

Review

The Potential Role of Neurogranin in Alzheimer's Disease

Xin Zhou¹ , Xiao-jun Jing¹ , Hua Zhang^{1,*} 

¹Department of Neurology, The First Affiliated Hospital of Chongqing Medical University, 400042 Chongqing, China

*Correspondence: zhanghuapro@hospital.cqmu.edu.cn (Hua Zhang)

Academic Editor: Ulises Gomez-Pinedo

Submitted: 25 June 2024 Revised: 29 August 2024 Accepted: 23 September 2024 Published: 20 March 2025

Abstract

Alzheimer's disease (AD) is the most common form of dementia and is characterized by the excessive deposition of amyloid- β (A β) plaques and the formation of neurofibrillary tangles. Numerous new studies also indicate that synaptic damage and loss play crucial roles in AD and form the basis of cognitive impairment. In recent years, synaptic-related proteins have emerged as important biomarkers for the early diagnosis of AD. Among these proteins, neurogranin (Ng), a postsynaptic protein widely present in the dendritic spines of the associative cortex in the brain, plays a significant role in memory, learning, synaptic plasticity, and long-term potentiation (LTP). This review aims to reveal the link between Ng and AD, as well as the potential for the diagnosis of AD, the prediction of the development of the disease, and the identification of a therapeutic target for AD.

Keywords: Alzheimer's disease; synapse; neurogranin; biomarkers; cognitive function

1. Introduction

Alzheimer's disease (AD) and related dementias affect approximately 50 million people worldwide [1]. AD is one of the most prevalent neurodegenerative disorders globally and is characterized by manifestations of memory loss and cognitive impairment. The pathological hallmarks of this disease include the accumulation of extracellular amyloid- β (A β) plaques, the development of neurofibrillary tangles composed of phosphorylated tau (P tau) protein within cells, synaptic loss, and neuroinflammation [2,3]. AD follows gradual clinical progression and is typically categorized into three stages: early, middle, and late. Dementia represents the final stage of the disease. However, in the early stages of AD, various pathophysiological processes occur, including synaptic dysfunction [4,5], neuronal and axonal damage, neuroinflammation, and glial reactions [6]. Notably, synaptic dysfunction in AD manifests several years before the onset of cognitive decline, underscoring the utility of cerebrospinal fluid (CSF) biomarkers for early AD diagnosis and differential diagnosis from other forms of dementia [7,8].

1.1 Synaptic Damage and Loss Play Crucial Roles in the Progression of AD

Synapses are important components of neurons and essential structures for neural network connections, and there is a fundamental tenet in neuroscience in which synaptic function is fundamental to cognition. The hypothesis that synaptic damage or loss serves as an objective manifestation of neurodegenerative changes, most relevant to the decline in cognitive abilities observed in AD, is widely accepted. This concept is supported by clinical, postmortem, and nonclinical evidence [9]. The study indicates a reduc-

tion in cortical synaptic density of 25% to 30% in AD patients, with the synaptic density per neuron decreasing by 15% to 35% [10]. The idea that synaptic changes modulate information storage gained popularity in the mid-20th century with Hebb's postulation that synapses between neurons that are simultaneously active undergo strengthening, contributing to the learning process [9]. The discovery of long-term potentiation (LTP) by Bliss and Lomo [11], as well as the plasticity of hippocampal synapses in memory formation by Takeuchi and colleagues [12], further emphasized this concept.

Synaptic dysfunction and loss are closely associated with the pathological cognitive decline experienced in AD [13–15]. Most studies to date indicate that mechanisms such as amyloidosis, neuroinflammation, and oxidative stress can lead to synaptic injury, suggesting that synaptic loss may be a downstream effect of amyloidosis, neuroinflammation, oxidative stress, and other mechanisms in AD [9,16]. Biomarkers of synaptic damage reflect the consequences of disease-induced synaptic injury and loss in the brain. The injury and loss of synapses mirror the cumulative effects of various pathological substrates in AD, making them potentially optimal surrogate indicators of AD progression in clinical and radiological contexts [8,17].

1.2 Synaptic Proteins Associated with AD

Many researchers have applied proteomic analysis to identify synaptic biomarkers for AD. Numerous studies have identified various potential synaptic biomarkers, including growth-associated protein 43 (GAP-43), neurogranin (Ng), synaptosome-associated protein of 25 kDa (SNAP-25), synaptoagmin-1, neuronal pentraxin 2 (NPTX2), neurexins, and synaptic vesicle glycoprotein 2A (SV2A) [18,19].



A recent study revealed that AD patients had higher concentrations of 14 synaptic proteins than non-AD participants did. Compared with other neurodegenerative diseases, AD patients presented with particularly elevated levels of synaptic proteins, with SNAP-25, 14-3-3 zeta/delta, β -synuclein, and neuronal granule proteins being especially effective in distinguishing between AD and control groups as well as non-AD individuals [20]. Furthermore, an additional study investigating the impact of AD on three types of synaptic proteins revealed significant increases in the levels of postsynaptic density protein 95 (PSD-95), SNAP-25, and Ng. Importantly, no elevated levels of Ng or SNAP-25 were detected in the cerebrospinal fluid of patients with other neurodegenerative disorders. Notably, a strong correlation between SNAP-25 and Ng was identified in AD [21]. These findings are consistent with those of previous studies, indicating that Ng is specifically elevated in AD-related neurodegenerative diseases but not in other neurodegenerative diseases or patients with nonneurodegenerative cognitive impairment [22,23].

Neurogranin is a postsynaptic protein found in the dendritic spines of postsynaptic neurons and consists of a 78-amino acid peptide [24]. It is expressed predominantly in the associative cortex regions of the brain and plays a crucial role in LTP [25]. Among these synaptic-related proteins, Ng is essential for memory, learning, synaptic plasticity, and LTP [26]. It is considered to reflect synaptic degeneration [27,28] and has the potential to predict cognitive decline in AD [29–33]. Ng is believed to be AD specific, as its CSF levels are higher in AD patients than in those with other forms of dementia and neurological disorders [22,23,34,35]. It holds promise as a significant biomarker for the early diagnosis of AD. A variety of synapse-associated proteins are increased or decreased in AD, but most of them are not specifically increased or decreased in AD. Compared with other synapse-associated proteins, Ng is the only postsynaptic protein at present, and these studies have used the specific mechanism of Ng in AD. In addition, the studies mentioned above have also shown that Ng is increased in AD and is specific in AD. Therefore, CSF Ng may be the best CSF biomarker for synaptic loss or dysfunction in AD patients.

Based on current research, synaptic damage represents an early manifestation of AD. Among numerous synaptic proteins, Ng has been extensively investigated as a specific biomarker for AD. To increase our understanding of the precise role played by Ng in AD, this article presents a comprehensive review encompassing Ng's physiological mechanisms, involvement in AD processes, diagnostic efficacy, and potential therapeutic targets. The aim is to provide a thorough understanding of Ng.

2. Biological Effects of Ng

Ng is the most abundant postsynaptic calmodulin binding protein (CaMBP) and is an essential regulator of

LTP and long-term depression (LTD) [36,37]. LTP involves persistent strengthening of synaptic connections between neurons, playing a crucial role in the formation of long-term memories [38]. Impaired brain function is associated with insufficient induction and shorter maintenance of LTP, which is crucial for the generation of new connections between neurons [39]. LTD is induced by low-frequency stimulation (LFS) *in vivo*, resulting in weakened connections between synapses and a continuous decline in synaptic efficacy [40]. In the signalling pathway, Ng binds to calmodulin (CaM) through its intact IQ (AAAKIQASFRGHMARKKIK) [41] motif, limiting the interaction of CaM with other calmodulin-binding proteins (CaMBPs) [36]. Ng can target CaM to the postsynaptic membrane, increasing the sensitivity of CaM to calcium ions through the positioning of CaM. This, in turn, fine-tunes the regulation of LTP and LTD through the modulation of calcium ion/calmodulin-dependent protein kinase II (CaMKII) and the CaM-dependent protein phosphatase calcineurin (CaN) [42,43]. Ng primarily localizes near the postsynaptic density, and this spatial positioning may allow preferential activation of targets, such as CaMKII, which are required for LTP induction [36].

A study indicates that a rapid and relatively large increase in the calcium ion concentration (several micromolar) within a short duration (seconds) activates CaMKII, leading to the induction of LTP [36]. In contrast, a modest increase in calcium activates only CaM, which in turn regulates LTD through the activation of CaN [44]. In its nonphosphorylated state, Ng binds to apo-CaM through its IQ motif, conferring the potential ability to isolate, localize, concentrate, and/or control the availability of this regulatory protein near the synaptic membrane [37,45]. The increase in the synaptic calcium ion concentration also depends on the activation of N-methyl-D-aspartate receptors (NMDARs). Ng recruits Ca^{2+} /CaM signals to the postsynaptic region, where Ng enhances NMDAR-mediated synaptic transmission [36]. In the signalling pathway, protein kinase C (PKC) phosphorylates Ng, reducing its ability to bind to CaM. This allows CaM to activate CaMKII for an extended period, a potentially critical event for LTP [46]. Calcium binding to CaM enables it to activate CaMKII, which phosphorylates alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid glutamate receptors (AMPA), allowing these receptors to move to the synapse. The localization of AMPARs in the membrane provides evidence for LTP [36,41]. Additionally, an increase in local calcium ions can dissociate CaM from Ng [42,47]. Therefore, an intact IQ motif and the ability to bind to CaM are essential for Ng's involvement in LTP. The entire process is summarized in Fig. 1.

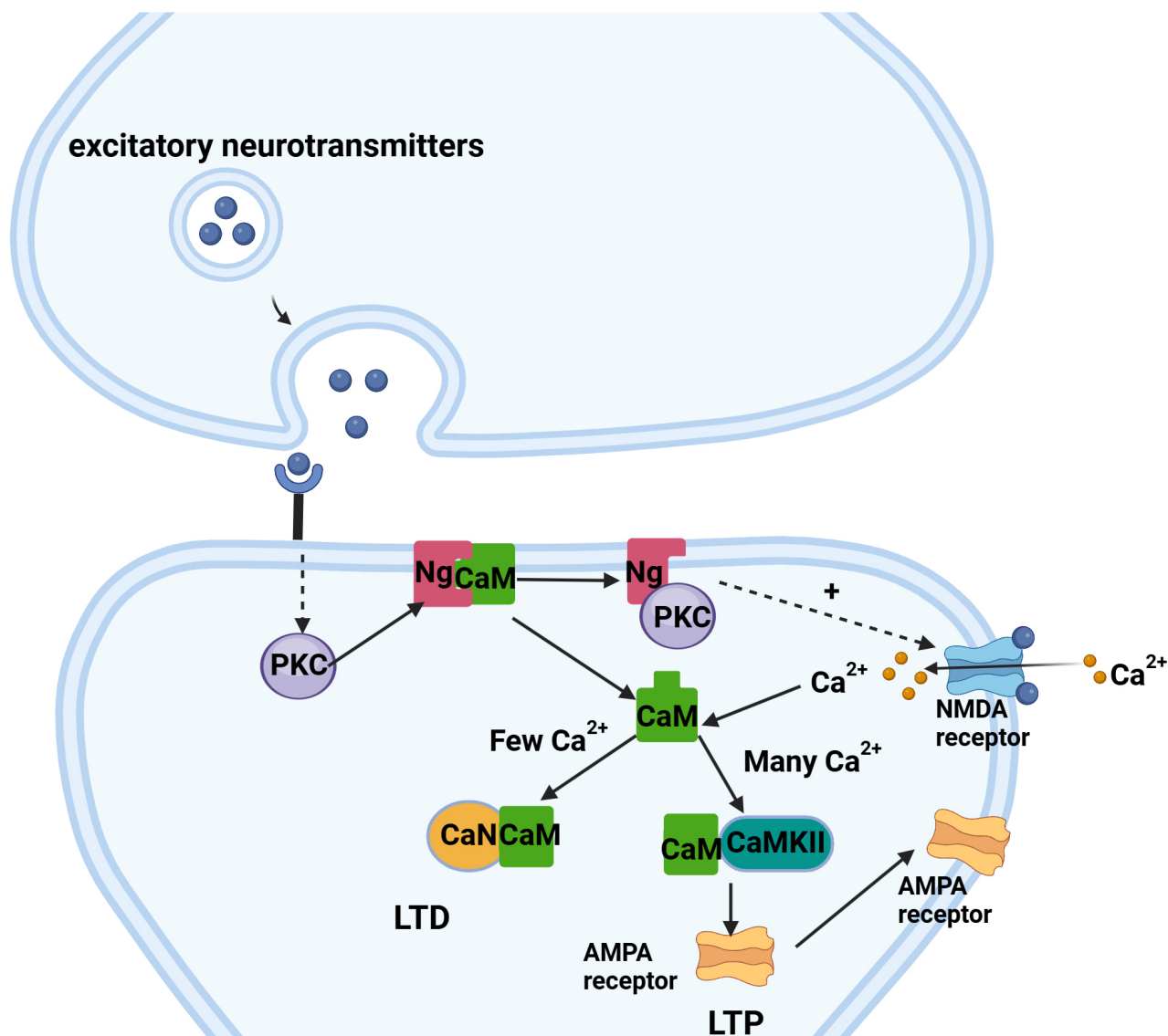


Fig. 1. Ng biological pathways. The binding of calmodulin (CaM) to calmodulin-dependent protein kinase II (CaMKII) in the presence of a large amount of Ca^{2+} activates the alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid glutamate receptor (AMPA) to complete long-term potentiation (LTP), and the binding of CaM to calcineurin (CaN) in the presence of a small amount of Ca^{2+} completes long-term depression (LTD). This figure was drawn using Biorender (<https://www.biorender.com/>). Ng, neurogranin; PKC, protein kinase C; NMDA, N-methyl-D-aspartic acid receptor.

3. Relationship between Ng and the Pathology of AD

In mouse models, the hippocampal Ng concentration decreases with age and is associated with central nervous system functional impairments [48]. Nakajima *et al.* [49] conducted behavioural studies on *Ng* gene knockout mice, revealing phenotypes related to hyperactivity, spatial learning deficits, impaired social skills, motor dysfunction, and altered anxiety levels. In the CaMKII-TetOp25 mouse model (Severe Synaptic loss and Brain atrophy induction model), Ng levels in the CSF increase during neurodegeneration induction, whereas Ng levels in the brain decrease, suggesting that CSF Ng is a biomarker for synaptic degen-

eration [50]. In many postmortem human samples, research has revealed that *Ng* gene (*NRGN*) expression is negatively correlated with amyloid and tau protein pathology in the cortex, as well as with functional clinical dementia rating (CDR) scores, demonstrating an association with the neuropathological diagnosis of AD [51].

3.1 Relationships between Ng and $\text{A}\beta$

The relationship between Ng and $\text{A}\beta$ is manifested primarily through synapses. The $\text{A}\beta$ hypothesis [52] focuses on the accumulation of excessive $\text{A}\beta$, particularly $\text{A}\beta_{42}$, leading to synaptic loss and neuronal death [53]. Substantial evidence indicates that the soluble form of $\text{A}\beta$

is detrimental to synapses [14,54]. The presence of A β oligomers (A β O), particularly in soluble forms, is associated with synaptic plasticity damage and significant loss of synapses in mouse and cell models [55–58] as well as in the human brains of AD patients [59–61]. Additionally, high-resolution imaging techniques reveal the presence of A β O at individual synapses in mouse models and AD cases [59,62,63]. Lambert and colleagues [64] reported that the nonfibrillar form of A β O can inhibit Ng-mediated LTP *in vitro*. Walsh and colleagues [65] also demonstrated that naturally secreted A β O can interfere with LTP *in vivo*.

Recent studies have indicated that, in the preclinical stages of AD when soluble A β biomarkers become positive, there is a sharp increase in tau proteins (p-tau and t-tau) and CSF Ng. This elevation continues throughout the entire preclinical AD process. When CSF p-tau becomes positive, CSF Ng reaches nearly twice the baseline level [52]. Therefore, it can be inferred that changes in soluble A β occur early and that soluble A β may impact alterations in Ng. One study revealed that exogenously added A β 42 predominantly targets the dendritic synapses of CaMKII-positive neurons rather than the axonal synapses [66]. The specific binding of A β 42 to CaMKII-positive synapses may provide an explanation for the selective impact on excitatory neurons.

A β affects synapses in multiple ways, leading to changes in Ng, as follows: (1) A β induces excitotoxic and stress-related signalling pathways and alters the transcriptional regulation of genes critical for neuronal development and plasticity [67–69]. A β oligomers can aggravate glutamate excitotoxicity by damaging glutamate transporters resulting from interruption of glutamate receptors, including NMDARs, AMPARs, and metabotropic glutamate receptors [70]. Excessive activation of NMDARs results in excessive influx of Ca²⁺, triggering the activation of various degrading enzymes, including phospholipase C, CaMKII, PKC, and NO synthase. Consequently, these enzymes induce damage to neuronal lipid membranes and protein scaffolds, thereby disrupting synaptic transmission [71]. The A β peptide interferes with NMDAR and AMPAR receptor transport, resulting in a reduction in surface receptor levels, synaptic dysfunction, and dendritic spine loss [16,40]. These strategies effectively prevent the impairment of Ng-mediated LTP. (2) Additionally, some key proteins in the A β processing pathway have been confirmed to be CaM-binding proteins, and others possess potential CaM-binding domains (CaMBDs) [72,73]. For example, β -secretase 1 (BACE1; β -site amyloid precursor protein cleaving enzyme 1) and amyloid precursor protein (A β PP) can bind to and be regulated by CaM [44]. Ng, being the most abundant CaMBP in the postsynaptic region, may inhibit the binding of CaM to other CaMBPs. There might be a competitive inhibitory effect between Ng and other CaMBPs, thereby influencing synaptic function. These pathways could collectively affect the signalling pathways involving Ng, thus im-

pacting synaptic function. In summary, A β primarily modulates NMDAR and AMPAR receptors to regulate synaptic LTP and LTD, resulting in synaptic damage and an increase in Ng levels in the CSF. However, the direct mechanism by which A β influences Ng remains unclear.

3.2 Relationship between Ng and Tau

Tau is a microtubule-stabilizing protein, and the phosphorylation of tau protein is considered one of the reasons for the formation of neurofibrillary tangles in AD [74]. When tau becomes excessively phosphorylated, it shifts from microtubules, leading to microtubule instability and disruption of neuronal transport mechanisms [75]. CSF t-tau is considered a marker of neurodegeneration, reflecting the intensity of neuronal damage, whereas CSF p-tau is a marker of phosphorylated tau found in neurofibrillary tangles, representing tau pathology. Ng is a neuron-specific postsynaptic protein and a marker of synaptic integrity and function [76]. Synaptic dysfunction is believed to occur before neuronal degeneration and death [77], as evidenced earlier, with an increase in CSF Ng preceding changes in p-tau. Additionally, a study by [78] has indicated a strong correlation between t-tau, p-tau, and Ng, especially in individuals with mild cognitive impairment (MCI) and AD. On the one hand, synaptic loss or impaired function may further affect the transport of microtubules in neurons. On the other hand, A β can also induce excessive phosphorylation of the tau protein, disrupting the stability of microtubule tracks and altering mitochondrial axonal transport and synaptic docking. The translocation of the tau protein to dendritic spines may also have a synergistic effect, leading to the instability of NMDA receptors, increased excitotoxicity, and oxidative stress, thereby adversely affecting synaptic function [69]. A recent study of the genome in AD confirmed five components, with the first component exhibiting a strong association with tau measurements and a moderate association with Ng and Chitinase-3-like protein 1 (YKL-40). This particular component is interpreted as indicative of tau pathology and neurodegeneration [79]. The findings from this study on specific variants suggest a further genetic-level association between tau and Ng compared with wild-type tau expression. Expression of the Tau^{P301L} mutant in cell culture medium was found to increase the levels of Ng, whereas the overall levels of neuronal granule protein and any discernible neurotoxic morphological indicators remained unchanged [80]. The increase in phosphorylated serine at position 262 (pSer262) levels observed with tau^{P301L} may be attributed to the concomitant up-regulation of its association with the kinase CaMKII and downregulation of its association with the phosphatase calcineurin, both of which affect the Ser262 phosphorylation site [81,82]. These findings indicate that excessive expression of pathological tau protein affects its association with specific neuronal binding partners without causing neuronal cell death, resulting in synaptic marker abnormalities.

3.3 Relationships between Ng and Inflammatory Markers Associated with AD

Inflammation is another common feature in the pathology of AD, and an epidemiological study suggests that long-term use of nonsteroidal anti-inflammatory drugs can reduce the risk of AD [2]. Neuroinflammation is a process that primarily involves microglia and astrocytes and is essential for healthy brain function [83]. In the context of AD pathology, neuroinflammation is driven mainly by activated microglia, which stimulate astrocytes, triggering a cascade of inflammatory responses that ultimately lead to synaptic loss and neuronal death [84,85]. One of the most extensively studied inflammatory factors is chitinase-3-like protein 1 (CHI3L1), a protein primarily expressed by astrocytes in the brain and known as YKL-40 in humans [83]. Several studies have reported elevated levels of YKL-40 in AD patients [34,83,86]. While Hellwig *et al.* [34] reported no correlation between YKL-40 and Ng in AD patients, a recent study by Connolly *et al.* [83] suggested a correlation between Ng and YKL-40 in AD patients. In the brain, YKL-40 levels in the cerebrospinal fluid are elevated in several central nervous system (CNS) infectious and noninfectious diseases such as stroke, viral encephalitis, traumatic brain injury, amyotrophic lateral sclerosis, multiple sclerosis, and AD [87–89]. The area under the curve (AUC) of YKL-40 as a biomarker for AD is 0.66, indicating that YKL-40 is a relatively nonspecific marker that is strongly influenced by patient comorbidities [34]. This may also explain why there are some contradictory data regarding the correlation with YKL-40. Further research is needed to understand the relationship between YKL-40 and Ng.

Multiple studies have consistently demonstrated the involvement of CHI3L1 in amyloid plaque formation, phosphorylated tau protein accumulation, and alterations in synaptic plasticity [90–92]. However, in addition to AD, the expression of CHI3L1 is also linked to other neurodegenerative disorders [93–95]. Recent research has revealed that a key regulator of the inflammatory state of the CNS, C-X3-C motif chemokine ligand 1 (CX3CL1), is significantly elevated in AD patients. The levels of Ng are also significantly correlated with both AD and MCI patients [83]. This may be attributed to the importance of CX3CL1/C-X3-C motif chemokine receptor 1 (CX3CR1) signalling in microglia for synaptic plasticity and neural regulation. The significant correlation between CX3CL1 and Ng in AD may be because CX3CL1 can inhibit the maintenance of LTP, reduce spontaneous glutamate release and postsynaptic glutamate currents, and promote the decay of hippocampal synaptic plasticity. It can also inhibit glutamate-induced calcium influx, especially in hippocampal neurons [96–98].

4. Relationship between Ng Hydrolysis and AD

Relevant study indicate that Ng hydrolysis may be associated with AD. Compared with that in the control group,

the ratio of peptide segments to full-length Ng increases in the parietal and temporal cortices of AD patients [99], suggesting that protein hydrolysis of full-length Ng occurs during synaptic degeneration. Mass spectrometry detection in CSF from AD patients revealed that endogenous Ng fragments represent approximately half of the C-terminus, and all fragments lack the IQ structural domain except for one [78]. Moreover, a study suggests that C-terminal fragments are not generated through extracellular protein degradation in CSF but are released from degenerating synapses in the CNS during the progression of the disease [76]. In the brains of AD patients, the activity of calpain-1 increases [100], and animal experiments have also demonstrated the cleavage of Ng by calpain-1, with cleavage sites either within or in proximity to its IQ structural domain. The protease prolyl endopeptidase (PREP) may also participate in further cleavage of Ng [101]. A proteomic study of isolated brain extracts revealed increased levels of α -synuclein and Ng peptides after *in vitro* incubation with brain extracts containing PREP [102]. Currently known substrates for PREP are small neuropeptides that are less than 30 amino acids (AA) in length, such as substance P and thymosin β 4. Therefore, PREP may be involved in fragment cleavage after calpain-1 cleavage. In experimental animals, a study indicates an increase in PREP activity in the brain tissue of aged animals [103]. The application of a PREP inhibitor has been shown to restore memory function in animal models with cognitive impairments [104].

5. Relationships between Ng and AD-associated Genes

A recent study revealed that the gene expression of chitinase domain containing 1 (*CHID1*) is downregulated in the brains of AD patients and that the expression levels of *CHID1* are positively correlated with those of Ng in both normal control and AD groups [105]. Therefore, in the pathogenesis of AD, the *CHID1* gene may interact synergistically with the Ng gene. Multiple reports have linked the apolipoprotein E (APOE) genotype to synaptic function [106]. A study suggests that in the MCI group, CSF Ng levels are significantly greater in APOE ϵ 4 carriers than in noncarriers ($p = 0.001$). In the AD group, there was no statistically significant difference in CSF Ng levels between APOE ϵ 4 carriers and noncarriers ($p = 0.57$). Furthermore, CSF Ng levels increase in a gene dose-dependent manner (heterozygous APOE ϵ 4 vs. homozygous APOE ϵ 4), suggesting that the interaction between Ng, tau protein, and A β 42 may be a crucial mechanism for synaptic damage leading to AD in APOE ϵ 4 carriers [107].

6. Value of Ng in the Diagnosis, Prediction, and Prognosis of AD

In a postmortem study, Ng levels were found to be lower in the hippocampal region and frontal cortex of AD patients, including those with early-onset AD (EOAD),

suggesting its association with synaptic loss and neurodegeneration [27]. A recent autopsy revealed that in cases of intermediate AD, neurons in the CA2 and CA3 regions of the hippocampus exhibit a more rounded morphology, with Ng distribution near the cell body. In contrast, control cases present with Ng in neurons and prominent apical dendrites in the CA2 and CA3 regions of the hippocampus [108]. A study has indicated the specificity of Ng for AD diagnosis. In a study of patients with AD, frontotemporal lobar degeneration (FTLD), Lewy body spectrum of disorders (LBD), and healthy controls, researchers reported that the expression of a synaptic protein was increased in biomarkers specific to AD [109]. These biomarkers include neurogranin and β -synuclein precursor proteins. A study has suggested a reduction in the dendritic shift of Ng mRNA towards dendrites in neocortical tissues in AD patients, whereas in frontotemporal dementia patients, the dendritic shift of Ng mRNA is preserved [110]. Additionally, several studies have reported elevated Ng levels in AD and MCI patients compared with those in control individuals. For example, Saunders *et al.* [111] reported significantly higher Ng levels in the middle-aged temporal lobe inferior gyrus (BA20/21), primary visual cortex (BA17), and hippocampal region (HC) in AD patients than in healthy elderly individuals, whereas BA20/21, BA17, and HC Ng levels were significantly greater in the control group [111]. Kvartsberg *et al.* [78] also obtained similar results, showing higher Ng levels in progressive MCI patients than in stable MCI patients. CSF Ng demonstrates diagnostic accuracy comparable to that of other biomarkers (tau, p-tau, and A β 42) in differentiating AD patients from control individuals [31]. Ng has great potential as a diagnostic biomarker for AD. Research has shown that combining Ng with other AD biomarkers leads to a better assessment of AD. For example, a study revealed that the A β 42/Ng ratio performs better than Ng alone in distinguishing between MCI and moderate AD, as well as exhibiting higher sensitivity and specificity when differentiating AD patients from control individuals. Furthermore, in distinguishing between AD patients and non-AD patients (n-AD), the AUC of A β 42/Ng is slightly superior to that of A β 42/A β 40 [112].

First, changes in blood neuron-derived exosome synaptic protein biomarkers caused by AD were validated, and the biomarkers in blood exosomes could be used to distinguish patients with AD, control patients, and patients with mild cognitive impairment (aMCI). Among the exosome proteins, Ng showed the highest accuracy in distinguishing AD patients from aMCI patients and control groups [113]. Recent study has also yielded similar results, showing significant differences in all plasma neuron-derived extracellular vesicle (NDEV) biomarkers between the control group and individuals with AD. Compared with those in the control group, the plasma levels of Ng are lower in patients with mild to moderate AD, and the levels of Ng in plasma NDEV increase with the severity of

AD and are associated with cognitive and functional decline [114]. Similar to a previous study on Ng in plasma NDEV, this finding can distinguish control subjects from those with mild to moderate AD as well as individuals with MCI, demonstrating high sensitivity [115]. However, a recent study suggested that Ng may not be specific to AD, as CSF Ng does not show a specific increase in clinical or confirmed AD patients. Notably, there was a significant difference in Ng levels between the high-Tau AD group and the AD group and non-AD dementia group [116]. Ng may also be an indicator of disease progression in AD, as previous studies have shown that high tau levels in AD patients can lead to the stabilization of Ng.

Several proteins in CSF have been reported as potential biomarkers for AD diagnosis, but fewer markers are associated with the progression of AD. Current research indicates that Ng, a synaptic protein, has significant potential as a biomarker for the progression of AD. In the early stages of AD, particularly MCI, elevated levels of Ng in CSF predict the rate of future cognitive decline [30,78,117]. Additionally, Ng levels are associated with future hippocampal atrophy and cortical glucose metabolism reduction, especially in patients with positive amyloid- β positron emission tomography (A β -PET) results [30,117]. Importantly, this metabolic reduction is independent of increased brain A β deposition [118,119], reflecting primarily synaptic dysfunction.

In a study by Kvartsberg *et al.* [78], the Cox proportional hazards model predicted the conversion from MCI to dementia, with a hazard ratio of 12.8 (95% CI, 1.6–103.0; $p = 0.02$). Moreover, baseline Ng was highly correlated with an annual decline in mini-mental state examination (MMSE) scores. A meta-analysis of 13 high-quality evidence studies suggested that CSF Ng could predict declines in MMSE scores in patients with MCI associated with A β positivity (A β +MCI), with moderate-quality evidence supporting predictions of declines in memory and executive function [120]. For example, in Portelius *et al.*'s study [31], among 32 individuals from the healthy control group who progressed to MCI or AD during follow-up, the median CSF Ng levels were significantly higher than those in cognitively stable healthy controls. In Tarawneh *et al.*'s study [32], individuals with higher baseline CSF Ng or Ng/A β 42 levels at the 15th percentile progressed more rapidly to cognitive impairment than those with lower levels at the 85th percentile. Baseline CSF Ng levels predict annual changes in clinical dementia rating sum of boxes (CDR-SB), overall memory, episodic memory, and semantic memory scores [31]. In comparison, CSF tau and CSF A β , which are excellent early diagnostic biomarkers, showed weak correlations with cognitive impairment and were less predictive of prognosis [15]. Furthermore, when combined with other biomarkers for AD, Ng exhibits enhanced predictive efficacy. The Ng/BACE1 ratio serves as a reliable predictor of cognitive decline within the continuum of AD. As the

staging of the Ng/BACE1 ratio progresses along the AD continuum, it has a stronger correlation with baseline memory performance and subsequently declines compared with individual measurements of Ng or BACE1 [121]. The undeniable value of Ng in the diagnosis and prognostic assessment of AD is evident, leading to its significance in the field.

7. Potential Therapeutic Role of Ng in AD

The level of neuronal granule protein in synapses decreases before the loss of synapses, indicating that the loss of neuronal granule protein is not only related to synaptic loss. In recent study investigating AD-specific mutant variants, five components have been validated [111]. One component primarily loads on Ng and exhibits weaker loadings on tau indicators, but it is not associated with dementia symptoms. This component is referred to as the NonAD synaptic function. However, the aforementioned study was limited to cross-sectional analysis and did not incorporate longitudinal analysis to investigate gene effects [79]. In conclusion, Ng could serve as a viable target for the treatment of AD. Research has demonstrated that Ng can repair A β -mediated synaptic functional defects by influencing the signalling pathways involved [28]. The overexpression of Ng enhances synaptic transmission in a manner similar to LTP, whereas its removal impedes LTP [36]. Pharmacological interventions that increase Ng levels, such as vitamin A, have partially alleviated age-related synaptic plasticity and memory impairments [122]. However, Ng's hydrolytic enzymes may also play a role in the development of AD, as clinical trials with calpain and/or PREP inhibitors revealed decreased Ng fragment concentrations in the CSF of patients, potentially improving cognitive function [85].

8. Conclusion

Dysfunction, loss, or damage to synapses can lead to cognitive decline. Synaptic proteins can serve as indicators of synaptic status, and Ng is widely present in the cerebral cortex, where it participates in signalling pathways related to LTP and LTD. Therefore, Ng may reflect the functional state of synapses and cognitive changes. In the context of AD, cognitive decline is associated with synaptic damage, making Ng one of the most promising synaptic proteins as a potential AD biomarker. Numerous studies on Ng in AD have demonstrated a clear upwards trend in its levels in the CSF of patients with MCI and AD. However, further research is needed to establish Ng as a diagnostic criterion for AD or for differentiating AD from other types of dementia and neurodegenerative diseases.

Currently, there are limited treatment options for AD, most of which are symptomatic. Several experimental treatments, including those targeting Ng, are in the early stages of development. Numerous studies have also suggested that increasing Ng levels can improve cognitive function in AD patients. The mechanisms underlying the reduction in Ng

in the cortical regions of the brain in AD patients remain unclear and warrant further investigation. Therefore, therapeutic approaches targeting Ng require additional research.

The direct mechanism by which Ng is reduced in AD is still unclear. As mentioned earlier, studies have shown that changes in Ng may occur before significant synaptic damage occurs, and animal experiments have shown that knocking out the *Ng* gene can affect cognitive function. However, this does not necessarily indicate an independent mechanism unrelated to A β and tau that leads to an increase in Ng levels in AD. Further research is needed to clarify the following issues: (1) whether there are pathological mechanisms directly leading to a decrease in Ng; (2) what is the mechanism by which Ng moves from inside of the cells to cerebrospinal fluid when synapses are relatively intact; (3) whether there are pathogenic mutations related to Ng in AD; and (4) whether the pathological mechanisms of AD affect the transcription, translation, and degradation processes of Ng.

Author Contributions

XZ conceived the project, carried out analysis of previous topics, and wrote the preliminary draft. XJJ conceived the project. HZ contributed to the conception of the study, design of the content framework, final manuscript preparation, and project supervision. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

This research received no external funding.

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Irwin MR, Vitiello MV. Implications of sleep disturbance and inflammation for Alzheimer's disease dementia. *The Lancet. Neurology*. 2019; 18: 296–306.
- [2] McGeer PL, McGeer EG. The amyloid cascade-inflammatory hypothesis of Alzheimer disease: implications for therapy. *Acta Neuropathologica*. 2013; 126: 479–497.
- [3] Vickers JC, Mitew S, Woodhouse A, Fernandez-Martos CM, Kirkcaldie MT, Canty AJ, *et al*. Defining the earliest pathological changes of Alzheimer's disease. *Current Alzheimer Research*. 2016; 13: 281–287.
- [4] Tzioras M, McGeachan RI, Durrant CS, Spires-Jones TL.

Synaptic degeneration in Alzheimer disease. *Nature Reviews. Neurology*. 2023; 19: 19–38.

- [5] Masliah E, Mallory M, Alford M, DeTeresa R, Hansen LA, McKeel DW, Jr, *et al.* Altered expression of synaptic proteins occurs early during progression of Alzheimer's disease. *Neurology*. 2001; 56: 127–129.
- [6] Milà-Alomà M, Salvadó G, Gispert JD, Vilor-Tejedor N, Grau-Rivera O, Sala-Vila A, *et al.* Amyloid beta, tau, synaptic, neurodegeneration, and glial biomarkers in the preclinical stage of the Alzheimer's continuum. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2020; 16: 1358–1371.
- [7] Ito K, Ahadié S, Corrigan B, French J, Fullerton T, Tensfeldt T, *et al.* Disease progression meta-analysis model in Alzheimer's disease. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2010; 6: 39–53.
- [8] Jack CR, Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, *et al.* Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *The Lancet. Neurology*. 2010; 9: 119–128.
- [9] Colom-Cadena M, Spires-Jones T, Zetterberg H, Blennow K, Caggiano A, DeKosky ST, *et al.* The clinical promise of biomarkers of synapse damage or loss in Alzheimer's disease. *Alzheimer's Research & Therapy*. 2020; 12: 1–12.
- [10] DeKosky ST, Scheff SW, Styren SD. Structural correlates of cognition in dementia: quantification and assessment of synapse change. *Neurodegeneration: a Journal for Neurodegenerative Disorders, Neuroprotection, and Neuroregeneration*. 1996; 5: 417–421.
- [11] Bliss TV, Lomo T. Long-lasting potentiation of synaptic transmission in the dentate area of the anesthetized rabbit following stimulation of the perforant path. *The Journal of Physiology*. 1973; 232: 331–356.
- [12] Takeuchi T, Duszkiewicz AJ, Morris RGM. The synaptic plasticity and memory hypothesis: encoding, storage and persistence. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*. 2013; 369: 20130288.
- [13] de Wilde MC, Overk CR, Sijben JW, Masliah E. Meta-analysis of synaptic pathology in Alzheimer's disease reveals selective molecular vesicular machinery vulnerability. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2016; 12: 633–644.
- [14] Spires-Jones TL, Hyman BT. The intersection of amyloid beta and tau at synapses in Alzheimer's disease. *Neuron*. 2014; 82: 756–771.
- [15] Blennow K, Bogdanovic N, Alafuzoff I, Ekman R, Davidsson P. Synaptic pathology in Alzheimer's disease: relation to severity of dementia, but not to senile plaques, neurofibrillary tangles, or the ApoE4 allele. *Journal of Neural Transmission (Vienna, Austria)*. 1996; 103: 603–618.
- [16] Tönnies E, Trushina E. Oxidative Stress, Synaptic Dysfunction, and Alzheimer's Disease. *Journal of Alzheimer's Disease: JAD*. 2017; 57: 1105–1121.
- [17] Jack CR, Jr, Shiung MM, Weigand SD, O'Brien PC, Gunter JL, Boeve BF, *et al.* Brain atrophy rates predict subsequent clinical conversion in normal elderly and amnesic MCI. *Neurology*. 2005; 65: 1227–1231.
- [18] Tible M, Sandelius Å, Höglund K, Brinkmalm A, Cognat E, Dumurgier J, *et al.* Dissection of synaptic pathways through the CSF biomarkers for predicting Alzheimer disease. *Neurology*. 2020; 95: e953–e961.
- [19] Mazzucchi S, Palermo G, Campese N, Galgani A, Della Vecchia A, Vergallo A, *et al.* The role of synaptic biomarkers in the spectrum of neurodegenerative diseases. *Expert Review of Proteomics*. 2020; 17: 543–559.
- [20] Nilsson J, Pichet Binette A, Palmqvist S, Brum WS, Janelidze S, Ashton NJ, *et al.* Cerebrospinal fluid biomarker panel for synaptic dysfunction in a broad spectrum of neurodegenerative diseases. *Brain: a Journal of Neurology*. 2024; 147: 2414–2427.
- [21] Kivisäkk P, Carlyle BC, Sweeney T, Quinn JP, Ramirez CE, Trombetta BA, *et al.* Increased levels of the synaptic proteins PSD-95, SNAP-25, and neurogranin in the cerebrospinal fluid of patients with Alzheimer's disease. *Alzheimer's Research & Therapy*. 2022; 14: 58.
- [22] Portelius E, Olsson B, Höglund K, Cullen NC, Kvartsberg H, Andreasson U, *et al.* Cerebrospinal fluid neurogranin concentration in neurodegeneration: relation to clinical phenotypes and neuropathology. *Acta Neuropathologica*. 2018; 136: 363–376.
- [23] Wellington H, Paterson RW, Portelius E, Törnqvist U, Magdalino N, Fox NC, *et al.* Increased CSF neurogranin concentration is specific to Alzheimer disease. *Neurology*. 2016; 86: 829–835.
- [24] Martínez de Arrieta C, Pérez Jurado L, Bernal J, Coloma A. Structure, organization, and chromosomal mapping of the human neurogranin gene (NRGN). *Genomics*. 1997; 41: 243–249.
- [25] Warpechowski M, Warpechowski J, Kulczyńska-Przybik A, Mroczko B. Biomarkers of Activity-Dependent Plasticity and Persistent Enhancement of Synaptic Transmission in Alzheimer Disease: A Review of the Current Status. *Medical Science Monitor: International Medical Journal of Experimental and Clinical Research*. 2023; 29: e938826.
- [26] Díez-Guerra FJ. Neurogranin, a link between calcium/calmodulin and protein kinase C signaling in synaptic plasticity. *IUBMB Life*. 2010; 62: 597–606.
- [27] Davidsson P, Blennow K. Neurochemical dissection of synaptic pathology in Alzheimer's disease. *International Psychogeriatrics*. 1998; 10: 11–23.
- [28] Kaleka KS, Gerges NZ. Neurogranin restores amyloid β -mediated synaptic transmission and long-term potentiation deficits. *Experimental Neurology*. 2016; 277: 115–123.
- [29] Casaletto KB, Elahi FM, Bettcher BM, Neuhaus J, Bendlin BB, Asthana S, *et al.* Neurogranin, a synaptic protein, is associated with memory independent of Alzheimer biomarkers. *Neurology*. 2017; 89: 1782–1788.
- [30] Headley A, De Leon-Benedetti A, Dong C, Levin B, Loewenstein D, Camargo C, *et al.* Neurogranin as a predictor of memory and executive function decline in MCI patients. *Neurology*. 2018; 90: e887–e895.
- [31] Portelius E, Zetterberg H, Skillbäck T, Törnqvist U, Andreasson U, Trojanowski JQ, *et al.* Cerebrospinal fluid neurogranin: relation to cognition and neurodegeneration in Alzheimer's disease. *Brain: a Journal of Neurology*. 2015; 138: 3373–3385.
- [32] Tarawneh R, D'Angelo G, Crimmins D, Herries E, Griest T, Fagan AM, *et al.* Diagnostic and Prognostic Utility of the Synaptic Marker Neurogranin in Alzheimer Disease. *JAMA Neurology*. 2016; 73: 561–571.
- [33] Xue M, Sun FR, Ou YN, Shen XN, Li HQ, Huang YY, *et al.* Association of cerebrospinal fluid neurogranin levels with cognition and neurodegeneration in Alzheimer's disease. *Aging*. 2020; 12: 9365–9379.
- [34] Hellwig K, Kvartsberg H, Portelius E, Andreasson U, Oberstein TJ, Lewczuk P, *et al.* Neurogranin and YKL-40: independent markers of synaptic degeneration and neuroinflammation in Alzheimer's disease. *Alzheimer's Research & Therapy*. 2015; 7: 74.
- [35] Janelidze S, Hertze J, Zetterberg H, Landqvist Waldö M, Santillo A, Blennow K, *et al.* Cerebrospinal fluid neurogranin and YKL-40 as biomarkers of Alzheimer's disease. *Annals of Clinical and Translational Neurology*. 2015; 3: 12–20.
- [36] Zhong L, Cherry T, Bies CE, Florence MA, Gerges NZ. Neurogranin enhances synaptic strength through its interaction with calmodulin. *The EMBO Journal*. 2009; 28: 3027–3039.
- [37] Li L, Lai M, Cole S, Le Novère N, Edelstein SJ. Neurogranin stimulates Ca²⁺/calmodulin-dependent kinase II by suppressing

- calcineurin activity at specific calcium spike frequencies. *PLoS Computational Biology*. 2020; 16: e1006991.
- [38] Smolen P. A model of late long-term potentiation simulates aspects of memory maintenance. *PloS One*. 2007; 2: e445.
 - [39] Burgdorf J, Zhang XL, Weiss C, Matthews E, Disterhoft JF, Stanton PK, *et al.* The N-methyl-D-aspartate receptor modulator GLYX-13 enhances learning and memory, in young adult and learning impaired aging rats. *Neurobiology of Aging*. 2011; 32: 698–706.
 - [40] Ma T, Klann E. Amyloid β : linking synaptic plasticity failure to memory disruption in Alzheimer's disease. *Journal of Neurochemistry*. 2012; 120 Suppl 1: 140–148.
 - [41] Baudier J, Deloulme JC, Van Dorsselaer A, Black D, Matthes HW. Purification and characterization of a brain-specific protein kinase C substrate, neurogranin (p17). Identification of a consensus amino acid sequence between neurogranin and neuromodulin (GAP43) that corresponds to the protein kinase C phosphorylation site and the calmodulin-binding domain. *The Journal of Biological Chemistry*. 1991; 266: 229–237.
 - [42] Petersen A, Gerges NZ. Neurogranin regulates CaM dynamics at dendritic spines. *Scientific Reports*. 2015; 5: 11135.
 - [43] Zhong L, Gerges NZ. Neurogranin targets calmodulin and lowers the threshold for the induction of long-term potentiation. *PloS One*. 2012; 7: e41275.
 - [44] O'Day DH. Calmodulin Binding Proteins and Alzheimer's Disease: Biomarkers, Regulatory Enzymes and Receptors That Are Regulated by Calmodulin. *International Journal of Molecular Sciences*. 2020; 21: 7344.
 - [45] Liu W, Lin H, He X, Chen L, Dai Y, Jia W, *et al.* Neurogranin as a cognitive biomarker in cerebrospinal fluid and blood exosomes for Alzheimer's disease and mild cognitive impairment. *Translational Psychiatry*. 2020; 10: 125.
 - [46] Zhong L, Kaleka KS, Gerges NZ. Neurogranin phosphorylation fine-tunes long-term potentiation. *The European Journal of Neuroscience*. 2011; 33: 244–250.
 - [47] Huang KP, Huang FL, Li J, Schuck P, McPhie P. Calcium-sensitive interaction between calmodulin and modified forms of rat brain neurogranin/RC3. *Biochemistry*. 2000; 39: 7291–7299.
 - [48] Mons N, Enderlin V, Jaffard R, Huguier P. Selective age-related changes in the PKC-sensitive, calmodulin-binding protein, neurogranin, in the mouse brain. *Journal of Neurochemistry*. 2001; 79: 859–867.
 - [49] Nakajima R, Hattori S, Funasaka T, Huang FL, Miyakawa T. Decreased nesting behavior, selective increases in locomotor activity in a novel environment, and paradoxically increased open arm exploration in Neurogranin knockout mice. *Neuropsychopharmacology Reports*. 2021; 41: 111–116.
 - [50] Höglund K, Schussler N, Kvartsberg H, Smailovic U, Brinkmalm G, Liman V, *et al.* Cerebrospinal fluid neurogranin in an inducible mouse model of neurodegeneration: A translatable marker of synaptic degeneration. *Neurobiology of Disease*. 2020; 134: 104645.
 - [51] Sun X, Wang Q, Blennow K, Zetterberg H, McCarthy M, Loewenstein DA, *et al.* Association of neurogranin gene expression with Alzheimer's disease pathology in the perirhinal cortex. *Alzheimer's & Dementia (New York, N. Y.)*. 2021; 7: e12162.
 - [52] Masters CL, Selkoe DJ. Biochemistry of amyloid β -protein and amyloid deposits in Alzheimer disease. *Cold Spring Harbor Perspectives in Medicine*. 2012; 2: a006262.
 - [53] Griffiths J, Grant SGN. Synapse pathology in Alzheimer's disease. *Seminars in Cell & Developmental Biology*. 2023; 139: 13–23.
 - [54] Cline EN, Bicca MA, Viola KL, Klein WL. The Amyloid- β Oligomer Hypothesis: Beginning of the Third Decade. *Journal of Alzheimer's Disease: JAD*. 2018; 64: S567–S610.
 - [55] Shankar GM, Li S, Mehta TH, Garcia-Munoz A, Shepardson NE, Smith I, *et al.* Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nature Medicine*. 2008; 14: 837–842.
 - [56] Lacor PN, Buniel MC, Furlow PW, Clemente AS, Velasco PT, Wood M, *et al.* Abeta oligomer-induced aberrations in synapse composition, shape, and density provide a molecular basis for loss of connectivity in Alzheimer's disease. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2007; 27: 796–807.
 - [57] Wang Z, Jackson RJ, Hong W, Taylor WM, Corbett GT, Moreno A, *et al.* Human Brain-Derived A β Oligomers Bind to Synapses and Disrupt Synaptic Activity in a Manner That Requires APP. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2017; 37: 11947–11966.
 - [58] Calabrese B, Shaked GM, Tabarean IV, Braga J, Koo EH, Halpain S. Rapid, concurrent alterations in pre- and postsynaptic structure induced by naturally-secreted amyloid-beta protein. *Molecular and Cellular Neurosciences*. 2007; 35: 183–193.
 - [59] Koffie RM, Hashimoto T, Tai HC, Kay KR, Serrano-Pozo A, Joyner D, *et al.* Apolipoprotein E4 effects in Alzheimer's disease are mediated by synaptotoxic oligomeric amyloid- β . *Brain: a Journal of Neurology*. 2012; 135: 2155–2168.
 - [60] Lue LF, Kuo YM, Roher AE, Brachova L, Shen Y, Sue L, *et al.* Soluble amyloid beta peptide concentration as a predictor of synaptic change in Alzheimer's disease. *The American Journal of Pathology*. 1999; 155: 853–862.
 - [61] Bao F, Wicklund L, Lacor PN, Klein WL, Nordberg A, Marutle A. Different β -amyloid oligomer assemblies in Alzheimer brains correlate with age of disease onset and impaired cholinergic activity. *Neurobiology of Aging*. 2012; 33: 825.e1–13.
 - [62] Koffie RM, Meyer-Luehmann M, Hashimoto T, Adams KW, Mielke ML, Garcia-Alloza M, *et al.* Oligomeric amyloid beta associates with postsynaptic densities and correlates with excitatory synapse loss near senile plaques. *Proceedings of the National Academy of Sciences of the United States of America*. 2009; 106: 4012–4017.
 - [63] Pickett EK, Koffie RM, Wegmann S, Henstridge CM, Herrmann AG, Colom-Cadena M, *et al.* Non-Fibrillar Oligomeric Amyloid- β within Synapses. *Journal of Alzheimer's Disease: JAD*. 2016; 53: 787–800.
 - [64] Lambert MP, Barlow AK, Chromy BA, Edwards C, Freed R, Liosatos M, *et al.* Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins. *Proceedings of the National Academy of Sciences of the United States of America*. 1998; 95: 6448–6453.
 - [65] Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, Wolfe MS, *et al.* Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. *Nature*. 2002; 416: 535–539.
 - [66] Willén K, Sroka A, Takahashi RH, Gouras GK. Heterogeneous Association of Alzheimer's Disease-Linked Amyloid- β and Amyloid- β Protein Precursor with Synapses. *Journal of Alzheimer's Disease: JAD*. 2017; 60: 511–524.
 - [67] Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi EY, *et al.* Regulation of NMDA receptor trafficking by amyloid-beta. *Nature Neuroscience*. 2005; 8: 1051–1058.
 - [68] Trushina E, McMurray CT. Oxidative stress and mitochondrial dysfunction in neurodegenerative diseases. *Neuroscience*. 2007; 145: 1233–1248.
 - [69] Bezprozvanny I, Mattson MP. Neuronal calcium mishandling and the pathogenesis of Alzheimer's disease. *Trends in Neurosciences*. 2008; 31: 454–463.
 - [70] Li S, Selkoe DJ. A mechanistic hypothesis for the impairment of synaptic plasticity by soluble A β oligomers from Alzheimer's brain. *Journal of Neurochemistry*. 2020; 154: 583–597.
 - [71] Zhang H, Jiang X, Ma L, Wei W, Li Z, Chang S, *et al.* Role

- of A β in Alzheimer's-related synaptic dysfunction. *Frontiers in Cell and Developmental Biology*. 2022; 10: 964075.
- [72] O'Day DH, Myre MA. Calmodulin-binding domains in Alzheimer's disease proteins: extending the calcium hypothesis. *Biochemical and Biophysical Research Communications*. 2004; 320: 1051–1054.
- [73] O'Day DH. Alzheimer's Disease: A short introduction to the calmodulin hypothesis. *AIMS Neuroscience*. 2019; 6: 231–239.
- [74] Cárdenas AM, Ardiles AO, Barraza N, Baéz-Matus X, Caviedes P. Role of tau protein in neuronal damage in Alzheimer's disease and Down syndrome. *Archives of Medical Research*. 2012; 43: 645–654.
- [75] Andreadis A, Brown WM, Kosik KS. Structure and novel exons of the human tau gene. *Biochemistry*. 1992; 31: 10626–10633.
- [76] Kvartsberg H, Portelius E, Andreasson U, Brinkmalm G, Hellwig K, Lelental N, *et al.* Characterization of the postsynaptic protein neurogranin in paired cerebrospinal fluid and plasma samples from Alzheimer's disease patients and healthy controls. *Alzheimer's Research & Therapy*. 2015; 7: 40.
- [77] Bertoni-Freddari C, Fattoretti P, Casoli T, Caselli U, Meier-Ruge W. Deterioration threshold of synaptic morphology in aging and senile dementia of Alzheimer's type. *Analytical and Quantitative Cytology and Histology*. 1996; 18: 209–213.
- [78] Kvartsberg H, Duits FH, Ingelsson M, Andreasen N, Öhrfelt A, Andersson K, *et al.* Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive decline in prodromal Alzheimer's disease. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2015; 11: 1180–1190.
- [79] Neumann A, Ohlei O, Küçükali F, Bos IJ, Timsina J, Vos S, *et al.* Multivariate GWAS of Alzheimer's disease CSF biomarker profiles implies GRIN2D in synaptic functioning. *Genome Medicine*. 2023; 15: 79.
- [80] Hole KL, Zhu B, Huggon L, Brown JT, Mason JM, Williams RJ. Tau^{P301L} disengages from the proteasome core complex and neurogranin coincident with enhanced neuronal network excitability. *Cell Death & Disease*. 2024; 15: 429.
- [81] Litersky JM, Johnson GV, Jakes R, Goedert M, Lee M, Seubert P. Tau protein is phosphorylated by cyclic AMP-dependent protein kinase and calcium/calmodulin-dependent protein kinase II within its microtubule-binding domains at Ser-262 and Ser-356. *The Biochemical Journal*. 1996; 316 (Pt 2): 655–660.
- [82] Wei Q, Holzer M, Brueckner MK, Liu Y, Arendt T. Dephosphorylation of tau protein by calcineurin triturated into neural living cells. *Cellular and Molecular Neurobiology*. 2002; 22: 13–24.
- [83] Connolly K, Lehoux M, O'Rourke R, Assetta B, Erdemir GA, Elias JA, *et al.* Potential role of chitinase-3-like protein 1 (CHI3L1/YKL-40) in neurodegeneration and Alzheimer's disease. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2023; 19: 9–24.
- [84] Liddelow SA, Barres BA. Reactive Astrocytes: Production, Function, and Therapeutic Potential. *Immunity*. 2017; 46: 957–967.
- [85] Liddelow SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, *et al.* Neurotoxic reactive astrocytes are induced by activated microglia. *Nature*. 2017; 541: 481–487.
- [86] Llorens F, Thüne K, Tahir W, Kanata E, Diaz-Lucena D, Xanthopoulos K, *et al.* YKL-40 in the brain and cerebrospinal fluid of neurodegenerative dementias. *Molecular Neurodegeneration*. 2017; 12: 83.
- [87] Bonneh-Barkay D, Wang G, Starkey A, Hamilton RL, Wiley CA. In vivo CHI3L1 (YKL-40) expression in astrocytes in acute and chronic neurological diseases. *Journal of Neuroinflammation*. 2010; 7: 34.
- [88] Bonneh-Barkay D, Zagadailov P, Zou H, Niyonkuru C, Figley M, Starkey A, *et al.* YKL-40 expression in traumatic brain injury: an initial analysis. *Journal of Neurotrauma*. 2010; 27: 1215–1223.
- [89] Comabella M, Fernández M, Martín R, Rivera-Vallvé S, Borrás E, Chiva C, *et al.* Cerebrospinal fluid chitinase 3-like 1 levels are associated with conversion to multiple sclerosis. *Brain: a Journal of Neurology*. 2010; 133: 1082–1093.
- [90] Rogers JT, Morganti JM, Bachstetter AD, Hudson CE, Peters MM, Grimmig BA, *et al.* CX3CR1 deficiency leads to impairment of hippocampal cognitive function and synaptic plasticity. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2011; 31: 16241–16250.
- [91] Merino JJ, Muñetón-Gómez V, Álvarez MI, Toledano-Díaz A. Effects of CX3CR1 and Fractalkine Chemokines in Amyloid Beta Clearance and p-Tau Accumulation in Alzheimer's Disease (AD) Rodent Models: Is Fractalkine a Systemic Biomarker for AD? *Current Alzheimer Research*. 2016; 13: 403–412.
- [92] Kulczyńska-Przybyk A, Dulewicz M, Doroszkiewicz J, Borawska R, Słowik A, Zetterberg H, *et al.* The Relationships between Cerebrospinal Fluid Glial (CXCL12, CX3CL, YKL-40) and Synaptic Biomarkers (Ng, NPTXR) in Early Alzheimer's Disease. *International Journal of Molecular Sciences*. 2023; 24: 13166.
- [93] Quintana E, Coll C, Salavedra-Pont J, Muñoz-San Martín M, Robles-Cedeño R, Tomás-Roig J, *et al.* Cognitive impairment in early stages of multiple sclerosis is associated with high cerebrospinal fluid levels of chitinase 3-like 1 and neurofilament light chain. *European Journal of Neurology*. 2018; 25: 1189–1191.
- [94] Vu L, An J, Kovalik T, Gendron T, Petrucelli L, Bowser R. Cross-sectional and longitudinal measures of chitinase proteins in amyotrophic lateral sclerosis and expression of CHI3L1 in activated astrocytes. *Journal of Neurology, Neurosurgery, and Psychiatry*. 2020; 91: 350–358.
- [95] Woollacott IOC, Nicholas JM, Heller C, Foiani MS, Moore KM, Russell LL, *et al.* Cerebrospinal Fluid YKL-40 and Chitotriosidase Levels in Frontotemporal Dementia Vary by Clinical, Genetic and Pathological Subtype. *Dementia and Geriatric Cognitive Disorders*. 2020; 49: 56–76.
- [96] Camacho-Hernández NP, Peña-Ortega F. Fractalkine/CX3CR1-Dependent Modulation of Synaptic and Network Plasticity in Health and Disease. *Neural Plasticity*. 2023; 2023: 4637073.
- [97] Basilico B, Ferrucci L, Ratano P, Golia MT, Grimaldi A, Rosito M, *et al.* Microglia control glutamatergic synapses in the adult mouse hippocampus. *Glia*. 2022; 70: 173–195.
- [98] Sheridan GK, Wdowicz A, Pickering M, Watters O, Halley P, O'Sullivan NC, *et al.* CX3CL1 is up-regulated in the rat hippocampus during memory-associated synaptic plasticity. *Frontiers in Cellular Neuroscience*. 2014; 8: 233.
- [99] Kvartsberg H, Lashley T, Murray CE, Brinkmalm G, Cullen NC, Höglund K, *et al.* The intact postsynaptic protein neurogranin is reduced in brain tissue from patients with familial and sporadic Alzheimer's disease. *Acta Neuropathologica*. 2019; 137: 89–102.
- [100] Liu J, Liu MC, Wang KKW. Calpain in the CNS: from synaptic function to neurotoxicity. *Science Signaling*. 2008; 1: re1.
- [101] Becker B, Nazir FH, Brinkmalm G, Camporesi E, Kvartsberg H, Portelius E, *et al.* Alzheimer-associated cerebrospinal fluid fragments of neurogranin are generated by Calpain-1 and prolyl endopeptidase. *Molecular Neurodegeneration*. 2018; 13: 47.
- [102] Brandt I, De Vriendt K, Devreese B, Van Beeumen J, Van Dongen W, Augustyns K, *et al.* Search for substrates for prolyl oligopeptidase in porcine brain. *Peptides*. 2005; 26: 2536–2546.
- [103] Jiang CH, Tsien JZ, Schultz PG, Hu Y. The effects of aging on gene expression in the hypothalamus and cortex of mice. *Proceedings of the National Academy of Sciences of the United States of America*. 2001; 98: 1930–1934.
- [104] Männistö PT, Venäläinen J, Jalkanen A, García-Horsman JA.

Prolyl oligopeptidase: a potential target for the treatment of cognitive disorders. *Drug News & Perspectives*. 2007; 20: 293–305.

- [105] Castrogiovanni P, Sanfilippo C, Imbesi R, Maugeri G, Lo Furno D, Tibullo D, *et al.* Brain *CHDI* Expression Correlates with *NRGN* and *CALB1* in Healthy Subjects and AD Patients. *Cells*. 2021; 10: 882.
- [106] Wang C, Wilson WA, Moore SD, Mace BE, Maeda N, Schmechel DE, *et al.* Human apoE4-targeted replacement mice display synaptic deficits in the absence of neuropathology. *Neurobiology of Disease*. 2005; 18: 390–398.
- [107] Sun X, Dong C, Levin B, Crocco E, Loewenstein D, Zetterberg H, *et al.* APOE ϵ 4 carriers may undergo synaptic damage conferring risk of Alzheimer's disease. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2016; 12: 1159–1166.
- [108] Vontell RT, Gober R, Dallmeier J, Brzostowski D, Barreda A, Blennow K, *et al.* Association of region-specific hippocampal reduction of neurogranin with inflammasome proteins in *post mortem* brains of Alzheimer's disease. *Alzheimer's & Dementia* (New York, N. Y.). 2024; 10: e12444.
- [109] Nilsson J, Cousins K, Gobom J, Portelius E, Chen-Plotkin A, Shaw LM, *et al.* Cerebrospinal fluid biomarker panel of synaptic dysfunction in Alzheimer's disease and other neurodegenerative disorders. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2023; 19: 1775–1784.
- [110] Chang JW, Schumacher E, Coulter PM, 2nd, Vinters HV, Watson JB. Dendritic translocation of RC3/neurogranin mRNA in normal aging, Alzheimer disease and fronto-temporal dementia. *Journal of Neuropathology and Experimental Neurology*. 1997; 56: 1105–1118.
- [111] Saunders T, Gunn C, Blennow K, Kvartsberg H, Zetterberg H, Shenkin SD, *et al.* Neurogranin in Alzheimer's disease and ageing: A human post-mortem study. *Neurobiology of Disease*. 2023; 177: 105991.
- [112] Piccoli T, Blandino V, Maniscalco L, Matranga D, Graziano F, Guajana F, *et al.* Biomarkers Related to Synaptic Dysfunction to Discriminate Alzheimer's Disease from Other Neurological Disorders. *International Journal of Molecular Sciences*. 2022; 23: 10831.
- [113] Jia L, Zhu M, Kong C, Pang Y, Zhang H, Qiu Q, *et al.* Blood neuro-exosomal synaptic proteins predict Alzheimer's disease at the asymptomatic stage. *Alzheimer's & Dementia: the Journal of the Alzheimer's Association*. 2021; 17: 49–60.
- [114] Alvarez XA, Winston CN, Barlow JW, Sarsoza FM, Alvarez I, Aleixandre M, *et al.* Modulation of Amyloid- β and Tau in Alzheimer's Disease Plasma Neuronal-Derived Extracellular Vesicles by Cerebrolisin® and Donepezil. *Journal of Alzheimer's Disease: JAD*. 2022; 90: 705–717.
- [115] Winston CN, Goetzl EJ, Akers JC, Carter BS, Rockenstein EM, Galasko D, *et al.* Prediction of conversion from mild cognitive impairment to dementia with neuronally derived blood exosome protein profile. *Alzheimer's & Dementia* (Amsterdam, Netherlands). 2016; 3: 63–72.
- [116] Willemse EAJ, Sieben A, Somers C, Vermeiren Y, De Roeck N, Timmers M, *et al.* Neurogranin as biomarker in CSF is non-specific to Alzheimer's disease dementia. *Neurobiology of Aging*. 2021; 108: 99–109.
- [117] Mattsson N, Insel PS, Palmqvist S, Portelius E, Zetterberg H, Weiner M, *et al.* Cerebrospinal fluid tau, neurogranin, and neurofilament light in Alzheimer's disease. *EMBO Molecular Medicine*. 2016; 8: 1184–1196.
- [118] Jagust WJ, Landau SM. Apolipoprotein E, not fibrillar β -amyloid, reduces cerebral glucose metabolism in normal aging. *Journal of Neuroscience*. 2012; 32: 18227–18233.
- [119] Mosconi L, Pupi A, De Leon MJ. Brain glucose hypometabolism and oxidative stress in preclinical Alzheimer's disease. *Annals of the New York Academy of Sciences*. 2008; 1147: 180–195.
- [120] Yoong SQ, Lu J, Xing H, Gyanwali B, Tan YQ, Wu XV. The prognostic utility of CSF neurogranin in predicting future cognitive decline in the Alzheimer's disease continuum: A systematic review and meta-analysis with narrative synthesis. *Ageing Research Reviews*. 2021; 72: 101491.
- [121] Kirsebom BE, Richter G, Nordengen K, Aarsland D, Bråthen G, Tijms BM, *et al.* Stable cerebrospinal fluid neurogranin and β -site amyloid precursor protein cleaving enzyme 1 levels differentiate predementia Alzheimer's disease patients. *Brain Communications*. 2022; 4: fcac244.
- [122] Etchamendy N, Enderlin V, Marighetto A, Vouimba RM, Pallet V, Jaffard R, *et al.* Alleviation of a selective age-related relational memory deficit in mice by pharmacologically induced normalization of brain retinoid signaling. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*. 2001; 21: 6423–6429.