

Review

Mechanism of Endothelial Cell Death in Cerebral Ischemic Stroke and Application of Protective Drugs

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Abstract

Brain endothelial cells (BECs) are situated at the interface between the bloodstream and the brain, serving a crucial function in the development and maturation of the brain, particularly in upholding the integrity of the blood-brain barrier (BBB). Consequently, any modifications or gradual breakdown of the endothelium can significantly disrupt brain homeostasis. Ischemic stroke (IS), characterized by the progressive compromise of the BBB and increased BECs mortality, stands as a prominent global cause of mortality and disability. This review will utilize recent research to explore mechanisms underlying death.

Keywords: ischemic stroke; brain endothelial cells death; neuroprotective drugs

1. Introduction

According to research, the frequency and occurrence of strokes rose with time, and they were associated with high rates of death and disability, endangering people's lives and health [1]. Of these, cerebral ischemic stroke (IS) accounts for around 70% of all stroke occurrences, making up the vast majority [2]. Insufficient blood flow and cerebral vascular congestion are characteristics of IS, which results in a lack of oxygen and nutrients reaching brain tissue. This, in turn, causes a number of pathophysiological alterations or injuries, including the death of brain cells, which ultimately causes neurological dysfunction [3].

The blood-brain barrier (BBB) is a specialized microvascular system that functions as an essential interface between the brain and blood, safeguarding the integrity and homeostasis of the central nervous system (CNS) [4]. They are mainly composed of brain microvascular endothelial cells (BMECs), astrocytes, pericytes and basement membrane. Endothelial cells constitute the capillary wall and are the main barrier of the BBB [5]. The endfeet of astrocytes are surrounded by brain microvascular endothelial cells (BMECs), whose secreted matrix proteins form the basement membrane. Additionally, pericytes are intricately embedded within the basement membrane that encompasses both the glial cells and the BMECs [6]. Investigating the mechanisms that maintain the integrity of the BBB is critical to our understanding of the regulation of exchange between the CNS and the periphery under both

healthy and diseased states. Among them, brain endothelial cells have a role in maintaining the integrity and functionality of the BBB as well as promoting the formation of neuronal axon. They are also engaged in the regulation of vascular relaxation ability, blood cell transit, platelet adhesion, and neovascularization [7]. They form a barrier that highly limits the passage of solutes between nerve tissue and circulating blood vessels [8].

Nearly a quarter of stroke survivors had another stroke after 5 years, almost doubling after 10 years. Recurrent strokes are associated with a significantly high mortality rate; approximately 50% of individuals who survive their initial stroke pass away within five years, while around 75% do so within ten years [9]. Long-term all-cause mortality was mainly due to diseases other than stroke. Both recurrent stroke and long-term mortality are influenced by several modifiable risk factors [10]. Certain clinical trials have indicated that the annual risk of myocardial infarction or vascular mortality following an ischemic stroke varies between 1.8% and 4.6%. In a recent study, cardiovascular deaths accounted for 5 to 39% of late deaths during 10 years of follow-up [11].

As the primary effector cells involved in the angiogenic response, endothelial cells (ECs) surrounding the infarcted brain area commence proliferation as early as 12 to 24 hours following the onset of ischemic stroke [12]. Additionally, upregulation of vascular endothelial growth factor (VEGF) in the peri-infarct region has been observed as early as three hours post-ischemic injury, indicating that an-

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giogenesis can initiate within hours after the stroke occurs [13]. In the context of ischemic stroke, the migration of ECs from their stationary locations is facilitated by the loosening of intercellular connections among the ECs, coupled with a reduction in support from adjacent cells such as pericytes and smooth muscle cells. This dynamic ultimately results in vascular instability [14]. Following the onset of ischemic stroke, reactive astrocytes play a crucial role in reorganizing the extracellular matrix (ECM), leading to the formation of ECM bundles that migrating endothelial cells utilize to establish new capillary structures [15]. Once the pathway for sprouting is established, VEGF binds to its receptors on vascular endothelial cells, thereby triggering a direct angiogenic response that fosters both proliferation and migration of the ECs [16].

Decades of preclinical research have underscore the potential advantages of neuroprotection in experimental stroke models. A study indicates that a wide array of therapeutic agents has undergone testing in clinical trials or is presently under evaluation in preclinical investigations to assess their effectiveness in acute IS [17]. In recent years, drugs and compounds for the protection of brain endothelial cells have also been developed rapidly.

This review will reflect the importance of brain endothelial cells (BECs) death mechanisms in IS by introducing current information on different types of brain endothelial cell death mechanisms and mediators involved in regulating these responses, and discuss the application prospects of neuroprotective drugs, to offer dependable therapeutic strategies for the future treatment of IS (Fig. 1).

2. Morphological and Functional Characteristics of BECs

Compared to endothelial cells generated from peripheral organ arteries, BECs have distinct morphological and functional characteristics [7]. BECs are polarized cells distinguished by their intricate tight junctions, which primarily consist of zonula occludens-1 (ZO-1), claudin, and occludin, along with membrane solute carriers and ATPbinding cassette (ABC) efflux transporters. These highly specialized cells form the blood-brain barrier (BBB), playing a crucial role in regulating the selective and active transport of potentially harmful substances from the bloodstream into the brain [18]. In addition, BECs have a great number of mitochondria and a smaller number of pinopoditic vesicles, thereby limiting endocytosis and transcellular action [19,20]. These characteristics enable BECs to collaborate with astrocytes, pericytes, and perivascular microglia to create the BBB in addition to enabling them to quickly supply oxygen, glucose, and other nutrients to fulfill the high metabolic demands of the brain.

Trauma, infections, oxidative stress, DNA-damaging chemicals, and the deposition of amyloid- β peptide (A β) fragments are a few examples of acute or chronic insults that offer significant risks to BECs [21–25]. These insults have

the potential to disrupt microvascular structures and have a deleterious effect on a variety of endothelial cell activities, including survival. Reduced BEC survival ultimately jeopardizes the cerebral vascular niche, the BBB, and the morphological and functional integrity of the neurovascular unit, there by disrupting brain homeostasis and resulting in illness.

3. Mechanisms of Endothelial Cell Death in IS

To date, studies have described a variety of regulatory-cell-death mechanisms [26–28]. These include apoptosis, necroptosis, pyroptosis, ferroptosis, and autophagy-dependent cell death. These cell-death mechanisms provide a basis for further characterizing endothelial cell death in nervous system diseases, bringing hope for the treatment of these diseases [29] and to provide new therapeutic targets for the treatment of IS by studying the potential mechanism of BEC death. In this section, we will introduce the latest research on the mechanism of endothelial cell death, and classify, supplement and summarize the previous research.

3.1 Apoptosis

Of the different Endothelial-cells-death mechanisms identified in IS, apoptosis is the most extensively studied. Two distinct processes can cause apoptosis: The death receptor pathway, commonly referred to as the extrinsic pathway, operates in conjunction with the endogenous pathway, also known as the mitochondrial pathway or the Bcell lymphoma-2 (Bcl-2) regulated pathway, which is classified as the intrinsic pathway [30,31]. The intrinsic pathway is regulated by pro-apoptotic and anti-apoptotic members of the Bcl-2 protein family [32]. Activation of the extrinsic pathway occurs via members of the tumor necrosis factor receptor (TNFR) superfamily through the binding of their respective ligands [33]. Both the intrinsic and extrinsic pathways ultimately converge on a final common pathway, which emphasizes the activation of the caspase protease family. This activation culminates in the hallmark features of apoptosis, including DNA fragmentation, chromatin condensation, and membrane blistering [34]. Additionally, the extrinsic pathway can induce intrinsic mitochondrial apoptosis by activating caspase-8, which subsequently generates truncated BID (tBID) [31]. Various regulatory proteins—including tumor protein 53 (p53), nuclear factor kappa-B (NF- κ B), the ubiquitin-proteasome system, and the phosphoinositide 3-kinase (PI3K) pathway—play critical roles in modulating both the intrinsic and extrinsic pathways. Given that multiple stimuli can trigger both pathways, there exists significant crosstalk between these two apoptotic mechanisms.

ASK1-K716R point mutation was found to inhibit apoptosis signal-regulating kinase 1 (ASK1) activity, as well as the activation of pro-apoptotic Jun N-terminal kinases (JNKs) pathway and pro-inflammatory p38 pathway



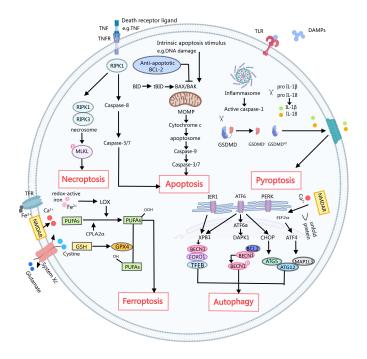


Fig. 1. A simple schematic of several death mechanisms in ischemic stroke. Necroptosis: Death ligands such as tumor necrosis factor (TNF) with its cognate receptor (TNFR) cause pleiotropic signaling, including inflammation and cell survival, cell apoptosis, and necrotizing apoptosis by key signaling molecules, receptor interaction protein kinase 1 (RIPK1) of serine/threonine decision. After inhibition of caspase-8, RIPK1 activates receptor interaction protein kinase 1 (RIPK3), leading to the formation of necrosomes. The necrosomes then phosphorylate and activate mixed lineage kinase-like (MLKL), leading to rapid membrane permeabilization and necroptosis. Apoptosis: The activation of caspase-8 triggers the subsequent activation of caspase-3 and caspase-7, ultimately resulting in cell death through the extrinsic apoptotic pathway. In contrast, the intrinsic apoptotic pathway is initiated by disturbances within the internal cellular environment, such as DNA damage, which leads to the permeabilization of the mitochondrial outer membrane (MOMP). MOMP is regulated by interactions between pro-apoptotic and anti-apoptotic B-cell lymphoma 2 (BCL-2) family proteins. Promoting apoptosis protein the Bcl-2-associated X protein (BAX) and related the BCL-2 cognate antagonist killer protein (BAK) formed in the mitochondrial outer membrane aperture, resulted in the release of cytochrome c and the formation of apoptotic body apoptotic protease activating factor-1 (APAF-1). Apoptotic bodies activate caspase-9 and subsequently caspase-3 and caspase-7, leading to apoptosis. Caspase-8 through interaction will promote apoptosis BH3 domain death (BID) cut to activate BAX and BAK truncated BID (tBID), crosstalk between inner and external mediated apoptosis pathway. Pyroptosis: Damage-associated molecular patterns (DAMPs) released from dying cells play a crucial role in activating pattern recognition receptors, including Toll-like receptors (TLRs). This leads to the activation of the classical inflammasome that activates caspase-1. Cracking gasdermin D (GSDMD) activated caspase-1, from GSDMD C inhibition of fragments of release GSDMD NT into a hole. GSDMD NT formed holes in membrane air and coke. Active caspase 1 also proinflammatory cytokines can be interleukin 1 beta (IL-1 beta) and IL-18 GSDMD hole to release the mature form of cracking. Ferroptosis: Typically, glutathione peroxidase 4 (GPX4) utilizes glutathione (GSH) to catalyze the conversion of lipid hydroperoxides into corresponding alcohols. However, elevated levels of extracellular glutamate can inhibit the function of System Xc, consequently disrupting the cysteine-GSH-GPX4 axis. In addition, Ca^{2+} overload triggers the activation of cytosolic phospholipase $A2\alpha$ (cPLA2 α), which generates substrates essential for lipid peroxidation. Furthermore, Fe²⁺ activates lipoxygenase (LOX), further contributing to this lipid peroxidation process. The rise in blueox-active iron, coupled with excessive lipid peroxidation, plays a significant role in the induction of ferroptosis. Autophagy: Nmethyl-D-aspartic acid receptor (NMDAR) leads to Ca²⁺ influx, causing unfolded protein and endoplasmic reticulum (ER) stress. The three transmembrane sensors inositol-requiring enzyme 1 (IRE1), activating transcription factor 6 (ATF6), and PKR-like endoplasmic reticulum kinase (PERK) activate a complex cascade with autophagic induction. IRE1 upregulates Beclin 1 (BECN1) via box-binding protein 1 (XBP1). XBP1 also functions as a transactivator for forkhead box O1 (FOXO1) and transcription factor EB (TFEB), effectively bypassing BECN1 to induce autophagy. The protein PERK enhances the expression of activating transcription factor 4 (ATF4) and CCAAT/enhancer binding protein homologous protein (CHOP) through the eukaryotic initiation factor 2 (eIF2) pathway. This process subsequently increases the transcription levels of microtubule-associated protein 1 light chain 3 beta (MAP1LC3), autophagy-related protein 5 (ATG5), and autophagy-related protein 12 (ATG12). Moreover, ATF6 promotes the phosphorylation of BECN1 mediated by death-associated protein kinase 1 (DAPK1) and is believed to concurrently activate both XBP1 and CHOP. Created using MedPeer.cn.



in the brain. Further studies confirmed that ASK1-K716R inhibited downstream endothelial cell c-Jun N-terminal kinases (JNKs) activation, endothelial cell apoptosis and tight junction (TJ) protein loss, and protected the integrity of the blood-brain barrier. It can also reduce neuronal damage and white matter structure/function damage, reduce the inflammatory response in the brain, and finally improve the long-term sensory motor and memory dysfunction after traumatic brain injury (TBI) in mice [35].

Additionally, research has indicated that exosomes derived from endothelial cells have the capability to directly shield nerve cells against cerebral ischemic injury. They achieve this protective effect by fostering cell growth, enhancing migration and invasion, as well as inhibiting apoptosis [36]. Endothelial-cell-derived exosomes modification has the potential to be developed as a new therapeutic strategy to treat neuronal damage during cerebral ischemic injury.

Several case-control and epidemiological studies have shown that stroke risk is associated with clinical periodontitis [37,38]. Gingipains of Porphyromonas gingivalis have been found in brain tissue [39]. Live Porphyromonas gingivalis and its virulence factors are powerful initiators of inflammation in the brain, directly affecting memory and lesion development [40]. Research has indicated that viable but non-heat-killed Porphyromonas gingivalis can induce apoptosis and contribute to cell death in brain endothelial cells. The infection by Porphyromonas gingivalis activates the reactive oxygen species (ROS)/NF- κ B p65 signaling pathway, which in turn leads to the increased expression of interleukin-1 beta (IL-1 β) and tumor necrosis factor-alpha (TNF- α) [41].

Apoptosis is the most common mechanism of cell death. The above studies show that cerebral endothelial cell apoptosis plays a very important role in IS, but many mechanisms still need to be further explored.

3.2 Necroptosis

Necroptosis represents a specific type of regulated necrosis that is initiated by death receptors [42]. This particular form of necrosis plays a crucial role in the body's defense against infections caused by pathogens and is morphologically distinguished by the phenomenon of cell swelling, which is subsequently accompanied by the disruption of the plasma membrane [43]. Necrosis is triggered by the activation of other cellular receptors, such as Fas and TNF-related apoptosis-inducing ligand (TRAIL), which induce the production of interferon- γ (Ifn- γ) and TNF- α , thereby promoting necroptosis in an autocrine feedback loop [44].

The key step underlying TNF- α -induced necroptosis pathway is the kinase regulation process of receptor-interacting protein 1 (RIP1)-RIP3-mixed lineage kinase-like (MLKL). Phosphorylation of these molecules not only promotes the formation of necrosomes but also simulta-

neously triggers multiple cascades that mediate multiple pathological processes leading to further damage [45,46]. Necroptosis has been demonstrated in an increasing number of cell types in different pathophysiological microenvironments, such as aging [47], cancer [48], neurodegeneration [49,50], and autoimmune diseases [51]. Furthermore, necroptosis is a major factor in a number of ischemic illnesses. The function of necroptosis in mediating EC harm following ischemia/reperfusion (I/R) injury, however, has not received much attention in research. Determining the fundamental processes that connect necroapoptosis to EC loss would therefore be beneficial and could lead to the development of sensible treatment plans for vascular injury and BBB disruption after cerebral I/R injury.

In IS, Chen et al. [51] found for the first time that necroptosis of brain endothelial cells induced by TNF- α may be an important cause of BBB destruction after IS [52]. Activation of phosphoinositide 1-kinase (RIPK1) is one of the major detrimental mechanisms processes that occur after cerebral ischemic injury. Activation of RIPK1 in endothelial cells promotes necroptosis and cerebrovascular injury. Necroptosis can be simultaneously blocked by inhibition of RIPK1 kinase, and necroptosis of endothelial cells can be inhibited by Ripk3^{-/-} to rescue intracerebral hemorrhage and inflammation [53]. Macrophage migration inhibitory factor (MIF) has recently been found to induce endothelial cell apoptosis and necroptosis in perioperative stroke (PIS) through a RIPK1-dependent pathway. A combined strategy targeting MIF and RIPK1 may be more effective than using a single agent targeting either alone [54].

The above studies suggest an important role of CEs necroptosis in the development of IS. Therefore, understanding the mechanisms of endothelial necroptosis in IS may help us to better understand the pathogenesis of the disease and provide new therapeutic approaches.

3.3 Pyroptosis in Endothelial Cells

A proinflammatory type of cell death is called pyroptosis [41]. Pyroptotic cell death shares morphological characteristics with both necrosis and apoptosis. The morphological alterations include nuclear condensation and DNA fragmentation akin to apoptosis, cellular swelling, hole creation, necrosis-like cell membrane rupture, and proinflammatory intracellular content release [55].

Pyroptosis is a molecular signature of gasderminmediated cell death [56]. Normally, two caspase-dependent mechanisms activate the gasdermin [57]. The inflammasome in the traditional caspase-1 pathway identifies substances linked with danger and pathogens that are released by dying cells, along with certain proinflammatory cytokines.

Gasdermin D (GSDMD) is cleaved by activated caspase 1, which leads to the oligomerization of the GSDMD-N domain in the cells. This finally forms pores and releases cell contents, including high mobility group box 1 protein



(HMGB1) and IL-1 α [58]. Furthermore, IL-1 β /18, which promotes inflammation, is processed and matured by activated caspase 1 [59]. In the non-classical caspase 4/5/11 pathway, lipopolysaccharide in the cytoplasm directly interacts with caspase 4/5/11 to initiate its activation. Then, GSDMD breakage and pyroptosis are directly triggered by activated caspase 4/5/11 [60].

The various cell types that undergo pyroptotic cell death in the brain have not been extensively studied. However, studies have shown that BBB cells, especially BECs, are damaged and release proinflammatory cytokines after stroke [61]. In the early stage of stroke, brain endothelial pyroptosis may be induced mainly by IL-1 and reactive oxygen species (ROS) released from microglia, so the result is carbon skeleton rearrangement and loss of ligand protein function in brain endothelial cells, ultimately leading to initial BBB damage [62]. Several mitochondria are seen in BECs. As a result, a significant amount of ROS is generated following pyroptosis as a result of mitochondrial malfunction, which might harm other BBB-forming cells and compromise the integrity of the BBB [63].

MCC950, an inhibitor of the NOD-like receptor protein 3 (NLRP3) inflammasome, has been shown to specifically block the activation of NOD-like receptor protein 1 (NLRP1) and the secretion of IL18b, IL3, and GS-DMD by preventing the oligomerization of apoptosisassociated speck-like protein containing a card (Apoptosisassociated speck-like protein) induced by NLRP3 [64]. In recent years, pyroptosis mediated by the NLRP3 inflammasome has been recognized as a potential contributor to the death of brain endothelial cells (BECs). Advanced glycation end products (AGEs) can initiate pyroptosis in endothelial cells through the hypoxia-inducible factor (HIF)-the receptor of advanced glycation endproducts (RAGE)-NLRP3 pathway, thereby exacerbating brain injury [65]. Emerging research indicates that the activation of peroxisome proliferator activated receptor γ coactivator-1 (PGC-1)/peroxisome proliferators-activated receptor (PPAR) pathways, which stimulates phenylalanine metabolism and reduces mitochondrial reactive oxygen species (ROS), may serve as potential mechanisms to mitigate pyroptosis in brain endothelial cells [66]. The other study has shown that inhibition of long non-coding RNA (lncRNA) Xist can alleviate brain endothelial cell pyroptosis to alleviate IS injury [63].

A recent study has shown that gram-negative bacterial infection or lipopolysaccharide stimulation activates the caspase-4/22-GSDMD signaling pathway in brain endothelial cells, leading to inflammatory destruction of the bloodbrain barrier [67]. This finding not only provides a new perspective to understand the mechanism of BBB disruption in the inflammatory state caused by pyroptosis, but also provides a potential therapeutic approach for central nervous system diseases associated with BBB loss.

Although our understanding of the effects of pyroptosis in BECs during IS is increasing, the exact mechanism of how pyroptosis senses different stimulus information is still largely unknown and remains a direction for future exploration.

3.4 Ferroptosis

Ferroptosis is an iron-dependent, planned cell death that is brought on by the build-up of lipid peroxides, increased ROS generation, and glutathione peroxidase 4 (GPX4) inactivation [68]. Ferroptosis apparently has a role in a number of neurological conditions, including Parkinson's, Alzheimer's, and stroke [69,70]. Moreover, IS has been shown to improve with ferroptosis inhibitor therapy [71].

Ferroptosis plays a key role in neuronal cell death. However, any studies targeting other cell types have not been performed. Following a hemorrhagic stroke, disorders in iron metabolism and the accumulation of reactive oxygen species (ROS) are observed in endothelial cells, microglia, and astrocytes [72], but studies targeting other cell types have not been performed. Endothelial ferroptosis is a potential pathogenic mechanism of stroke.

Findings that show that hypoxia-induced ferroptosis in BECs and the inhibitory effect of ferrostatin-1 (Fer-1) on ferroptosis, alleviated hypoxia-induced BBB disruption, open the possibility of new potential targets for the treatment of central nervous system (CNS) diseases associated with BBB breakdown [73]. A recent study has shown that the middle cerebral artery occlusion middle cerebral artery occlusion (MCAO) model in zebrafish leads to ferroptosis of BECs, accompanied by lipid peroxidation, increased iron concentration, and decreased expression of solute carrier family 7 member 11 (SLC7A11) and GPX4, indicating that ferroptosis of endothelial cells is a potential pathogenic mechanism for IS [74].

Ferroportin 1 (FPN1) is the sole identified iron export protein located in brain endothelial cells, serving as a crucial molecule that facilitates the transfer of iron from the bloodstream into the brain [75]. Specific knockdown of FPN1 in ECs was found to produce differential effects in the acute and recovery phases of IS. In the acute phase of IS, the knockout of FPN1 in endothelial cells results in a decrease in brain iron accumulation, which subsequently alleviates oxidative stress and the inflammatory response. This reduction in oxidative and inflammatory processes leads to a decrease in ferroptosis and apoptosis, ultimately resulting in a reduction of cerebral infarct volume and an improvement in neurological function. Conversely, the knockout of FPN1 in recovering brain endothelial cells significantly hinders the restoration of neurological function following IS. This impairment occurs due to increased brain iron accumulation, promotion of gliosis, and suppression of neural stem cell migration and differentiation. Iron restriction pla-



ys a neuroprotective role in the acute phase of IS and an inhibition of the recovery phase [41].

The heme oxygenase 1 (HO-1) pathway apparently may be overexpressed, which may lessen ferroptosis [76]. According to other research, ferroptosis is significantly aided by HO-1 overexpression, which contributes to the rise in the labile iron pool [77,78]. HO-1 and transferrin receptor protein 1 (TFR1) are thought to have a role in cellular adaptation processes, iron homeostasis, and oxidative stress in the brain [79]. It is possible to ameliorate cerebral ischemia-reperfusion injury by inhibiting HO-1 [80]. Its function in acute damage may be influenced by HO-1 expression level. According to these findings, ischemia/reperfusion and hyperglycaemia (HG) cause increased HO-1 production in endothelium, which leads to ferroptosis and the progression of BBB damage both in vitro and in vivo. However, purinergic receptor P2X7 (P2RX7) blockade can reverse this process. These findings shown that P2RX7 blocking has a significant role in ferroptosis pathways and can control SLC7A11/GPX4 via inhibiting the extracellular-regulated kinase 1/2 (ERK1/2) and p53 pathway [81].

Cerebral ischemia can cause ferroptosis, which exacerbates the damage caused by cerebral ischemia. By inhibiting ferroptosis, this ischemia harm is lessened. Iron overload and the regulatory mechanisms of GPX4 and 12/15 lipoxygenases (LOX) upstream remain to be further investigated, as do the regulatory processes involved in brain ferroptosis.

3.5 Autophagy

Autophagy is a self-eating process that occurs in a variety of cells, including neurons, glia cells, and brain microvessel cells (BMVECs), and it is involved in maintaining cellular homeostasis and proper cellular functions [82,83]. After cerebral ischemia, endothelial autophagy activation restores and preserves BBB integrity, which reduces brain edema and prevents stroke [84,85].

A recent study has shown that oxygen-glucose deprivation/reoxygenation (OGD/R)-mediated upregulation of circ-forkhead box O3 (FOXO3) in cerebral endothelial cells promotes autophagy. Knockdown of circ-FOXO3 can inhibit autophagy and enhance the permeability of endothelial cells. Circ-FOXO3 plays a protective role against OGD/R-induced endothelial cell injury in an autophagy-dependent manner [86].

Furthermore, occludin degradation is mediated by ischemia-induced autophagy, and BBB dysfunction is lessened and occludin degradation is restored by 3-methyladenine (3-MA)-induced autophagy inhibition [87]. Additionally, in BECs exposed to OGD, autophagic lysosomal activation degrades claudin-5 [88], and in mice lacking the *p50* gene, autophagic activation was linked to BBB damage [89], indicating a potential role for autophagy in ischemia neuronal and vascular injury.

In contrast to these findings, recent reports have high-lighted the protective role of body BECs in autophagy and their involvement in BBB dysfunction during I/R injury [90]. In their investigation, they found that whereas pretreatment with 3-MA inhibited autophagy and increased I/R-induced BBB damage, activation of the autophagic process with the autophagy induce rapamycin and lithium carbonate dramatically reversed BBB disruption following I/R injury. In their investigation, the scientists came to the conclusion that autophagy prevented cells from producing reactive oxygen species and restored lower levels of ZO-1, despite the fact that no particular biological targets of activated autophagy were found.

Thus, autophagy after cerebral ischemia increases cerebral endothelial permeability, and this permeability can be reduced by the inhibition of autophagy. These findings enhance our comprehensive understanding of the role that autophagy plays in the dysfunction of the BBB following ischemic events.

3.6 Other Types of Endothelial Cell Death

In addition to the aforementioned types, programmed cell death encompasses parthanatos, entotic cell death, alkalosis-induced cell death, oxygen ptosis, autophagy-dependent cell death, reticulocyte death, and lysosome-dependent cell death.

Parthanatos is a kind of planned cell death that requires polymerase 1 (PARP-1) (ADP-ribosome) [91,92]. Poly (ADP-ribose) (PAR) accumulates as a result of PARP-1 overactivation following after substantial DNA damage [93]. Apoptosis-inducing factor (AIF) is released when PAR is translocated from the nucleus to the mitochondria. AIF forms a complex with macrophage migration inhibitory factor (MIF) within the cytoplasm after its release from the mitochondria. The translocation of the AIF/MIF complex into the nucleus leads to chromatin condensation and DNA fragmentation, ultimately culminating in cell death [94,95]. Neurodegenerative illnesses and other conditions associated with DNA damage may be affected by the interplay between this type of dependent cell death and other types of dependent cell death, such as necroptosis.

Autophagy-dependent cell death is a kind of programmed cell death driven by the molecular mechanism of autophagy [96]. Researches believe that ferroptosis is an autophagy-dependent cell death through a complex feedback loop, this provides new ideas for the study of autophagy-dependent cell death in endothelial cells [97].

Lysosome-dependent cell death (LCD), is a form of regulatory cell death mediated by iron translocation caused by lysosomal components or lysosomal membrane permeabilization in order to amplify or initiate cell death during apoptosis, autophagy, and ferroptosis [98]. However, the research on lysosome-dependent cell death in BECs still needs to be expanded.



In conclusion, there are various forms of endothelial cell death after stroke, but the research investigation shows that apoptosis is the main form of endothelial cell death after stroke [99], and the research progress on alleviating endothelial cell apoptosis should be further strengthened.

3.7 Effect of Other Factors on Endothelial Cell Death

Research has uncovered a previously unrecognized function of myeloid-derived MIF (macrophage migration inhibitory factor) in facilitating apoptosis and necrosis in cerebrovascular endothelial cells (ECs) through the activation of RIPK1 (receptor interacting protein kinase 1). In a clinically relevant model of postischemic syndrome (PIS), it was demonstrated that surgical trauma-induced aseptic inflammation can enhance MIF expression. Moreover, MIF has been shown to serve as a potent trigger for the death of cerebrovascular ECs and the disruption of the BBB following ischemic brain injury. By integrating both genetic and pharmacological evidence, it is suggested that targeting the release of myeloid MIF or inhibiting RIPK1 activation may confer protective effects on cerebrovascular ECs and the BBB in the context of ischemic brain injury [53].

In addition, Microglia are macrophages in the brain, which are considered to be the sentinel of neuroinflammatory response caused by different brain injuries [100]. Therefore, the regulatory effect of microglia in central nervous system diseases has received extensive attention. Microglia is one of the important components of the neurovascular unit, which plays a coordinated role with other cell types. Acute or subacute cerebral ischemic injury, which is common in the brain, will destroy the cerebral microvasculature and lead to subsequent inflammatory response, accompanied by aggravated neuronal damage and activation of microglia [101].

Microglial activation can be detected at different stages of the pathological process of stroke. Liang *et al.* [102] verified that chemokine ligand receptor CX3CL1-CX3CR1 signaling is involved in basal intercellular communication for microglial chemotaxis and activation [103]. This study reveals that there exist various categories of cell signaling cascades between neurons, microglia and brain microvascular cells. The mechanism of bidirectional regulation of microglia in neurovascular coupling is an important molecular event that needs to be clarified urgently, which will provide key ideas for clinical treatment. In addition, pericytes act as vascular smooth muscle in brain capillaries. Some pericytes respond to vasoactive signals generated by the brain by contracting, thereby affecting capillary diameter [104].

Pericytes are essential in establishing and maintaining vascular structure and BBB function. Pericyte depletion has been found in both acute and chronic CNS diseases, with rapid apoptosis following IS and traumatic brain injury (TBI). This study has reported the behavior of pericyte series changes at different stages of ischemic stroke [105].

In the acute phase of stroke, pericytes shrink to clog capillaries and cause no-return of blood. Interestingly, however, pericytes subsequently have proinflammatory and immunomodulatory effects, stabilizing the BBB and protecting the brain parenchyma by protecting endothelial cells on the lateral side of the lumen and releasing neurotrophin. In addition, pericytes have neuroprotective activity and promote angiogenesis and neuronal growth during the poststroke recovery phase. It is evident that pericytes play multiple intervening roles in the complex process of ischemia-reperfusion injury and repair in response to the dynamic changes of endothelial cells and neurons.

4. Drugs for the Treatment of IS

Although there are still few therapeutic options for stroke, recanalization therapy involving medication and mechanical thrombolysis have shown some beneficial effects in people recover from IS. Therapeutic medicines for neuroprotection in acute IS are still needed in order to avoid brain damage before and after recanalization, to extend the therapeutic window for intervention, and to improve functional outcome.

The existing literature clearly indicates that numerous therapeutic agents have undergone testing in clinical trials or are actively being assessed in preclinical studies to determine their efficacy in acute ischemic stroke (IS) [106–108]. However, despite these extensive endeavors, the identification of clinically effective neuroprotective agents continues to be a challenging pursuit. However, these studies lay the foundation for future treatment of IS with neuroprotective BECs. In recent years, several compounds targeting the inhibition of BEC-death signals have been used in IS experiments.

Dichloroacetic acid (DCA) is a small molecule that has been employed as a therapeutic agent for various genetic mitochondrial disorders [98]. Research has demonstrated that DCA may mitigate the detrimental effects of cerebral ischemia/reperfusion (I/R) by decreasing infarct volume, improving neurological scores, and reducing brain water content. Furthermore, DCA treatment has been found to diminish the impact of OGD on mitochondrial metabolism, BBB permeability, and oxidative stress in human brain microvascular endothelial cells (HBMEC) [109].

Medioresinol, a natural product [110], is a novel PGC- 1α activator that can promote the interaction between PGC- 1α and PPAR α in BMVECs and increase the expression of glutamic-oxaloacetic transaminase 1(GOT1) and phenylalanine hydroxylase (PAH), thereby improving the accumulation of phenylalanine caused by ischemia and reducing mitochondrial ROS (mtROS) [66].

Ergothioneine is a naturally occurring antioxidant. Research has demonstrated that ergothioneine can cross the blood-brain barrier and function as a cytoprotective and antioxidant agent, making it a viable treatment option for neurodegenerative diseases in which oxidative stress is fre-



Table 1. Compounds targeting BECs death.

Drugs or compounds	Mechanism of action	Effects	Type of research
DCA	Inhibition of PDK2 activates PDH, thereby activating Nrf2 and reducing	Apoptosis [109]	preclinical
	oxidative stress.		
PNS	Increased Akt phosphorylation, nuclear Nrf2 activity, and downstream	Apoptosis [122]	preclinical
	antioxidant enzyme HO-1 expression.		
Ergothioneine	Significantly reduced the proinflammatory genes IL-1 β , IL-6, IL-8,	Apoptosis [111]	preclinical
	$TNF\alpha$, NF-kB, and COX2's increase caused by 7KC.		
Baicalin	It increased the expression of Nrf2, HO-1, and NQO1, reduced the gen-	Apoptosis [119]	preclinical
	eration of ROS and MDA, and encouraged the creation of SOD.		
Cardamonin	MACO-induced brain damage and OGD/R-induced HBMEC damage	Apoptosis [116]	preclinical
	are avoided when the HIF-1 α /VEGFA pathway is activated.		
PCA	Decreased the expression of NIMA-associated kinase 7, GSDMD,	Pyroptosis [124]	preclinical
	Caspase-1, IL-1 β , and NLRP3.		
Medioresinol	Pyroptosis, mtROS, and the expression of associated proteins (NLRP3,	Pyroptosis [66]	preclinical
	ASC, cleaved caspase-1, IL-1 β , GSDMD-NT) were all dramatically de-		
	creased with PGC-1 α activation.		
Caffeic acid	To withstand ferroptosis, TFR1 and ACSL4 were downregulated, and	Ferroptosis [127]	preclinical
	glutathione synthesis was increased via the Nrf2 signaling pathway.		
PBT434	Increased in the abundance of the transcripts for TfR and ceruloplasmin	Ferroptosis [129]	preclinical
3-MA	Inhibition of autophagy reversed occludin degradation and attenuated	Autophagy. [131]	preclinical
	BBB dysfunction		
Cariside II	To reduce damage caused by cerebral I/R by disrupting the PKG/GSK-	Autophagy [134]	preclinical
	3β /autophagy axis.		

Abbreviations: DCA, dichloroacetic acid; PNS, panax notoginseng saponins; ET, ergothionein; PCA, protocatechuic aldehyde; MDN, medioresinol; 3-MA, 3-Methyladenine; PDK2, pyruvate dehydrogenase kinase 2; PDH, pyruvate dehydrogenase; Nrf2, nuclear factor erythroid 2-related factor 2; HO-1, heme oxygenase 1; TNF α , tumor necrosis factor- α ; NF-kB, nuclear factor kappa-B; COX2, cyclooxygenase 2; 7KC, 7-ketocholesterol; NQO1, NAD(P)H quinone oxidoreductase 1; ROS, reactive oxygen species; MDA, malondialdehyde; SOD, superoxide dismutase; MCAO, middle cerebral artery occlusion; OGD/R, oxygen-glucose deprivation/reoxygenation; HBMEC, human brain microvascular endothelial cells; HIF-1 α , hypoxia inducible factor-1 α ; VEGFA, vascularendothelial growth factor A; GS-DMD, gasdermin D; NOD, nucleotide-binding oligomer-zation domain; NLRP3, NOD-like receptor protein 3; ASC, apoptosis-associated speck-like protein containing a CARD; PGC, proliferator-activated receptor-gamma coactivator; TFR1, Transferrin receptor protein 1; ACSL4, acyl-CoA synthetase long chain family member 4; BBB, blood-brain barrier; Akt, protein kinase B; MACO, middle cerebral artery occlusion; PKG, protein kinases G.

quently the driving force behind disease progression [111]. A study has also shown that ergothioneine can attenuate the apoptosis of brain endothelial cells caused by the accumulation of 7-ketocholesterol, and can treat neurovascular diseases by reducing the damage to brain endothelial cells [112].

Cardamonin is a chalcone with neuroprotective activity [113]. Previous research has indicated that cardamonin has a protective effect against apoptosis induced by adriamycin or lipopolysaccharide in cardiac cells [114]. More significantly, it has been observed to reduce oxidative-stress-induced apoptosis in PC12 cells [115]. Recent research has additionally demonstrated that cardamonin can reduce OGD/R-induced increased cerebral endothelial-cell permeability, apoptosis, and brain damage in mice with MCAO by activating the HIF- 1α /vascularendothelial growth factor A (VEGFA) pathway, suggesting that cardamonin may have a neuroprotective effect in IS [116].

A significant traditional Chinese medicinal herb is Scutellaria baicalensis [117]. One of the primary bioactive ingredients in Scutellaria baicalensis extract is baicalin. Reported pharmaceutical qualities of baicalin include anti-inflammatory, anti-tumor, anti-diabetic, anti-cancer, cardioprotective, liver-protective, and neuroprotective effects [118]. The BBB can be shielded from a lipopolysaccharide challenge by baicalin administration. Inhibition of the generation of ROS by the nuclear factor erythroid 2-related factor 2 (Nrf2) antioxidant pathway, and BBB endothelial cell inflammatory response, can influence this process [119]. These results suggest that baicalin has a strong protective effect against brain damage caused by lipopolysaccharide, which offers a therapeutic treatment alternative.

Panax notoginseng saponins (PNS) have gained widespread application in the treatment of ischemic stroke (IS) and cardiovascular diseases within China [120]. PNS seems to possess a multitude of pharmacological actions, including anti-thromboembolism, anti-inflammation, anti-



apoptosis, hemostasis, cerebral vasodilatation, anticoagulation, anti-hyperglycemia, and anti-hyperlipidemia effects [121]. According to an earlier study, PNS can reduce the degradation of ZO-1 and claudin-5 tight-junction proteins by antioxidant activation of Nrf2 antioxidant signaling through the PI3K/Akt pathway and prevent OGD/R-induced BBB integrity degradation *in vitro* [122].

Protocatechuic aldehyde (PCA) is a hydrophilic phenolic compound that is extracted from the dried roots of the traditional Chinese herb, sage [123]. Numerous investigations have verified that PCA reduces the inflammatory damage to endothelial cells, preserving endothelial cell function [124]. Recent research has demonstrated that PCA reverses brain endothelial cell pyroptosis via lncRNA Xist, which strongly enhances the protective effect of PCA on IS [67].

Caffeic acid, as a natural bioactive phenolic acid, has been tested for its potential antioxidant properties [125]. Vegetables, fruits, and coffee are high in caffeic acid. According to earlier research, caffeic acid helped rat brain damage during cerebral I/R [126]. According to recent research, in the brain of the MCAO rat, caffeic acid suppresses oxidative stress-mediated neuronal death, regulates ferroptosis, upregulates glutathione production through the Nrf2 signaling pathway, and downregulates transferrin receptor protein 1 (TFR1) and acyl-CoA synthetase long chain family member 4 (ACSL4). Caffeic acid has been proposed as a possible treatment to lessen brain damage after cerebral ischemia [127].

PBT434 methanesulfonate is a potent and orally active α -synucleinaggregation inhibitor that can cross the BBB [128]. It has been shown that the brain supports plasma membrane iron reduction of transferrin bound iron; PBT434, like ferrozine, prevents 55Fe uptake from 55Fe-Tf by hBMVEC [129].

3-MA is a mature autophagy inhibitor. A study has shown that 3-MA helps to enhance the cell viability of brain endothelial cells by inhibiting autophagy, and reduces the level of occludin and the cell hyperpermeability induced by glucose and oxygen deprivation [130]. In addition, 3-MA administration significantly reduced the I/C values increased by Evans blue in an *in vivo* study [131].

Icariin (icariside II - ICS II) Icariin, also known as icariside II (ICS II), is one of the primary active components derived from Epimedium, a Traditional Chinese Medicine widely employed in the treatment of various clinical conditions such as dementia, erectile dysfunction, and cardiovascular diseases [132]. ICS II is recognized for its antioxidant properties and demonstrated neuroprotective effects [133]. According to recent research, ICS II both inactivates glycogen synthase kinase-3 β (GSK-3 β) and restores the cyclic guanosine monophosphate-protein kinase G (cGMP/PKG) pathway, which reduces the damage caused by cerebral I/R. This reduction is attributed to the suppression of excessive autophagy. Consequently, the PKG/GSK-3 β /autophagy axis is crucial in controlling the beneficial effects of ICS

II on excessive autophagic neuronal death brought on by cerebral I/R [134]. The mechanisms of drugs for the treatment of cerebral ischemia are summarized in Table 1 (Ref. [66,109,111,116,119,122,124,127,129,131,134]).

5. Conclusions

IS is a prevalent and often fatal disease globally, yet the precise pathogenic mechanism remains elusive despite advancements in experimental and clinical research. Endothelial cell dysfunction, a key element of the blood-brain barrier, is intricately linked to the development of IS. Although some headway has been made in identifying specific targets for endothelial cell death through experimental studies, the efficacy of therapeutic drugs in treating ischemic stroke remains uncertain due to variations between experimental models and actual clinical cases. Numerous challenges persist in the development of novel and precise models, as well as more efficient treatment modalities. Nevertheless, opportunities for advancement in disease research remain abundant, particularly in the investigation of endothelial-cell-death mechanisms which may inform the development of synergistic therapies involving other neural cells. Consequently, elucidating the mechanisms underlying endothelial cell death serves as a crucial step in the advancement of therapeutic strategies for IS.

Author Contributions

HRY and HH jointly designed and conceived the article and made revisions to the important intellectual content of the paper. QWM and QBL collected the references, made the figures and tables, made the final revisions to the version to be published and agreed to take responsibility for all aspects of the research work to ensure that any issues related to the accuracy or integrity of any part of the paper are properly investigated and resolved. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

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Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Wu S, Wu B, Liu M, Chen Z, Wang W, Anderson CS, *et al.* Stroke in China: advances and challenges in epidemiology, prevention, and management. The Lancet. Neurology. 2019; 18: 394–405. https://doi.org/10.1016/S1474-4422(18)30500-3.
- [2] Wang L, Zhang X, Xiong X, Zhu H, Chen R, Zhang S, et al. Nrf2 Regulates Oxidative Stress and Its Role in Cerebral Ischemic Stroke. Antioxidants. 2022; 11: 2377. https://doi.org/10.3390/antiox11122377.
- [3] Hitomi E, Simpkins AN, Luby M, Latour LL, Leigh RJ, Leigh R. Blood-ocular barrier disruption in patients with acute stroke. Neurology. 2018; 90: e915–e923. https://doi.org/10.1212/WN L.0000000000005123.
- [4] Obermeier B, Daneman R, Ransohoff RM. Development, maintenance and disruption of the blood-brain barrier. Nature Medicine. 2013; 19: 1584–1596. https://doi.org/10.1038/nm.3407.
- [5] Abbott NJ, Rönnbäck L, Hansson E. Astrocyte-endothelial interactions at the blood-brain barrier. Nature Reviews. Neuroscience. 2006; 7: 41–53. https://doi.org/10.1038/nrn1824.
- [6] Greene C, Hanley N, Campbell M. Claudin-5: gatekeeper of neurological function. Fluids and Barriers of the CNS. 2019; 16: 3. https://doi.org/10.1186/s12987-019-0123-z.
- [7] Aird WC. Phenotypic heterogeneity of the endothelium: I. Structure, function, and mechanisms. Circulation Research. 2007; 100: 158–173. https://doi.org/10.1161/01.RES. 0000255691.76142.4a.
- [8] Fisher M. Injuries to the vascular endothelium: vascular wall and endothelial dysfunction. Reviews in Neurological Diseases. 2008; 5: S4–S11.
- [9] Sacco RL, Wolf PA, Kannel WB, McNamara PM. Survival and recurrence following stroke. The Framingham study. Stroke. 1982; 13: 290–295. https://doi.org/10.1161/01.str.13.3.290.
- [10] Ray BK, Hazra A, Ghosal M, Banerjee T, Chaudhuri A, Singh V, *et al.* Early and delayed fatality of stroke in Kolkata, India: results from a 7-year longitudinal population-based study. Journal of Stroke and Cerebrovascular Diseases. 2013; 22: 281–289. https://doi.org/10.1016/j.jstrokecerebrovasdis.2011.09.002.
- [11] Singh RJ, Chen S, Ganesh A, Hill MD. Long-term neurological, vascular, and mortality outcomes after stroke. International Journal of Stroke. 2018; 13: 787–796. https://doi.org/10.1177/ 1747493018798526.
- [12] Yang Y, Torbey MT. Angiogenesis and Blood-Brain Barrier Permeability in Vascular Remodeling after Stroke. Current Neuropharmacology. 2020; 18: 1250–1265. https://doi.org/10.2174/ 1570159X18666200720173316.
- [13] Marti HJ, Bernaudin M, Bellail A, Schoch H, Euler M, Petit E, et al. Hypoxia-induced vascular endothelial growth factor expression precedes neovascularization after cerebral ischemia. The American Journal of Pathology. 2000; 156: 965–976. https://doi.org/10.1016/S0002-9440(10)64964-4.
- [14] Carmeliet P. Mechanisms of angiogenesis and arteriogenesis. Nature Medicine. 2000; 6: 389–395. https://doi.org/10.1038/74651.
- [15] Iadecola C. The pathobiology of vascular dementia. Neuron. 2013; 80: 844–866. https://doi.org/10.1016/j.neuron.2013.10. 008
- [16] Liu J, Wang Y, Akamatsu Y, Lee CC, Stetler RA, Lawton MT, et al. Vascular remodeling after ischemic stroke: mechanisms and therapeutic potentials. Progress in Neurobiology. 2014; 115: 138–156. https://doi.org/10.1016/j.pneurobio.2013.11.004.
- [17] Wiciński M, Puk O, Malinowski B. Cenobamate: Neuropro-

- tective Potential of a New Antiepileptic Drug. Neurochemical Research. 2021; 46: 439–446. https://doi.org/10.1007/s11064-020-03188-8.
- [18] Vargas-Valderrama A, Messina A, Mitjavila-Garcia MT, Guenou H. The endothelium, a key actor in organ development and hPSC-derived organoid vascularization. Journal of Biomedical Science. 2020; 27: 67. https://doi.org/10.1186/s12929-020-00661-y.
- [19] Oldendorf WH, Cornford ME, Brown WJ. The large apparent work capability of the blood-brain barrier: a study of the mitochondrial content of capillary endothelial cells in brain and other tissues of the rat. Annals of Neurology. 1977; 1: 409–417. https://doi.org/10.1002/ana.410010502.
- [20] Sedlakova R, Shivers RR, Del Maestro RF. Ultrastructure of the blood-brain barrier in the rabbit. Journal of Submicroscopic Cytology and Pathology. 1999; 31: 149–161.
- [21] Chan PH. Reactive oxygen radicals in signaling and damage in the ischemic brain. Journal of Cerebral Blood Flow and Metabolism. 2001; 21: 2–14. https://doi.org/10.1097/00004647-200101000-00002.
- [22] Heo JH, Han SW, Lee SK. Free radicals as triggers of brain edema formation after stroke. Free Radical Biology & Medicine. 2005; 39: 51–70. https://doi.org/10.1016/j.freeradbiomed.2005. 03.035.
- [23] Yin KJ, Chen SD, Lee JM, Xu J, Hsu CY. ATM gene regulates oxygen-glucose deprivation-induced nuclear factorkappaB DNA-binding activity and downstream apoptotic cascade in mouse cerebrovascular endothelial cells. Stroke. 2002; 33: 2471–2477. https://doi.org/10.1161/01.str.0000030316. 79601.03.
- [24] Kimura H, Gules I, Meguro T, Zhang JH. Cytotoxicity of cytokines in cerebral microvascular endothelial cell. Brain Research. 2003; 990: 148–156. https://doi.org/10.1016/ s0006-8993(03)03450-4.
- [25] van Sorge NM, Ebrahimi CM, McGillivray SM, Quach D, Sabet M, Guiney DG, et al. Anthrax toxins inhibit neutrophil signaling pathways in brain endothelium and contribute to the pathogenesis of meningitis. PLoS ONE. 2008; 3: e2964. https://doi.org/10.1371/journal.pone.0002964.
- [26] Shen S, Shao Y, Li C. Different types of cell death and their shift in shaping disease. Cell Death Discovery. 2023; 9: 284. https://doi.org/10.1038/s41420-023-01581-0.
- [27] Dang X, Huan X, Du X, Chen X, Bi M, Yan C, et al. Correlation of Ferroptosis and Other Types of Cell Death in Neurodegenerative Diseases. Neuroscience Bulletin. 2022; 38: 938–952. https://doi.org/10.1007/s12264-022-00861-6.
- [28] Ye S, Cheng Z, Zhuo D, Liu S. Different Types of Cell Death in Diabetic Neuropathy: A Focus on Mechanisms and Therapeutic Strategies. International Journal of Molecular Sciences. 2024; 25: 8126. https://doi.org/10.3390/ijms25158126.
- [29] Galluzzi L, Vitale I, Aaronson SA, Abrams JM, Adam D, Agostinis P, et al. Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. Cell Death and Differentiation. 2018; 25: 486–541. https://doi.org/10.1038/s41418-017-0012-4.
- [30] Green DR. Apoptotic pathways: paper wraps stone blunts scissors. Cell. 2000; 102: 1–4. https://doi.org/10.1016/ s0092-8674(00)00003-9.
- [31] Wang X. The expanding role of mitochondria in apoptosis. Genes & Development. 2001; 15: 2922–2933.
- [32] Kelly GL, Strasser A. Toward targeting antiapoptotic MCL-1 for cancer therapy. Annual Review of Cancer Biology. 2020; 4: 299–313. https://doi.org/10.1146/annurev-cancerbio-030419-033510.
- [33] Strasser A, Jost PJ, Nagata S. The many roles of FAS receptor signaling in the immune system. Immunity. 2009; 30: 180–192.



- https://doi.org/10.1016/j.immuni.2009.01.001.
- [34] Fulda S, Debatin KM. Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy. Oncogene. 2006; 25: 4798–4811. https://doi.org/10.1038/sj.onc.1209608.
- [35] Meng S, Cao H, Huang Y, Shi Z, Li J, Wang Y, et al. ASK1-K716R reduces neuroinflammation and white matter injury via preserving blood-brain barrier integrity after traumatic brain injury. Journal of Neuroinflammation. 2023; 20: 244. https://doi.org/10.1186/s12974-023-02923-6.
- [36] Xiao B, Chai Y, Lv S, Ye M, Wu M, Xie L, et al. Endothelial cell-derived exosomes protect SH-SY5Y nerve cells against ischemia/reperfusion injury. International Journal of Molecular Medicine. 2017; 40: 1201–1209. https://doi.org/10.3892/ijmm.2017.3106.
- [37] Palm F, Lahdentausta L, Sorsa T, Tervahartiala T, Gokel P, Buggle F, *et al.* Biomarkers of periodontitis and inflammation in ischemic stroke: A case-control study. Innate Immunity. 2014; 20: 511–518. https://doi.org/10.1177/1753425913501214.
- [38] Pussinen PJ, Alfthan G, Jousilahti P, Paju S, Tuomilehto J. Systemic exposure to Porphyromonas gingivalis predicts incident stroke. Atherosclerosis. 2007; 193: 222–228. https://doi.org/10.1016/j.atherosclerosis.2006.06.027.
- [39] Dominy SS, Lynch C, Ermini F, Benedyk M, Marczyk A, Konradi A, et al. Porphyromonas gingivalis in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors. Science Advances. 2019; 5: eaau3333. https://doi.org/10.1126/sciadv.aau3333.
- [40] Singhrao SK, Olsen I. Assessing the role of *Porphyromonas gingivalis* in periodontitis to determine a causative relationship with Alzheimer's disease. Journal of Oral Microbiology. 2019; 11: 1563405. https://doi.org/10.1080/20002297.2018.1563405.
- [41] Charoensaensuk V, Chen YC, Lin YH, Ou KL, Yang LY, Lu DY. *Porphyromonas gingivalis* Induces Proinflammatory Cytokine Expression Leading to Apoptotic Death through the Oxidative Stress/NF-κB Pathway in Brain Endothelial Cells. Cells. 2021; 10: 3033. https://doi.org/10.3390/cells10113033.
- [42] Vanden Berghe T, Linkermann A, Jouan-Lanhouet S, Walczak H, Vandenabeele P. Regulated necrosis: the expanding network of non-apoptotic cell death pathways. Nature Reviews. Molecular Cell Biology. 2014; 15: 135–147. https://doi.org/10.1038/nrm3737.
- [43] Dhuriya YK, Sharma D. Necroptosis: a regulated inflammatory mode of cell death. Journal of Neuroinflammation. 2018; 15: 199. https://doi.org/10.1186/s12974-018-1235-0.
- [44] Taylor RC, Cullen SP, Martin SJ. Apoptosis: controlled demolition at the cellular level. Nature Reviews. Molecular Cell Biology. 2008; 9: 231–241. https://doi.org/10.1038/nrm2312.
- [45] Weinlich R, Oberst A, Beere HM, Green DR. Necroptosis in development, inflammation and disease. Nature Reviews. Molecular Cell Biology. 2017; 18: 127–136. https://doi.org/10.1038/nrm.2016.149.
- [46] Xu D, Jin T, Zhu H, Chen H, Ofengeim D, Zou C, et al. TBK1 Suppresses RIPK1-Driven Apoptosis and Inflammation during Development and in Aging. Cell. 2018; 174: 1477–1491.e19. https://doi.org/10.1016/j.cell.2018.07.041.
- [47] Strilic B, Yang L, Albarrán-Juárez J, Wachsmuth L, Han K, Müller UC, et al. Tumour-cell-induced endothelial cell necroptosis via death receptor 6 promotes metastasis. Nature. 2016; 536: 215–218. https://doi.org/10.1038/nature19076.
- [48] Caccamo A, Branca C, Piras IS, Ferreira E, Huentelman MJ, Liang WS, et al. Necroptosis activation in Alzheimer's disease. Nature Neuroscience. 2017; 20: 1236–1246. https://doi.org/10. 1038/nn.4608.
- [49] Morrice JR, Gregory-Evans CY, Shaw CA. Necroptosis in amyotrophic lateral sclerosis and other neurological disorders. Biochimica et Biophysica Acta. Molecular Basis of Disease.

- 2017; 1863: 347–353. https://doi.org/10.1016/j.bbadis.2016.11.
- [50] Ofengeim D, Ito Y, Najafov A, Zhang Y, Shan B, DeWitt JP, et al. Activation of necroptosis in multiple sclerosis. Cell Reports. 2015; 10: 1836–1849. https://doi.org/10.1016/j.celrep.2015.02.
- [51] Chen AQ, Fang Z, Chen XL, Yang S, Zhou YF, Mao L, et al. Microglia-derived TNF-α mediates endothelial necroptosis aggravating blood brain-barrier disruption after ischemic stroke. Cell Death & Disease. 2019; 10: 487. https://doi.org/10.1038/s41419-019-1716-9.
- [52] Naito MG, Xu D, Amin P, Lee J, Wang H, Li W, et al. Sequential activation of necroptosis and apoptosis cooperates to mediate vascular and neural pathology in stroke. Proceedings of the National Academy of Sciences of the United States of America. 2020; 117: 4959–4970. https://doi.org/10.1073/pnas.1916427117.
- [53] Li Y, Zou C, Chen C, Li S, Zhu Z, Fan Q, et al. Myeloid-derived MIF drives RIPK1-mediated cerebromicrovascular endothelial cell death to exacerbate ischemic brain injury. Proceedings of the National Academy of Sciences of the United States of America. 2023; 120: e2219091120. https://doi.org/10.1073/pnas.2219091120.
- [54] Chen S, Mei S, Luo Y, Wu H, Zhang J, Zhu J. Gasdermin Family: a Promising Therapeutic Target for Stroke. Translational Stroke Research. 2018; 9: 555–563. https://doi.org/10.1007/s12975-018-0666-3.
- [55] Shi J, Gao W, Shao F. Pyroptosis: Gasdermin-Mediated Programmed Necrotic Cell Death. Trends in Biochemical Sciences. 2017; 42: 245–254. https://doi.org/10.1016/j.tibs.2016.10.004.
- [56] Tan MS, Tan L, Jiang T, Zhu XC, Wang HF, Jia CD, et al. Amyloid-β induces NLRP1-dependent neuronal pyroptosis in models of Alzheimer's disease. Cell Death & Disease. 2014; 5: e1382. https://doi.org/10.1038/cddis.2014.348.
- [57] Dinarello CA. A clinical perspective of IL-1β as the gatekeeper of inflammation. European Journal of Immunology. 2011; 41: 1203–1217. https://doi.org/10.1002/eji.201141550.
- [58] Xu YJ, Zheng L, Hu YW, Wang Q. Pyroptosis and its relationship to atherosclerosis. Clinica Chimica Acta; International Journal of Clinical Chemistry. 2018; 476: 28–37. https://doi.org/10.1016/j.cca.2017.11.005.
- [59] Cheng KT, Xiong S, Ye Z, Hong Z, Di A, Tsang KM, et al. Caspase-11-mediated endothelial pyroptosis underlies endotoxemia-induced lung injury. The Journal of Clinical Investigation. 2017; 127: 4124–4135. https://doi.org/10.1172/JC 194495.
- [60] Wang Y, Wu J, Wang J, He L, Lai H, Zhang T, et al. Mitochondrial oxidative stress in brain microvascular endothelial cells: Triggering blood-brain barrier disruption. Mitochondrion. 2023; 69: 71–82. https://doi.org/10.1016/j.mito.2023.01.007.
- [61] Thornton C, Leaw B, Mallard C, Nair S, Jinnai M, Hagberg H. Cell Death in the Developing Brain after Hypoxia-Ischemia. Frontiers in Cellular Neuroscience. 2017; 11: 248. https://doi.org/10.3389/fncel.2017.00248.
- [62] Wei C, Jiang W, Wang R, Zhong H, He H, Gao X, et al. Brain endothelial GSDMD activation mediates inflammatory BBB breakdown. Nature. 2024; 629: 893–900. https://doi.org/10.1038/s41586-024-07314-2.
- [63] Coll RC, Robertson AAB, Chae JJ, Higgins SC, Muñoz-Planillo R, Inserra MC, et al. A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases. Nature Medicine. 2015; 21: 248–255. https://doi.org/10.1038/nm .3806.
- [64] Han C, Zhai L, Shen H, Wang J, Guan Q. Advanced Glycation End-Products (AGEs) Promote Endothelial Cell Pyroptosis Under Cerebral Ischemia and Hypoxia via HIF-1α-RAGE-



- NLRP3. Molecular Neurobiology. 2023; 60: 2355–2366. https://doi.org/10.1007/s12035-023-03228-8.
- [65] Wang Y, Guan X, Gao CL, Ruan W, Zhao S, Kai G, et al. Medioresinol as a novel PGC-1α activator prevents pyroptosis of endothelial cells in ischemic stroke through PPARα-GOT1 axis. Pharmacological Research. 2021; 169: 105640. https://doi.org/10.1016/j.phrs.2021.105640.
- [66] Guo Y, Yang JH, He Y, Zhou HF, Wang Y, Ding ZS, et al. Protocatechuic aldehyde prevents ischemic injury by attenuating brain microvascular endothelial cell pyroptosis via lncRNA Xist. Phytomedicine: International Journal of Phytotherapy and Phytopharmacology. 2022; 94: 153849. https://doi.org/10.1016/j.phymed.2021.153849.
- [67] Weiland A, Wang Y, Wu W, Lan X, Han X, Li Q, et al. Ferroptosis and Its Role in Diverse Brain Diseases. Molecular Neurobiology. 2019; 56: 4880–4893. https://doi.org/10.1007/s12035-018-1403-3.
- [68] Su L, Jiang X, Yang C, Zhang J, Chen B, Li Y, et al. Pannexin 1 mediates ferroptosis that contributes to renal ischemia/reperfusion injury. The Journal of Biological Chemistry. 2019; 294: 19395–19404. https://doi.org/10.1074/jbc.RA119. 010949.
- [69] Cheng Y, Song Y, Chen H, Li Q, Gao Y, Lu G, et al. Ferroptosis Mediated by Lipid Reactive Oxygen Species: A Possible Causal Link of Neuroinflammation to Neurological Disorders. Oxidative Medicine and Cellular Longevity. 2021; 2021: 5005136. https://doi.org/10.1155/2021/5005136.
- [70] Chen B, Chen Z, Liu M, Gao X, Cheng Y, Wei Y, et al. Inhibition of neuronal ferroptosis in the acute phase of intracerebral hemorrhage shows long-term cerebroprotective effects. Brain Research Bulletin. 2019; 153: 122–132. https://doi.org/10.1016/j. brainresbull.2019.08.013.
- [71] Li Q, Weiland A, Chen X, Lan X, Han X, Durham F, *et al.* Corrigendum: Ultrastructural characteristics of neuronal death and white matter injury in mouse brain tissues after intracerebral hemorrhage: coexistence of ferroptosis, autophagy, and necrosis. Frontiers in Neurology. 2024; 15: 1385719. https://doi.org/10.3389/fneur.2024.1385719.
- [72] Liu Q, Song T, Chen B, Zhang J, Li W. Ferroptosis of brain microvascular endothelial cells contributes to hypoxia-induced blood-brain barrier injury. FASEB Journal. 2023; 37: e22874. https://doi.org/10.1096/fj.202201765R.
- [73] Liu Q, Song T, Chen B, Zhang J, Li W. Ferroptosis of brain microvascular endothelial cells contributes to hypoxia-induced blood-brain barrier injury. FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology. 2023; 37: e22874. https://doi.org/10.1096/fj.202201765R.
- [74] Donovan A, Brownlie A, Zhou Y, Shepard J, Pratt SJ, Moynihan J, *et al.* Positional cloning of zebrafish ferroportin1 identifies a conserved vertebrate iron exporter. Nature. 2000; 403: 776–781. https://doi.org/10.1038/35001596.
- [75] Zheng H, Guo X, Kang S, Li Z, Tian T, Li J, et al. Cdh5-mediated Fpn1 deletion exerts neuroprotective effects during the acute phase and inhibitory effects during the recovery phase of ischemic stroke. Cell Death & Disease. 2023; 14: 161. https://doi.org/10.1038/s41419-023-05688-1.
- [76] Chang LC, Chiang SK, Chen SE, Yu YL, Chou RH, Chang WC. Heme oxygenase-1 mediates BAY 11-7085 induced ferroptosis. Cancer Letters. 2018; 416: 124–137. https://doi.org/10.1016/j.canlet.2017.12.025.
- [77] Tang Z, Ju Y, Dai X, Ni N, Liu Y, Zhang D, et al. HO-1-mediated ferroptosis as a target for protection against retinal pigment epithelium degeneration. Redox Biology. 2021; 43: 101971. https://doi.org/10.1016/j.redox.2021.101971.
- [78] Ozen M, Kitase Y, Vasan V, Burkhardt C, Ramachandra S, Robinson S, et al. Chorioamnionitis Precipitates Perinatal Al-

- terations of Heme-Oxygenase-1 (HO-1) Homeostasis in the Developing Rat Brain. International Journal of Molecular Sciences. 2021; 22: 5773. https://doi.org/10.3390/ijms22115773.
- [79] Zhao J, Zhao X, Tian J, Xue R, Luo B, Lv J, et al. Theanine attenuates hippocampus damage of rat cerebral ischemia-reperfusion injury by inhibiting HO-1 expression and activating ERK1/2 pathway. Life Sciences. 2020; 241: 117160. https://doi.org/10.1016/j.lfs.2019.117160.
- [80] Liu C, Tian Q, Wang J, He P, Han S, Guo Y, et al. Correction to: Blocking P2RX7 Attenuates Ferroptosis in Endothelium and Reduces HG-induced Hemorrhagic Transformation After MCAO by Inhibiting ERK1/2 and P53 Signaling Pathways. Molecular Neurobiology. 2023; 60: 480. https://doi.org/10.1007/s12035-022-03138-1.
- [81] He C, Klionsky DJ. Regulation mechanisms and signaling pathways of autophagy. Annual Review of Genetics. 2009; 43: 67– 93. https://doi.org/10.1146/annurev-genet-102808-114910.
- [82] Wang P, Shao BZ, Deng Z, Chen S, Yue Z, Miao CY. Autophagy in ischemic stroke. Progress in Neurobiology. 2018; 163-164: 98-117. https://doi.org/10.1016/j.pneurobio.2018.01.001.
- [83] van Vliet EA, Forte G, Holtman L, den Burger JC, Sinjewel A, de Vries HE, *et al*. Inhibition of mammalian target of rapamycin reduces epileptogenesis and blood-brain barrier leakage but not microglia activation. Epilepsia. 2012; 53: 1254–1263.
- [84] Fang L, Li X, Zhong Y, Yu J, Yu L, Dai H, et al. Autophagy protects human brain microvascular endothelial cells against methylglyoxal-induced injuries, reproducible in a cerebral ischemic model in diabetic rats. Journal of Neurochemistry. 2015; 135: 431–440. https://doi.org/10.1111/jnc.13277.
- [85] Yang Z, Huang C, Wen X, Liu W, Huang X, Li Y, et al. Circular RNA circ-FoxO3 attenuates blood-brain barrier damage by inducing autophagy during ischemia/reperfusion. Molecular Therapy. 2022; 30: 1275–1287. https://doi.org/10.1016/j.ymthe.2021.11.004.
- [86] Ryter SW, Cloonan SM, Choi AMK. Autophagy: a critical regulator of cellular metabolism and homeostasis. Molecules and Cells. 2013; 36: 7–16. https://doi.org/10.1007/s10059-013-0140-8.
- [87] Liu J, Weaver J, Jin X, Zhang Y, Xu J, Liu KJ, et al. Nitric Oxide Interacts with Caveolin-1 to Facilitate Autophagy-Lysosome-Mediated Claudin-5 Degradation in Oxygen-Glucose Deprivation-Treated Endothelial Cells. Molecular Neurobiology. 2016; 53: 5935–5947. https://doi.org/10.1007/s12035-015-9504-8.
- [88] Wei N, Yu SP, Gu XH, Chen DD, Whalin MK, Xu GL, et al. The involvement of autophagy pathway in exaggerated ischemic brain damage in diabetic mice. CNS Neuroscience & Therapeutics. 2013; 19: 753–763. https://doi.org/10.1111/cns.12123.
- [89] Li H, Gao A, Feng D, Wang Y, Zhang L, Cui Y, et al. Evaluation of the protective potential of brain microvascular endothelial cell autophagy on blood-brain barrier integrity during experimental cerebral ischemia-reperfusion injury. Translational Stroke Research. 2014; 5: 618–626. https://doi.org/10.1007/s12975-014-0354-x.
- [90] Fatokun AA, Dawson VL, Dawson TM. Parthanatos: mitochondrial-linked mechanisms and therapeutic opportunities. British Journal of Pharmacology. 2014; 171: 2000–2016. https://doi.org/10.1111/bph.12416.
- [91] Robinson N, Ganesan R, Hegedűs C, Kovács K, Kufer TA, Virág L. Programmed necrotic cell death of macrophages: Focus on pyroptosis, necroptosis, and parthanatos. Redox Biology. 2019; 26: 101239. https://doi.org/10.1016/j.redox.2019.101239.
- [92] Andrabi SA, Dawson TM, Dawson VL. Mitochondrial and nuclear cross talk in cell death: parthanatos. Annals of the New York Academy of Sciences. 2008; 1147: 233–241. https://doi.org/10.1196/annals.1427.014.



- [93] Wang Y, An R, Umanah GK, Park H, Nambiar K, Eacker SM, et al. A nuclease that mediates cell death induced by DNA damage and poly(ADP-ribose) polymerase-1. Science (New York, N.Y.). 2016; 354: aad6872. https://doi.org/10.1126/science.aad6872.
- [94] Liu L, Li J, Ke Y, Zeng X, Gao J, Ba X, et al. The key players of parthanatos: opportunities for targeting multiple levels in the therapy of parthanatos-based pathogenesis. Cellular and Molecular Life Sciences. 2022; 79: 60. https://doi.org/10.1007/s00018-021-04109-w.
- [95] Klionsky DJ. Autophagy: from phenomenology to molecular understanding in less than a decade. Nature Reviews. Molecular Cell Biology. 2007; 8: 931–937. https://doi.org/10.1038/nr m2245.
- [96] Dikic I, Elazar Z. Mechanism and medical implications of mammalian autophagy. Nature Reviews. Molecular Cell Biology. 2018; 19: 349–364. https://doi.org/10.1038/s41580-018-0003-4.
- [97] Wang F, Gómez-Sintes R, Boya P. Lysosomal membrane permeabilization and cell death. Traffic. 2018; 19: 918–931. https://doi.org/10.1111/tra.12613.
- [98] Sun Y, Li T, Xie C, Zhang Y, Zhou K, Wang X, et al. Dichloroacetate treatment improves mitochondrial metabolism and reduces brain injury in neonatal mice. Oncotarget. 2016; 7: 31708–31722. https://doi.org/10.18632/oncotarget.9150.
- [99] Zille M, Ikhsan M, Jiang Y, Lampe J, Wenzel J, Schwaninger M. The impact of endothelial cell death in the brain and its role after stroke: A systematic review. Cell Stress. 2019; 3: 330–347. https://doi.org/10.15698/cst2019.11.203.
- [100] Qin C, Zhou LQ, Ma XT, Hu ZW, Yang S, Chen M, et al. Dual Functions of Microglia in Ischemic Stroke. Neuroscience Bulletin. 2019; 35: 921–933. https://doi.org/10.1007/s12264-019-00388-3.
- [101] Wang H, Hong LJ, Huang JY, Jiang Q, Tao RR, Tan C, et al. P2RX7 sensitizes Mac-1/ICAM-1-dependent leukocyte-endothelial adhesion and promotes neurovascular injury during septic encephalopathy. Cell Research. 2015; 25: 674–690. https://doi.org/10.1038/cr.2015.61.
- [102] Liang KJ, Lee JE, Wang YD, Ma W, Fontainhas AM, Fariss RN, et al. Regulation of dynamic behavior of retinal microglia by CX3CR1 signaling. Investigative Ophthalmology & Visual Science. 2009; 50: 4444–4451. https://doi.org/10.1167/iovs .08-3357.
- [103] Nikolakopoulou AM, Montagne A, Kisler K, Dai Z, Wang Y, Huuskonen MT, et al. Pericyte loss leads to circulatory failure and pleiotrophin depletion causing neuron loss. Nature Neuroscience. 2019; 22: 1089–1098. https://doi.org/10.1038/s41593-019-0434-z.
- [104] Sweeney MD, Sagare AP, Zlokovic BV. Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. Nature Reviews. Neurology. 2018; 14: 133–150. https://doi.org/10.1038/nrneurol.2017.188.
- [105] Yang S, Jin H, Zhu Y, Wan Y, Opoku EN, Zhu L, et al. Diverse Functions and Mechanisms of Pericytes in Ischemic Stroke. Current Neuropharmacology. 2017; 15: 892–905. https://doi.org/ 10.2174/1570159X15666170112170226.
- [106] Lusk JB, Xu H, Peterson ED, Bhatt DL, Fonarow GC, Smith EE, et al. Antithrombotic Therapy for Stroke Prevention in Patients With Ischemic Stroke With Aspirin Treatment Failure. Stroke. 2021; 52: e777–e781. https://doi.org/10.1161/STROKE AHA.121.034622.
- [107] Tanswell P, Modi N, Combs D, Danays T. Pharmacokinetics and pharmacodynamics of tenecteplase in fibrinolytic therapy of acute myocardial infarction. Clinical Pharmacokinetics. 2002; 41: 1229–1245. https://doi.org/10.2165/00003088-200241150-00001.
- [108] Goyal M, Menon BK, van Zwam WH, Dippel DWJ, Mitchell

- PJ, Demchuk AM, *et al.* Endovascular thrombectomy after large-vessel ischaemic stroke: a meta-analysis of individual patient data from five randomised trials. Lancet (London, England). 2016; 387: 1723–1731. https://doi.org/10.1016/S0140-6736(16)00163-X.
- [109] Zhao X, Li S, Mo Y, Li R, Huang S, Zhang A, et al. DCA Protects against Oxidation Injury Attributed to Cerebral Ischemia-Reperfusion by Regulating Glycolysis through PDK2-PDH-Nrf2 Axis. Oxidative Medicine and Cellular Longevity. 2021; 2021: 5173035. https://doi.org/10.1155/2021/5173035.
- [110] Schroder K, Tschopp J. The inflammasomes. Cell. 2010; 140: 821–832. https://doi.org/10.1016/j.cell.2010.01.040.
- [111] Cheah IK, Halliwell B. Ergothioneine; antioxidant potential, physiological function and role in disease. Biochimica et Biophysica Acta. 2012; 1822: 784–793. https://doi.org/10.1016/j. bbadis.2011.09.017.
- [112] Leow DMK, Cheah IKM, Fong ZWJ, Halliwell B, Ong WY. Protective Effect of Ergothioneine against 7-Ketocholesterol-Induced Mitochondrial Damage in hCMEC/D3 Human Brain Endothelial Cells. International Journal of Molecular Sciences. 2023; 24: 5498. https://doi.org/10.3390/ijms24065498.
- [113] Qi W, Boliang W, Xiaoxi T, Guoqiang F, Jianbo X, Gang W. Cardamonin protects against doxorubicin-induced cardiotoxicity in mice by restraining oxidative stress and inflammation associated with Nrf2 signaling. Biomedicine & Pharmacotherapy. 2020; 122: 109547. https://doi.org/10.1016/j.biopha.2019. 109547.
- [114] Wang K, Lv Q, Miao YM, Qiao SM, Dai Y, Wei ZF. Cardamonin, a natural flavone, alleviates inflammatory bowel disease by the inhibition of NLRP3 inflammasome activation via an AhR/Nrf2/NQO1 pathway. Biochemical Pharmacology. 2018; 155: 494–509. https://doi.org/10.1016/j.bcp.2018.07.039.
- [115] Liu EYL, Xu ML, Xia Y, Kong X, Wu Q, Dong TTX, et al. Activation of G protein-coupled receptor 30 by flavonoids leads to expression of acetylcholinesterase in cultured PC12 cells. Chemico-biological Interactions. 2019; 306: 147–151. https://doi.org/10.1016/j.cbi.2019.04.031.
- [116] Ni H, Li J, Zheng J, Zhou B. Cardamonin attenuates cerebral ischemia/reperfusion injury by activating the HIF-1α/VEGFA pathway. Phytotherapy Research. 2022; 36: 1736–1747. https://doi.org/10.1002/ptr.7409.
- [117] Huang J, Zhang X, Yang X, Yv Q, Ye F, Chen S, et al. Baicalin exerts neuroprotective actions by regulating the Nrf2-NLRP3 axis in toxin-induced models of Parkinson's disease. Chemicobiological Interactions. 2024; 387: 110820. https://doi.org/10. 1016/j.cbi.2023.110820.
- [118] Martínez Leo EE, Segura Campos MR. Systemic Oxidative Stress: A key Point in Neurodegeneration - A Review. The Journal of Nutrition, Health & Aging. 2019; 23: 694–699. https: //doi.org/10.1007/s12603-019-1240-8.
- [119] Wang X, Yu JY, Sun Y, Wang H, Shan H, Wang S. Baicalin protects LPS-induced blood-brain barrier damage and activates Nrf2-mediated antioxidant stress pathway. International Immunopharmacology. 2021; 96: 107725. https://doi.org/10.1016/ j.intimp.2021.107725.
- [120] Zhang X, Wu J, Zhang B. Xuesaitong injection as one adjuvant treatment of acute cerebral infarction: a systematic review and meta-analysis. BMC Complementary and Alternative Medicine. 2015; 15: 36. https://doi.org/10.1186/s12906-015-0560-4.
- [121] Yang X, Xiong X, Wang H, Wang J. Protective effects of panax notoginseng saponins on cardiovascular diseases: a comprehensive overview of experimental studies. Evidence-Based Complementary and Alternative Medicine. 2014; 2014: 204840. https://doi.org/10.1155/2014/204840.
- [122] Li J, Lu K, Sun F, Tan S, Zhang X, Sheng W, et al.

 Panaxydol attenuates ferroptosis against LPS-induced acute



- lung injury in mice by Keap1-Nrf2/HO-1 pathway. Journal of Translational Medicine. 2021; 19: 96. https://doi.org/10.1186/s12967-021-02745-1.
- [123] Li SN, Li P, Liu WH, Shang JJ, Qiu SL, Zhou MX, et al. Danhong injection enhances angiogenesis after myocardial infarction by activating MiR-126/ERK/VEGF pathway. Biomedicine & Pharmacotherapy. 2019; 120: 109538. https://doi.org/10.1016/j.biopha.2019.109538.
- [124] Kong BS, Im SJ, Lee YJ, Cho YH, Do YR, Byun JW, et al. Vasculoprotective Effects of 3-Hydroxybenzaldehyde against VSMCs Proliferation and ECs Inflammation. PloS One. 2016; 11: e0149394. https://doi.org/10.1371/journal.pone.0149394.
- [125] Darendelioglu E. Caffeic acid suppresses HT-29 cell death induced by H2O2 via oxidative stress and apoptosis. Bratislavske Lekarske Listy. 2020; 121: 805–811. https://doi.org/10.4149/BLL 2020 132.
- [126] Yang L, Tao Y, Luo L, Zhang Y, Wang X, Meng X. Dengzhan Xixin injection derived from a traditional Chinese herb Erigeron breviscapus ameliorates cerebral ischemia/reperfusion injury in rats via modulation of mitophagy and mitochondrial apoptosis. Journal of Ethnopharmacology. 2022; 288: 114988. https://doi. org/10.1016/j.jep.2022.114988.
- [127] Li XN, Shang NY, Kang YY, Sheng N, Lan JQ, Tang JS, et al. Caffeic acid alleviates cerebral ischemic injury in rats by resisting ferroptosis via Nrf2 signaling pathway. Acta Pharmacologica Sinica. 2024; 45: 248–267. https://doi.org/10.1038/s41401-023-01177-5.
- [128] Finkelstein DI, Billings JL, Adlard PA, Ayton S, Sedjahtera A, Masters CL, et al. Correction to: The novel compound PBT434 prevents iron mediated neurodegeneration and alpha-synuclein toxicity in multiple models of Parkinson's disease. Acta Neuropathologica Communications. 2021; 9: 161. https://doi.org/

- 10.1186/s40478-021-01258-8.
- [129] Bailey DK, Clark W, Kosman DJ. The iron chelator, PBT434, modulates transcellular iron trafficking in brain microvascular endothelial cells. PLoS ONE. 2021; 16: e0254794. https://doi. org/10.1371/journal.pone.0254794.
- [130] Zhu T, Yao Y, Ding J, Zhang C, Xia N, Tao Y, et al. 3-Methyladenine attenuates neuroinflammation and improves cognitive function in sepsis-associated encephalopathy by inhibiting autophagy. International Immunopharmacology. 2024; 139: 112744. https://doi.org/10.1016/j.intimp.2024.112744.
- [131] Kim KA, Kim D, Kim JH, Shin YJ, Kim ES, Akram M, et al. Autophagy-mediated occludin degradation contributes to blood-brain barrier disruption during ischemia in bEnd.3 brain endothelial cells and rat ischemic stroke models. Fluids and Barriers of the CNS. 2020; 17: 21. https://doi.org/10.1186/s12987-020-00182-8.
- [132] Liu YQ, Yang QX, Cheng MC, Xiao HB. Synergistic inhibitory effect of Icariside II with Icaritin from Herba Epimedii on preosteoclastic RAW264.7 cell growth. Phytomedicine: International Journal of Phytotherapy and Phytopharmacology. 2014; 21: 1633–1637. https://doi.org/10.1016/j.phymed.2014.07.016.
- [133] Gao J, Long L, Xu F, Feng L, Liu Y, Shi J, *et al.* Icariside II, a phosphodiesterase 5 inhibitor, attenuates cerebral ischaemia/reperfusion injury by inhibiting glycogen synthase kinase-3β-mediated activation of autophagy. British Journal of Pharmacology. 2020; 177: 1434–1452. https://doi.org/10.1111/bph.14912.
- [134] Zhang Y, Huang N, Lu H, Huang J, Jin H, Shi J, *et al.* Icariin protects against sodium azide-induced neurotoxicity by activating the PI3K/Akt/GSK-3β signaling pathway. PeerJ. 2020; 8: e8955. https://doi.org/10.7717/peerj.8955.

