicon. 2023; 15: 37–60.
- [332] García-Espinoza F, García MJ, Quesada-Moraga E, Yousef-Yousef M. Entomopathogenic Fungus-Related Priming Defense Mechanisms in Cucurbits Impact Spodoptera littoralis (Boisduval) Fitness. Applied and Environmental Microbiology. 2023;

- 89: e0094023.
- [333] Yi HS, Yang JW, Choi HK, Ghim SY, Ryu CM. Benzothiadiazole-elicited defense priming and systemic acquired resistance against bacterial and viral pathogens of pepper under field conditions. Plant Biotechnology Reports. 2012; 6: 373–380.
- [334] Jing B, Xu S, Xu M, Li Y, Li S, Ding J, et al. Brush and spray: a high-throughput systemic acquired resistance assay suitable for large-scale genetic screening. Plant Physiology. 2011; 157: 973–980.
- [335] Ponsankar A, Senthil-Nathan S, Vasantha-Srinivasan P, Pandiyan R, Karthi S, Kalaivani K, *et al.* Systematic induced resistance in Solanum lycopersicum (L.) against vascular wilt pathogen (Fusarium oxysporum f. sp. lycopersici) by Citrullus colocynthis and Trichoderma viride. PLoS ONE. 2023; 18: e0278616.
- [336] Torres AM. Application of molecular markers for breeding disease resistant varieties in crop plants. In Jain SM, Brar DS (eds.) Molecular Techniques in Crop Improvement (pp. 185–205). 2nd edn. Springer: Dordrecht. 2009.
- [337] Stear MJ, Bishop SC, Mallard BA, Raadsma H. The sustainability, feasibility and desirability of breeding livestock for disease resistance. Research in Veterinary Science. 2001; 71: 1–7.
- [338] Hafeez U, Ali M, Hassan SM, Akram MA, Zafar A. Advances in Breeding and Engineering Climate-Resilient Crops: A Comprehensive Review. International Journal of Research and Advances in Agricultural Sciences. 2023; 2: 85–99.
- [339] Jabran M, Ali MA, Zahoor A, Muhae-Ud-Din G, Liu T, Chen W, et al. Intelligent reprogramming of wheat for enhancement of fungal and nematode disease resistance using advanced molecular techniques. Frontiers in Plant Science. 2023; 14: 1132699.
- [340] Song L, Wang R, Yang X, Zhang A, Liu D. Molecular Markers and Their Applications in Marker-Assisted Selection (MAS) in Bread Wheat (Triticum aestivum L.). Agriculture. 2023; 13: 642.
- [341] Jan S, Bhat M, Wani M, Bhat F, Kanth R, Hussain S, et al. Gene Identification and Marker Assisted Selection for Introgression of important traits in barley. Journal of Cereal Research. 2023; 15: 14–23.
- [342] Raina A, Laskar RA, Wani MR, Tomlekova N, Khan S. Plant Breeding from Classical Genetics to Molecular Approaches for Food and Nutrition Security. In Raina A, Wani MR, Laskar RA, Tomlekova N, Khan S (eds.) Advanced Crop Improvement, Volume 1: Theory and Practice (pp. 1–32). Springer: Cham. 2023.
- [343] Gocer EU, Kulak M. Marker assisted selection and breeding of crop plants. In Husen A, Ahmad A (eds.) Genomics, Transcriptomics, Proteomics and Metabolomics of Crop Plants (pp. 249–260). Elsevier: Amsterdam, The Netherlands. 2023.
- [344] Angmo D, Sharma SP, Kalia A. Breeding strategies for late blight resistance in potato crop: recent developments. Molecular Biology Reports. 2023; 50: 7879–7891.
- [345] Greenwood JR, Zhang X, Rathjen JP. Precision genome editing of crops for improved disease resistance. Current Biology. 2023; 33: R650–R657.
- [346] Kumar K, Gambhir G, Dass A, Tripathi AK, Singh A, Jha AK, *et al.* Genetically modified crops: current status and future prospects. Planta. 2020; 251: 91.
- [347] Kumar S, Chandra A, Pandey KC. Bacillus thuringiensis (Bt) transgenic crop: an environment friendly insect-pest management strategy. Journal of Environmental Biology. 2008; 29: 641–653.
- [348] Federici BA, Park HW, Bideshi DK. Overview of the basic biology of Bacillus thuringiensis with emphasis on genetic engineering of bacterial larvicides for mosquito control. Open Toxinology Journal. 2010; 3: 83–100.
- [349] Abebe A, Tafa Z. A Review On: Impact of Genetic Engineer-



- ing in Biotic Stresses Resistance Crop Breeding. International Journal for Research in Applied Sciences and Biotechnology. 2022; 9: 243–250.
- [350] Sanchis V. From microbial sprays to insect-resistant transgenic plants: history of the biospesticide Bacillus thuringiensis. A review. Agronomy for sustainable development. 2011; 31: 217–231.
- [351] Zhu F, Zhu PX, Xu F, Che YP, Ma YM, Ji ZL. Alphamomorcharin enhances Nicotiana benthamiana resistance to tobacco mosaic virus infection through modulation of reactive oxygen species. Molecular Plant Pathology. 2020; 21: 1212– 1226.
- [352] Gaffar FY, Koch A. Catch Me If You Can! RNA Silencing-Based Improvement of Antiviral Plant Immunity. Viruses. 2019; 11: 673.
- [353] Deo S, Turton KL, Kainth T, Kumar A, Wieden HJ. Strategies for improving antimicrobial peptide production. Biotechnology Advances. 2022; 59: 107968.
- [354] Lima AM, Azevedo MIG, Sousa LM, Oliveira NS, Andrade CR, Freitas CDT, et al. Plant antimicrobial peptides: An overview about classification, toxicity and clinical applications. International Journal of Biological Macromolecules. 2022; 214: 10–21.
- [355] Iqbal A, Khan RS, Shehryar K, Imran A, Ali F, Attia S, et al. Antimicrobial peptides as effective tools for enhanced disease resistance in plants. Plant Cell, Tissue and Organ Culture (PC-TOC). 2019; 139: 1–15.
- [356] Erdem Büyükkiraz M, Kesmen Z. Antimicrobial peptides (AMPs): A promising class of antimicrobial compounds. Journal of Applied Microbiology. 2022; 132: 1573–1596.
- [357] Saberi Riseh R, Dashti H, Gholizadeh Vazvani M, Dini A. Changes in the activity of enzymes phenylalanine ammonialyase, polyphenol oxidase, and peroxidase in some wheat genotypes against take-all disease. Journal of Agricultural Science and Technology. 2021; 23: 929–942.
- [358] Jin L, Wang J, Guan F, Zhang J, Yu S, Liu S, et al. Dominant point mutation in a tetraspanin gene associated with field-evolved resistance of cotton bollworm to transgenic Bt cotton. Proceedings of the National Academy of Sciences of the United States of America. 2018; 115: 11760–11765.
- [359] Wan P, Xu D, Cong S, Jiang Y, Huang Y, Wang J, et al. Hybridizing transgenic Bt cotton with non-Bt cotton counters resistance in pink bollworm. Proceedings of the National Academy of Sciences of the United States of America. 2017; 114: 5413–5418.
- [360] Ferreira SA, Pitz KY, Manshardt R, Zee F, Fitch M, Gonsalves D. Virus Coat Protein Transgenic Papaya Provides Practical Control of Papaya ringspot virus in Hawaii. Plant Disease. 2002: 86: 101–105.
- [361] Tripathi S, Suzuki J, Gonsalves D. Development of genetically engineered resistant papaya for papaya ringspot virus in a timely manner: a comprehensive and successful approach. Methods in Molecular Biology. 2007; 354: 197–240.
- [362] Gonsalves D, Tripathi S, Carr JB, Suzuki JY. Papaya ringspot virus. The Plant Health Instructor. 2010; 10: 1094.
- [363] Azad MAK, Amin L, Sidik NM. Gene technology for papaya ringspot virus disease management. The Scientific World Journal. 2014; 2014: 768038.
- [364] Tecson Mendoza EM, C Laurena A, Botella JR. Recent advances in the development of transgenic papaya technology. Biotechnology Annual Review. 2008; 14: 423–462.
- [365] Jia R, Zhao H, Huang J, Kong H, Zhang Y, Guo J, *et al.* Use of RNAi technology to develop a PRSV-resistant transgenic papaya. Scientific Reports. 2017; 7: 12636.
- [366] Colton LM, Groza HI, Wielgus SM, Jiang J. Marker-assisted selection for the broad-spectrum potato late blight resistance

- conferred by gene RB derived from a wild potato species. Crop Science. 2006; 46: 589-594.
- [367] Paluchowska P, Śliwka J, Yin Z. Late blight resistance genes in potato breeding. Planta. 2022; 255: 127.
- [368] Oosumi T, Rockhold D, Maccree M, Deahl K, McCue K, Belknap W. Gene Rpi-bt1 from Solanum bulbocastanum confers resistance to late blight in transgenic potatoes. American Journal of Potato Research. 2009; 86: 456–465.
- [369] Rakosy-Tican E, Thieme R, König J, Nachtigall M, Hammann T, Denes TE, et al. Introgression of Two Broad-Spectrum Late Blight Resistance Genes, Rpi-Blb1 and Rpi-Blb3, From Solanum bulbocastanum Dun Plus Race-Specific R Genes Into Potato Pre-breeding Lines. Frontiers in Plant Science. 2020; 11: 699.
- [370] Palti J. Cultural practices and infectious crop diseases. Springer Science & Business Media: Berlin, Germany. 2012.
- [371] Ogle H, Dale M. Disease management: cultural practices. Plant Pathogens and Plant Diseases. 1997; 390–404.
- [372] Moorman G, Gwinn KD. Cultural control of plant diseases. In Trigiano RN, Windham MT, Windham AS (eds.) Plant Pathology. Second edition. CRC press/Taylor and Francis Ggroup: Boca Raton, FL, USA. 2004.
- [373] Bowles TM, Mooshammer M, Socolar Y, Calderón F, Cavigelli MA, Culman SW, et al. Long-term evidence shows that crop-rotation diversification increases agricultural resilience to adverse growing conditions in North America. One Earth. 2020; 2: 284–293.
- [374] Reddy PP, Reddy PP. Cultural approaches. In Reddy PP (ed.) Sustainable Intensification of Crop Production (pp. 289–304). Springer: Singapore. 2016.
- [375] Dury J, Schaller N, Garcia F, Reynaud A, Bergez JE. Models to support cropping plan and crop rotation decisions. A review. Agronomy for Sustainable Development. 2012; 32: 567–580.
- [376] Liu Y, Yu Q, Zhou Q, Wang C, Bellingrath-Kimura SD, Wu W. Mapping the Complex Crop Rotation Systems in Southern China Considering Cropping Intensity, Crop Diversity, and Their Seasonal Dynamics. IEEE Journal of Selected Topics in Applied Earth Observations and Remote Sensing. 2022; 15: 9584–9598.
- [377] Richard B, Qi A, Fitt BD. Control of crop diseases through Integrated Crop Management to deliver climate-smart farming systems for low-and high-input crop production. Plant Pathology. 2022; 71: 187–206.
- [378] Samaddar S, Karp DS, Schmidt R, Devarajan N, McGarvey JA, Pires AFA, *et al.* Role of soil in the regulation of human and plant pathogens: soils' contributions to people. Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences. 2021; 376: 20200179.
- [379] Bashir H, Zafar S, Rehman R, Hussain M, Haris M, Khalid M, et al. Impact of potentially soil mineralizable Nitrogen (Pmn) on soil health and crop production. Biological and Agricultural Sciences Research Journal. 2023; 2023: 12.
- [380] Li Y, Mmbaga MT, Zhou B, Joshua J, Rotich E, Parikh L. Diseases of hydrangea. Handbook of florists' crops diseases (pp. 987–1006). Springer International Publishing AG: USA. 2018.
- [381] Eldred ZD, Tian S, Vallad GE, Zhao X, Paret M, Dufault NS. Common Tomato High Tunnel Production Diseases in Florida. EDIS. 2023; 2023: 368.
- [382] Oweis T, Hachum A. Water harvesting and supplemental irrigation for improved water productivity of dry farming systems in West Asia and North Africa. Agricultural Water Management. 2006; 80: 57–73.
- [383] Rawat L, Bisht T, Naithani DC. Plant disease management in organic farming system: strategies and challenges. In Singh KP, Jahagirdar S, Sarma BK (eds.) Emerging trends in plant pathology (pp. 611–642). Springer: Singapore. 2021.
- [384] Brar NS, Dhillon BS, Saini K, Sharma P. Agronomy of sugarbeet cultivation-A review. Agricultural Reviews. 2015; 36:



- 184-197.
- [385] Yang GZ, Luo XJ, Nie YC, Zhang XL. Effects of plant density on yield and canopy micro environment in hybrid cotton. Journal of Integrative Agriculture. 2014; 13: 2154–2163.
- [386] Ferrante A, Mariani L. Agronomic management for enhancing plant tolerance to abiotic stresses: High and low values of temperature, light intensity, and relative humidity. Horticulturae. 2018; 4: 21.
- [387] Gan Y, Siddique K, MacLeod W, Jayakumar P. Management options for minimizing the damage by ascochyta blight (Ascochyta rabiei) in chickpea (Cicer arietinum L.). Field Crops Research. 2006; 97: 121–134.
- [388] Elad Y. Cultural and Integrated Control of Botrytis spp. In Fillinger S, Elad Y (eds.) Botrytis—the fungus, the pathogen and its management in agricultural systems (pp. 149–164). Springer: Cham. 2016.
- [389] Kingra P, Kaur H. Microclimatic modifications to manage extreme weather vulnerability and climatic risks in crop production. Journal of Agricultural Physics. 2017; 17: 1–15.
- [390] Chartzoulakis K, Bertaki M. Sustainable water management in agriculture under climate change. Agriculture and Agricultural Science Procedia. 2015; 4: 88–98.
- [391] Sanogo S, Ji P. Water management in relation to control of Phytophthora capsici in vegetable crops. Agricultural water management. 2013; 129: 113–119.
- [392] Srivastav AL, Dhyani R, Ranjan M, Madhav S, Sillanpää M. Climate-resilient strategies for sustainable management of water resources and agriculture. Environmental Science and Pollution Research International. 2021; 28: 41576–41595.
- [393] Hernández-Martínez NR, Blanchard C, Wells D, Salazar-Gutiérrez MR. Current state and future perspectives of commercial strawberry production: A review. Scientia Horticulturae. 2023; 312: 111893.
- [394] Ghaffar A, Rahman MHU, Ahmed S, Haider G, Ahmad I, Khan MA, et al. Adaptations in cropping system and pattern for sustainable crops production under climate change scenarios. Improvement of Plant Production in the Era of Climate Change (pp. 1–34). 1st edn. CRC Press: Boca Raton. 2022.
- [395] Zuccarini P, Galindo A, Torrecillas A, Pardossi A, Clothier B. Hydraulic relations and water use of mediterranean ornamental shrubs in containers. Journal of Horticultural Research. 2020; 28: 49–56.
- [396] Gu Z, Qi Z, Burghate R, Yuan S, Jiao X, Xu J. Irrigation scheduling approaches and applications: A review. Journal of Irrigation and Drainage Engineering. 2020; 146: 04020007.
- [397] Giordano M, Petropoulos S, Cirillo C, Rouphael Y. Biochemical, physiological, and molecular aspects of ornamental plants adaptation to deficit irrigation. Horticulturae. 2021; 7: 107.
- [398] Aziz DA, Asgarnezhad R, Mahmood SN. The recent advances in IoT based smart plant irrigation systems: a brief review. In 2021 5th International Symposium on Multidisciplinary Studies and Innovative Technologies (ISMSIT). IEEE. 2021; 97–104.
- [399] Naveed M, Mitter B, Reichenauer TG, Wieczorek K, Sessitsch A. Increased drought stress resilience of maize through endophytic colonization by Burkholderia phytofirmans PsJN and Enterobacter sp. FD17. Environmental and Experimental Botany. 2014; 97: 30–39.
- [400] Wang Z, Luo W, Cheng S, Zhang H, Zong J, Zhang Z. Ralstonia solanacearum - A soil borne hidden enemy of plants: Research development in management strategies, their action mechanism and challenges. Frontiers in Plant Science. 2023; 14: 1141902.
- [401] Rasool F, Singh P, Anwar A, Mughal N, Kumar S. Disease management approaches-strategies and concepts. The Pharma Innovation Journal. 2022; 11: 413–417.
- [402] Gupta A, Kumar R. Management of seed-borne diseases: an integrated approach. In Kumar R, Gupta A (eds.) Seed-borne diseases

- eases of agricultural crops: detection, diagnosis & management (pp. 717–745). Springer: Singapore. 2020.
- [403] Vega D, Ibarra S, Varela Pardo R, Poggio S. Agroecological management of crop diseases: a review. Agroecology and Sustainable Food Systems. 2023; 47: 919–949.
- [404] Kumar D, Shivay Y. Integrated crop management. National Science Digital Library, NISCAIR: New Delhi, India. 2008.
- [405] Shakeel Q, Li G, Long Y, Tahir HAS. Development and Implementation of IDM Program for Annual and Perennial Crops. In Ul Haq I, Ijaz S (eds.) Plant Disease Management Strategies for Sustainable Agriculture through Traditional and Modern Approaches (pp. 295–327). Springer: Cham. 2020.
- [406] Khoury WE, Makkouk K. Integrated plant disease management in developing countries. Journal of Plant Pathology. 2010; 92: S35–S42.
- [407] Degani O, Gordani A, Becher P, Chen A, Rabinovitz O. Crop Rotation and Minimal Tillage Selectively Affect Maize Growth Promotion under Late Wilt Disease Stress. Journal of Fungi. 2022; 8: 586.
- [408] Ghimire B, Liyanage KHE, Hall C, Baysal-Gurel F. A Modified Delphi Study on Boxwood Blight Disease Management in the US Nursery Industry. HortScience. 2023; 58: 898–906.
- [409] Iftikhar Y, Sajid A, Shakeel Q, Ahmad Z, Ul Haq Z. Biological antagonism: a safe and sustainable way to manage plant diseases. In Ul Haq I, Ijaz S (eds.) Plant Disease Management Strategies for Sustainable Agriculture through Traditional and Modern Approaches (pp. 83–109). Springer: Cham. 2020.
- [410] Lahlali R, Ezrari S, Radouane N, Kenfaoui J, Esmaeel Q, El Hamss H, et al. Biological Control of Plant Pathogens: A Global Perspective. Microorganisms. 2022; 10: 596.
- [411] Prajapati S, Kumar N, Kumar S, Maurya S. Biological control a sustainable approach for plant diseases management: A review. Journal of Pharmacognosy and Phytochemistry. 2020; 9: 1514– 1523
- [412] Aguilar-Marcelino L, Mendoza-de-Gives P, Al-Ani LKT, López-Arellano ME, Gómez-Rodríguez O, Villar-Luna E, et al. Using molecular techniques applied to beneficial microorganisms as biotechnological tools for controlling agricultural plant pathogens and pest. In Sharma V, Salwan R, Al-Ani LKT (eds.) Molecular Aspects of Plant Beneficial Microbes in Agriculture (pp. 333–349). Elsevier: Amsterdam, The Netherlands. 2020.
- [413] Ingwell LL, Kaplan I. Insect Exclusion Screens Reduce Cucumber Beetle Infestations in High Tunnels, Increasing Cucurbit Yield. Journal of Economic Entomology. 2019; 112: 1765– 1773
- [414] Kusakari SI, Matsuda Y, Toyoda H. Electrostatic Insect Repulsion, Capture, and Arc-Discharge Techniques for Physical Pest Management in Greenhouses. Agronomy. 2022; 13: 23.
- [415] Hussain Z, Luqman. Effects of Mulching Practices on the Management of Weeds. In Akhtar K, Arif M, Riaz M, Wang H (eds.) Mulching in Agroecosystems: Plants, Soil & Environment (pp. 135–150). Springer: Singapore. 2022.
- [416] Khan BA, Nijabat A, Khan MI, Khan I, Hashim S, Nadeem MA, et al. Implications of Mulching on weed management in crops and vegetable. In Akhtar K, Arif M, Riaz M, Wang H (eds.) Mulching in Agroecosystems: Plants, Soil & Environment (pp. 199–213). Springer: Singapore. 2022.
- [417] Adnan M, Asif M, Khalid M, Abbas B, Hayyat MS, Raza A, et al. Role of mulches in agriculture: A review. International Journal of Botany Studies. 2020; 3: 309–314.
- [418] Dragumilo A, Marković T, Vrbničanin S, Prijić Ž, Mrđan S, Radanović D, et al. Weed suppression by mulches in Mentha x piperita L. Journal of Applied Research on Medicinal and Aromatic Plants. 2023; 35: 100499.
- [419] Wani RA, Din S, Khan M, Hakeem SA, Jahan N, Lone RA, et al. Canopy management in fruit crops for maximizing pro-



- ductivity. International Journal of Chemical Sciences. 2021; 9: 160-165
- [420] Deguine JP, Aubertot JN, Flor RJ, Lescourret F, Wyckhuys KA, Ratnadass A. Integrated pest management: good intentions, hard realities. A review. Agronomy for Sustainable Development. 2021; 41: 38.
- [421] Zehra A, Raytekar NA, Meena M, Swapnil P. Efficiency of microbial bio-agents as elicitors in plant defense mechanism under biotic stress: A review. Current Research in Microbial Sciences. 2021; 2: 100054.
- [422] Thakur M, Sohal BS. Role of elicitors in inducing resistance in plants against pathogen infection: a review. International Scholarly Research Notices. 2013; 2013.
- [423] Thaler JS, Fidantsef AL, Duffey SS, Bostock RM. Trade-offs in plant defense against pathogens and herbivores: a field demonstration of chemical elicitors of induced resistance. Journal of Chemical Ecology. 1999; 25: 1597–1609.
- [424] Sobhy IS, Erb M, Lou Y, Turlings TCJ. The prospect of applying chemical elicitors and plant strengtheners to enhance the biological control of crop pests. Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences. 2014; 369: 20120283.
- [425] Pickett JA, Poppy GM. Switching on plant genes by external chemical signals. Trends in Plant Science. 2001; 6: 137–139.
- [426] Figueroa Pérez MG, Rocha-Guzmán NE, Mercado-Silva E, Loarca-Piña G, Reynoso-Camacho R. Effect of chemical elicitors on peppermint (Mentha piperita) plants and their impact on the metabolite profile and antioxidant capacity of resulting infusions. Food Chemistry. 2014; 156: 273–278.
- [427] Vivas M, Martín J, Gil L, Solla A. Evaluating methyl jasmonate for induction of resistance to Fusarium oxysporum, F. circinatum and Ophiostoma novo-ulmi. Forest Systems. 2012; 21: 289–299.
- [428] Boughton AJ, Hoover K, Felton GW. Impact of chemical elicitor applications on greenhouse tomato plants and population growth of the green peach aphid, Myzus persicae. Entomologia Experimentalis et Applicata. 2006; 120: 175–188.
- [429] Tripathi D, Raikhy G, Kumar D. Chemical elicitors of systemic acquired resistance—Salicylic acid and its functional analogs. Current Plant Biology. 2019; 17: 48–59.
- [430] Oostendorp M, Kunz W, Dietrich B, Staub T. Induced disease resistance in plants by chemicals. European Journal of Plant Pathology. 2001; 107: 19–28.
- [431] Dasgupta D, Paul A, Acharya K, Minkina T, Mandzhieva S, Gorovtsov AV, *et al.* Bioinoculant mediated regulation of signalling cascades in various stress responses in plants. Heliyon. 2023; 9: e12953.
- [432] Lyagin I, Aslanli A, Domnin M, Stepanov N, Senko O, Maslova O, *et al.* Metal Nanomaterials and Hydrolytic Enzyme-Based Formulations for Improved Antifungal Activity. International Journal of Molecular Sciences. 2023; 24: 11359.
- [433] Dann E, Meuwly P, Métraux JP, Deverall B. The effect of pathogen inoculation or chemical treatment on activities of chitinase and β -1, 3-glucanase and accumulation of salicylic acid in leaves of green bean, Phaseolus vulgarisL. Physiological and Molecular Plant Pathology. 1996; 49: 307–319.
- [434] Alnefaie RM, El-Sayed SA, Ramadan AA, Elmezien AI, El-Taher AM, Randhir TO, et al. Physiological and Anatomical Responses of Faba Bean Plants Infected with Chocolate Spot Disease to Chemical Inducers. Life. 2023; 13: 392.
- [435] Saberi Riseh R, Gholizadeh Vazvani M, Vatankhah M, Kennedy JF. Chitin-induced disease resistance in plants: A review. International Journal of Biological Macromolecules. 2024; 266: 131105.
- [436] Riseh RS, Vazvani MG, Kennedy JF. β-glucan-induced disease resistance in plants: A review. International Journal of Biologi-

- cal Macromolecules. 2023; 253: 127043.
- [437] Carvalho FP. Agriculture, pesticides, food security and food safety. Environmental Science & Policy. 2006; 9: 685–692.
- [438] Rocha I, Ma Y, Souza-Alonso P, Vosátka M, Freitas H, Oliveira RS. Seed Coating: A Tool for Delivering Beneficial Microbes to Agricultural Crops. Frontiers in Plant Science. 2019; 10: 1357.
- [439] Mansouri H, Ait Said H, Noukrati H, Oukarroum A, Ben youcef H, Perreault F. Advances in Controlled Release Fertilizers: Cost-Effective Coating Techniques and Smart Stimuli-Responsive Hydrogels. Advanced Sustainable Systems. 2023; 7: 2300149.
- [440] Bateman RP, Hidalgo E, Garcia J, Arroyo C, Ten Hoopen GM, Adonijah V, *et al.* Application of chemical and biological agents for the management of frosty pod rot (Moniliophthora roreri) in Costa Rican cocoa (Theobroma cacao). Annals of Applied Biology. 2005; 147: 129–138.
- [441] Krauss U, Hidalgo E, Bateman R, Adonijah V, Arroyo C, García J, *et al.* Improving the formulation and timing of application of endophytic biocontrol and chemical agents against frosty pod rot (Moniliophthora roreri) in cocoa (Theobroma cacao). Biological Control. 2010; 54: 230–240.
- [442] Riseh RS, Vazvani MG, Kennedy JF. The application of chitosan as a carrier for fertilizer: A review. International Journal of Biological Macromolecules. 2023; 252: 126483.
- [443] Saberi Riseh R, Vatankhah M, Hassanisaadi M, Kennedy JF. Increasing the efficiency of agricultural fertilizers using cellulose nanofibrils: A review. Carbohydrate Polymers. 2023; 321: 121313.
- [444] Ons L, Bylemans D, Thevissen K, Cammue BPA. Combining Biocontrol Agents with Chemical Fungicides for Integrated Plant Fungal Disease Control. Microorganisms. 2020; 8: 1930.
- [445] Talboys P. Chemical control of Verticillium wilts. Phytopathologia Mediterranea. 1984; 23: 163–175.
- [446] Noman A, Aquel M, He S. CRISPR-Cas9: Tool for Qualitative and Quantitative Plant Genome Editing. Frontiers in Plant Science. 2016; 7: 1740.
- [447] Bisht DS, Bhatia V, Bhattacharya R. Improving plant-resistance to insect-pests and pathogens: The new opportunities through targeted genome editing. Seminars in Cell & Developmental Biology. 2019; 96: 65–76.
- [448] Langner T, Kamoun S, Belhaj K. CRISPR Crops: Plant Genome Editing Toward Disease Resistance. Annual Review of Phytopathology. 2018; 56: 479–512.
- [449] Gratacap RL, Wargelius A, Edvardsen RB, Houston RD. Potential of Genome Editing to Improve Aquaculture Breeding and Production. Trends in Genetics. 2019; 35: 672–684.
- [450] Zhan J, Thrall PH, Papaïx J, Xie L, Burdon JJ. Playing on a pathogen's weakness: using evolution to guide sustainable plant disease control strategies. Annual Review of Phytopathology. 2015; 53: 19–43.
- [451] Parker IM, Gilbert GS. The evolutionary ecology of novel plant-pathogen interactions. Annual Review of Ecology, Evolution, and Systematics. 2004; 35: 675–700.
- [452] Chakraborty S. Migrate or evolve: options for plant pathogens under climate change. Global Change Biology. 2013; 19: 1985– 2000.
- [453] Mundt CC. Durable resistance: a key to sustainable management of pathogens and pests. Infection, Genetics and Evolution. 2014; 27: 446–455.
- [454] Fabre F, Rousseau E, Mailleret L, Moury B. Durable strategies to deploy plant resistance in agricultural landscapes. The New Phytologist. 2012; 193: 1064–1075.
- [455] Stuthman D, Leonard K, Miller-Garvin J. Breeding crops for durable resistance to disease. Advances in Agronomy. 2007; 95: 319–367.
- [456] Camenzind M, Koller T, Armbruster C, Jung E, Brunner



- S, Herren G, *et al.* Breeding for durable resistance against biotrophic fungal pathogens using transgenes from wheat. Molecular Breeding: New Strategies in Plant Improvement. 2024; 44: 8.
- [457] Passioura JB. Translational research in agriculture. Can we do it better? Crop and Pasture Science. 2020; 71: 517–528.
- [458] Züst T, Agrawal AA. Trade-Offs Between Plant Growth and
- Defense Against Insect Herbivory: An Emerging Mechanistic Synthesis. Annual Review of Plant Biology. 2017; 68: 513–534.
- [459] Albrecht T, Argueso CT. Should I fight or should I grow now? The role of cytokinins in plant growth and immunity and in the growth-defence trade-off. Annals of Botany. 2017; 119: 725–735

