

Original Research

## Coagulation Disorders in Patients With Acute Respiratory Distress Syndrome Following Acute Aortic Dissection: A Prospective Observational Study

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#### Abstract

Background: Coagulation disorders are potentially one of the most important pathogeneses of acute respiratory distress syndrome (ARDS) following acute type A aortic dissection (ATAAD). This study aimed to determine whether aortic dissection singularly and cardiopulmonary bypass (CPB) surgery can activate coagulation pathways, promoting ARDS development in patients with ATAAD. Methods: A total of 450 patients who received treatment at Beijing Anzhen Hospital, Capital Medical University, between March 2023 and February 2024 were consecutively enrolled in this prospective cohort study. We analyzed the clinical factors and measured serum coagulation biomarkers by enzyme-linked immunosorbent assay (ELISA) among patients with ATAAD, aortic aneurysm (AA), or unstable angina (UA). Logistic regression, two-way analysis of variance (ANOVA), and Spearman's correlation analysis were performed. Furthermore, the patients with ATAAD were divided into ARDS (based on chest radiographic findings and an oxygenation index ≤300 mmHg) and non-ARDS groups for subgroup comparisons. Results: The incidence of postoperative ARDS among patients with ATAAD was 20.7% (13.3% in the AA group and 7.3% in the UA group). Preoperatively, prothrombin time (PT) was longer in patients with ATAAD than in those with AA or UA ((odds ratio (OR): 12.0, 95% confidence interval (CI): 11.5-12.6) vs. (OR: 11.4, 95% CI: 10.9-12.1) vs. (OR: 11.2, 95% CI: 10.8–11.6), respectively; p < 0.001). The D-dimer levels, fibrin degradation products (FDPs), factor XIIa, and factor VIII-Ag (FVIII-Ag) were significantly elevated preoperatively and postoperatively in patients with ATAAD. The FDP levels in the ATAAD subgroup immediately after surgery were significantly higher in the ARDS group compared with those in the non-ARDS group (OR: 2.26, 95% CI: 1.13–4.54; p = 0.022). In addition, a negative correlation existed between the FXII level (correlation coefficient r = -0.682, p = 0.043) at 24 hours after surgery and the oxygenation index. **Conclusion**: Coagulation activation may be caused by a ortic dissection singularly and CPB, which promotes postoperative ARDS in patients with ATAAD.

Keywords: acute aortic dissection; acute respiratory distress syndrome; coagulation; fibrinolysis; cardiopulmonary bypass

### 1. Introduction

Acute lung injury (ALI) and acute respiratory distress syndrome (ARDS) are common complications that may occur during perioperative management, especially in high-risk populations of patients undergoing surgery [1–3]. ARDS is a primary cause of postoperative acute hypoxemic respiratory failure and its etiologies are diverse [4–7]. The occurrence of ARDS has also been investigated following cardiac surgery (CS) involving cardiopulmonary bypass (CPB) procedures [3,7–9]. Despite the continual refinement of perioperative management and surgical techniques, up to 20% of patients are affected by this postsurgical complication [3,10]. Postoperative ARDS can seriously affect patient outcomes, with mortality rates as high as 40–80% in severe cases [2,7,11,12] and functional limitations sometimes persisting 5 years after an ARDS episode [13].

Although the pathogenesis of ARDS is not entirely clear, recent studies suggest that the activation of coagulation is closely related to the development of ARDS [6,14–

18] and blockade of the initiating steps of the clotting cascade may be beneficial for patients with ARDS [6,19]. During cardiac surgery, CPB procedures and extensive surgical trauma can induce the widespread activation of the coagulation system [17,20,21], and the pathogenesis of acute type A aortic dissection (ATAAD) has been shown to be associated with coagulation disorders [22]. Therefore, both ATAAD itself and CPB can contribute to the development of multiple disorders of the coagulation system; however, there is little data supporting the activation of coagulation cascade plays a role in the etiology of ARDS after aortic surgery.

The occurrence of ALI/ARDS following aortic surgery is unpredictable, although recent studies have suggested that early identification and improved perioperative care can prevent postoperative ALI/ARDS. Although several risk factors have been identified are associated with the occurrence of post-operative ARDS, to our knowledge, barely any study has illustrated the association between coagulation markers and ARDS in ATAAD patients. There-

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fore, the aim of this study was to explore the correlations between the serum levels of various biomarkers of coagulation and the occurrence of ARDS following aortic surgery, as these markers could serve as reliable predictors of the risk of ALI/ARDS.

### 2. Methods

### 2.1 Design and Participants

This single-center prospective observational study was performed at the Center for Cardiac Intensive Care of Beijing Anzhen Hospital, Capital Medical University. A total of 450 patients were consecutively enrolled between March 2023 and February 2024, including 150 patients diagnosed with ATAAD, 150 with aortic aneurysm (AA), and 150 with unstable angina (UA). Patients with ATAAD and AA underwent Sun's surgery or partial aortic arch replacement, whereas patients with UA underwent off-pump coronary artery bypass grafting (OPCABG).

All adult subjects were over 18 years of age, had been diagnosed with AA (chronic onset), UA and ATAAD (with a time from onset of under two weeks and had been scheduled for emergency surgery). The exclusion criteria were as follows: patients with neoplastic or chronic coagulation and inflammatory disorders; patients with missing specimens or clinical data; and patients who experienced a failed surgical intervention or died within 24 hours after surgery. Clinical data of these patients were collected during the hospitalization period. In addition, nine patients in each group were randomly selected and venous plasma was collected and subjected to enzyme-linked immunosorbent assays (ELISAs) to quantify various markers of coagulation and inflammation. We randomly selected 9 patients from each group by using a random number generator to ensure the reproducibility and validity of the findings.

### 2.2 Definitions

All subjects underwent contrast-enhanced computed tomography (CT) to confirm the surgeons' diagnoses of ATAAD [23]. Based on the European Society of Intensive Care Medicine (ESICM) definition [5], ARDS was confirmed via arterial blood gas analysis and chest radiography. These criteria included an acute onset, an oxygenation index (partial pressure of oxygen in arterial blood/fraction of inspired oxygen, PaO<sub>2</sub>/FiO<sub>2</sub>) <300 mmHg for ARDS, regardless of the ventilator settings, the appearance of bilateral pulmonary infiltration on chest radiography, and the presence of respiratory failure that could not be fully explained by cardiac failure or fluid overload. Hypertension (HTN) is defined as having a systolic blood pressure (SBP) of ≥140 mmHg and/or a diastolic blood pressure (DBP) of >90 mmHg when no antihypertensive drugs are used. The diagnosis of diabetes is based on the recommended standards of the American Diabetes Association (ADA) in 2010. Acute kidney injury (AKI) was confirmed by the guideline published by the Kidney Disease Improving Global Outcomes (KIDGO) in 2012. Heart failure (HF) was defined as left ventricular ejection fraction ≤40% accompanied by symptoms and/or signs. Chronic lung diseases (CLD) include chronic obstructive pulmonary disease, bronchiectasis, asthma or tuberculosis, etc. Neurological disease (ND) refer to ischemic/hemorrhagic stroke, brain tumors or brain trauma. Altered consciousness was defined as drowsiness, stupor or coma that occurs before surgery due to various reasons. Chest radiography was performed daily, and the results were interpreted by two radiologists. Blood gas levels were measured every 4 hours. The primary outcome variable was the development of ARDS within two days after surgery.

### 2.3 Anesthesia and Surgery

Radial artery pressure, dorsalis pedis artery pressure, and central venous pressure were monitored before surgery in all patients using established methods. Anesthesia was administered in accordance with institutional standards. Aortic surgical procedures were performed under general anesthesia, CPB, deep hypothermic circulatory arrest (DHCA), and selective cerebral perfusion were conducted using a modified elephant trunk technique (Sun's procedure) as previously described [22].

### 2.4 Clinical Data and Blood Samples

Data related to clinical characteristics, demographic information, medical history, intraoperative variables, and details pertaining to the subsequent course of recovery in the intensive care unit (ICU) post-surgery were recorded. Each patient's preoperative risk profile was evaluated using the European System for Cardiac Operative Risk Evaluation (EuroSCORE) [24]. The dosage of vasoactive drug administration and organ function were assessed during the first 48 h post-surgery using the vasoactive inotrope score (VIS) [25] and the Sequential Organ Failure Assessment (SOFA) scale [26], respectively.

Blood samples were collected in sodium citrate tubes through a central venous catheter at the following three time points: immediately prior to anesthesia induction; immediately after surgery; and 24 hours after surgery. Samples were centrifuged at  $1550\times g$  for 15 min at 4 °C, and the supernatants were aliquoted and stored at –80 °C for later analysis. Serum levels of factor XII (FXII), factor XIIa (FXIIa), factor VIII-related antigen (FVIII-Ag), plasminantiplasmin complex (PAP), interleukin one beta (IL-1 $\beta$ ), and tumor necrosis factor-alpha (TNF $\alpha$ ) were assayed using ELISA kits (USCN KIT INC, Wuhan, China).

### 2.5 Statistical Analysis

We used the median to interpolate the missing data of continuous variables and conducted the Kolmogorov-Smirnov test. Continuous variables were expressed as the mean  $\pm$  standard deviation and were analyzed using a Student's *t*-test for normally distributed data; alternatively,



Table 1. Baseline among three groups.

Variables	ATAAD	AA	UA	$^{1}p$	$^2p$	$^{3}p$			
variables	(n = 150)	(n = 150)	(n = 150)	P	P				
Demographic information and preoperative complications									
Age (yrs)	$49.1 \pm 11.5$	$55.3\pm13.5$	$62.8 \pm 8.1$	< 0.001	< 0.001	< 0.001			
Sex (male%)	114 (76.0)	109 (72.7)	83 (55.3)	0.597	< 0.001	< 0.001			
BMI $(kg/m^2)$	$27.2 \pm 3.8$	$25.6 \pm 3.5$	$25.6\pm3.1$	< 0.001	< 0.001	< 0.001			
Smoking	65 (43.3)	72 (48.0)	49 (32.7)	0.487	0.074	0.022			
Alcohol	19 (12.7)	42 (28.0)	33 (22.0)	0.002	0.047	0.004			
HTN	126 (84.0)	94 (62.7)	90 (60.0)	< 0.001	< 0.001	< 0.001			
Diabetes	13 (8.7)	9 (6.0)	57 (38.0)	0.506	< 0.001	< 0.001			
AKI	23 (15.3)	7 (4.7)	5 (3.3)	0.004	0.001	< 0.001			
LD	11 (7.3)	7 (4.7)	4 (2.7)	0.466	0.112	0.171			
HF	49 (32.7)	57 (38.0)	88 (58.7)	0.398	< 0.001	< 0.001			
CLD	4 (2.7)	2 (1.3)	4 (2.7)	0.680	1	0.664			
ND	1 (0.7)	4 (2.7)	4 (2.7)	0.367	0.367	0.360			
Prior CS	2 (1.3)	13 (8.7)	1 (0.7)	0.008	1	< 0.001			
Shock	7 (4.7)	0 (0.0)	0 (0.0)	0.022	0.022	0.001			
AC	15 (10.0)	1 (0.7)	0 (0.0)	0.001	< 0.001	< 0.001			
EuroSCORE	5.0 (5.0, 6.0)	4.0 (3.3, 6.0)	2.0 (1.0, 3.0)	< 0.001	< 0.001	< 0.001			
Preoperative laborator	y values								
PaCO <sub>2</sub> (mmHg)	36.7 (33.3, 39.7)	34.6 (32.3, 36.8)	34.6 (31.7, 36.8)	< 0.001	< 0.001	< 0.001			
PaO <sub>2</sub> (mmHg)	98.1 (76.4, 122.8)	89.7 (81.9, 95.0)	90.9 (80.9, 101.1)	0.015	0.092	0.034			
Lac (mmol/L)	1.5 (1.0, 2.3)	1.5 (1.1, 1.8)	1.7 (1.4, 2.1)	0.338	0.113	0.004			
PT	12.0 (11.5, 12.6)	11.4 (10.9, 12.1)	11.2 (10.8, 11.6)	< 0.001	< 0.001	< 0.001			
APTT	30.3 (28.5, 32.3)	31.4 (29.6, 33.8)	31.0 (29.0, 33.2)	0.001	0.069	0.004			
D-dimer (ng/mL)	2589.0 (978.5, 7919.0)	180.0 (68.3, 670.8)	92.5 (56.0, 157.0)	< 0.001	< 0.001	< 0.001			
FBG (g/L)	2.3 (1.8, 2.9)	2.9 (2.4, 3.4)	3.1 (2.6, 3.6)	< 0.001	< 0.001	< 0.001			
FDPs ( $\mu g/mL$ )	27.5 (10.9, 69.3)	1.4 (0.5, 4.6)	0.8 (0.5, 1.2)	< 0.001	< 0.001	< 0.001			

 $^1p$ , p value of ATAAD group vs AA group;  $^2p$ , p value of ATAAD group vs UA group;  $^3p$ , p value of one-way ANOVA among three groups; ATAAD, acute type A aortic dissection; AA, aortic aneurysm; UA, unstable angina; BMI, body mass index; PT, prothrombin time; APTT, activated partial thromboplastin time; HTN, hypertension; AKI, acute kidney injury; LD, liver dysfunction; HF, heart failure; CLD, chronic lung disease; CS, cardiac surgery; ND, neurological dysfunction; AC, altered consciousness; EuroSCORE, European System for Cardiac Operative Risk Evaluation; Lac, lactic acid; FBG, fibrinogen; FDPs, fibrinogen degradation products.

 $^{1}p$  and  $^{2}p < 0.017$  indicates statistical significance;  $^{3}p < 0.05$  indicates statistical significance. The bolded data indicate statistical significance.

variables with a skewed distribution were expressed as medians and quartiles and were analyzed using a Mann-Whitney U-test. Categorical variables were expressed as numbers and percentages, and a  $\chi^2$  test or Fisher's exact test was used to compare groups as appropriate.

The baseline characteristics of the ATAAD group were first compared with those of the AA group and UA group separately, followed by ANOVA across all three groups with Bonferroni correction for multiple comparisons. Patients with ATAAD were further stratified into ARDS and Non-ARDS subgroups, with a comparative analysis conducted on their baseline characteristics. All coagulation biomarkers were normalized and standardized using logand z-score transformations. Logistic regression models were applied separately in the ATAAD group, AA group and UA group to assess the associations between coagula-

tion biomarkers and postoperative ALI/ARDS, with odds ratios (ORs) and 95% confidence intervals (95% CIs) calculated. Model 1 was a univariate model and Model 2 was adjusted for traditional confounders, including sex, age, body mass index (BMI), HTN, diabetes, AKI, HF, liver dysfunction (LD), CLD, ND, smoking status, alcohol consumption, prior CS, preoperative aspartate aminotransferase (AST) level, and the preoperative oxygenation index (PaO<sub>2</sub>/FiO<sub>2</sub>, P/F). These covariates include medical history data and some preoperative meaningful indicators (AST and P/F) between the ARDS group and Non-ARDS group in the baseline characteristics (Supplementary Tables 1,2). Collinearity test was conducted in the multivariate analysis (Supplementary Table 3). A two-way ANOVA was used to account for both group and time as independent variables in ELISA. Spearman correlation analysis was used to ana-



Table 2. Intraoperative and postoperative clinical factors among three groups.

Surgery-related variables           Surgery-related variation (h)         8.0 (7.0, 9.0)         7.1 (6.0, 9.0)         4.5 (4.0, 5.0)         0.001         < 0.001		ATAAD	AA	UA	1 p	$^2p$	<sup>3</sup> p	
Surgical duration (h)         8.0 (7.0, 9.0)         7.1 (6.0, 9.0)         4.5 (4.0, 5.0)         0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001	Variables	(n = 150)	(n = 150)	(n = 150)	- p	<i>-p</i>	~ <i>p</i>	
Surgical duration (h)         8.0 (7.0, 9.0)         7.1 (6.0, 9.0)         4.5 (4.0, 5.0)         0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001	Surgery-related variables							
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		8.0 (7.0, 9.0)	7.1 (6.0, 9.0)	4.5 (4.0, 5.0)	0.001	< 0.001	< 0.001	
ACC time (min)         96.5 (86.0, 116.0)         86.5 (66.3, 100.8)         0.0 (0.0, 0.0)         <0.001         <0.001         <0.001           Heparin input (mL)         1.0 (10.0, 10.0)         1.0 (7.6, 10.0)         0.0 (0.0, 0.0)         0.065         <0.001	CPB time (min)	179.0 (158.0, 206.0)	147.5 (123.0, 172.8)	0.0(0.0, 0.0)	< 0.001	< 0.001	< 0.001	
Heparin input (mL)	DHCA time (min)	20.0 (13.0, 30.0)	0.0 (0.0, 0.0)	0.0(0.0, 0.0)	< 0.001	< 0.001	< 0.001	
Plasma input (mL)         0.0 (0.0, 400.0)         0.0 (0.0, 400.0)         0.0 (0.0, 0.0)         0.658         <0.001         <0.001           Postoperative laboratory values         HR         89.0 (75.0, 98.0)         84.0 (75.0, 89.8)         79.0 (70.0, 86.0)         0.032         <0.001	ACC time (min)	96.5 (86.0, 116.0)	86.5 (66.3, 100.8)	0.0(0.0, 0.0)	< 0.001	< 0.001	< 0.001	
Postoperative laboratory values         Postoperative laboratory values         Postoperative laboratory values         Postoperative laboratory values         Co.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001         <0.001	Heparin input (mL)	10.0 (10.0, 10.0)	10.0 (7.6, 10.0)	0.0(0.0, 0.0)	0.005	< 0.001	< 0.001	
HR   89.0 (75.0, 98.0)   84.0 (75.0, 89.8)   79.0 (70.0, 86.0)   0.032   <0.001   <0.001   MAP (mm/Hg)   92.8 (83.0, 102.9)   80.0 (75.1, 86.7)   83.5 (74.3, 91.0)   <0.001   <0.001   <0.001   <0.001   <0.001   ACT   160.0 (147.0, 176.0)   151.0 (141.3, 159.0)   153.0 (144.3, 163.0)   <0.001   <0.001   <0.001   <0.001   PaCO <sub>2</sub> (mm/Hg) 0 h   46.5 (42.6, 50.1)   43.5 (39.1, 46.1)   39.5 (36.2, 42.4)   <0.001   <0.001   <0.001   PaCO <sub>2</sub> (mm/Hg) 24 h   39.4 (35.4, 42.7)   39.8 (36.5, 42.9)   39.3 (35.6, 41.4)   0.565   0.214   0.169   PaO <sub>2</sub> (mm/Hg) 24 h   39.2 (77.6, 115.8)   118.0 (95.1, 149.8)   127.0 (97.2, 153.0)   <0.001   <0.001   <0.001   Lac (mmol/L) 0 h   2.4 (1.5, 3.5)   1.8 (1.3, 2.6)   1.2 (0.9, 1.5)   0.001   <0.001   <0.001   Lac (mmol/L) 24 h   2.2 (16.3.2)   3.2 (1.9, 5.0)   1.8 (1.2, 2.5)   <0.001   <0.001   <0.001   PF 0 h (%)   180.0 (134.1, 264.6)   275.0 (178.3, 375.3)   268.4 (203.7, 348.3)   <0.001   <0.001   <0.001   NE (×10°/L) 24 h   10.0 (7.8, 12.4)   9.3 (7.0, 12.8)   9.7 (80.12.0)   0.573   0.938   0.791   NE (×10°/L) 48 h   13.6 (10.9, 16.5)   14.0 (10.2, 17.4)   11.8 (9.1, 14.9)   0.730   0.001   0.001   PT 0 h   13.2 (12.4, 14.1)   13.0 (12.3, 14.0)   13.5 (12.9, 14.5)   0.345   <0.001   <0.001   APTT 24 h   33.9 (31.0, 40.3)   32.2 (29.2, 36.1)   31.5 (29.7, 35.4)   0.001   0.001   0.001   APTT 24 h   30.4 (27.7, 33.9)   30.8 (28.6, 34.5)   31.0 (28.3, 34.1)   0.143   0.144   0.239   0.404	Plasma input (mL)	0.0 (0.0, 400.0)	0.0 (0.0, 400.0)	0.0(0.0, 0.0)	0.658	< 0.001	< 0.001	
MAP (mm/Hg)         92.8 (83.0, 102.9)         80.0 (75.1, 86.7)         83.5 (74.3, 91.0)         < 0.001         < 0.001         < 0.001           ACT         160.0 (147.0, 176.0)         151.0 (141.3, 159.0)         153.0 (144.3, 163.0)         < 0.001	Postoperative laboratory va	lues						
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	HR	89.0 (75.0, 98.0)	84.0 (75.0, 89.8)	79.0 (70.0, 86.0)	0.032	< 0.001	< 0.001	
$\begin{array}{llllllllllllllllllllllllllllllllllll$	MAP (mm/Hg)	92.8 (83.0, 102.9)	80.0 (75.1, 86.7)	83.5 (74.3, 91.0)	< 0.001	< 0.001	< 0.001	
$\begin{array}{llllllllllllllllllllllllllllllllllll$	ACT	160.0 (147.0, 176.0)	151.0 (141.3, 159.0)	153.0 (144.3, 163.0)	< 0.001	0.001	< 0.001	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	PaCO <sub>2</sub> (mm/Hg) 0 h	46.5 (42.6, 50.1)	43.5 (39.1, 46.1)	39.5 (36.2, 42.4)	< 0.001	< 0.001	< 0.001	
$\begin{array}{llllllllllllllllllllllllllllllllllll$	PaCO <sub>2</sub> (mm/Hg) 24 h	39.4 (35.4, 42.7)	39.8 (36.5, 42.9)	39.3 (35.6, 41.4)	0.565	0.214	0.169	
Lac (mmol/L) 0 h         2.4 (1.5, 3.5)         1.8 (1.3, 2.6)         1.2 (0.9, 1.5)         0.001         < 0.001         < 0.001           Lac (mmol/L) 24 h         2.2 (1.6, 3.2)         3.2 (1.9, 5.0)         1.8 (1.2, 2.5)         < 0.001	$PaO_2 \text{ (mm/Hg) } 0 \text{ h}$	110.0 (85.5, 160.5)	151.5 (104.5, 206.5)	158.0 (120.5, 205.8)	< 0.001	< 0.001	< 0.001	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	PaO <sub>2</sub> (mm/Hg) 24 h	93.2 (77.6, 115.8)	118.0 (95.1, 149.8)	127.0 (97.2, 153.0)	< 0.001	< 0.001	< 0.001	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Lac (mmol/L) 0 h	2.4 (1.5, 3.5)	1.8 (1.3, 2.6)	1.2 (0.9, 1.5)	0.001	< 0.001	< 0.001	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Lac (mmol/L) 24 h	2.2 (1.6, 3.2)	3.2 (1.9, 5.0)	1.8 (1.2, 2.5)	< 0.001	< 0.001	< 0.001	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	P/F 0 h (%)	180.0 (134.1, 264.6)	275.0 (178.3, 375.3)	268.4 (203.7, 348.3)	< 0.001	< 0.001	< 0.001	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	P/F 24 h (%)	210.8 (151.4, 281.3)	289.0 (230.2, 371.3)	295.7 (242.0, 373.6)	< 0.001	< 0.001	< 0.001	
PT 0 h         13.2 (12.4, 14.1)         13.0 (12.3, 14.2)         13.3 (12.5, 14.4)         0.628         0.344         0.354           PT 24 h         13.0 (12.1, 13.9)         13.0 (12.3, 14.0)         13.5 (12.9, 14.5)         0.345         <0.001	NE (×10 <sup>9</sup> /L) 24 h	10.0 (7.8, 12.4)	9.3 (7.0, 12.8)	9.7 (8.0, 12.0)	0.573	0.938	0.791	
PT 24 h APTT 0 h APTT 0 h APTT 24 h APTT 24 h APTT 24 h BOULL (11.0) APTT 24 h BOULL (11.0) APTT 24 h APTT	NE (×10 <sup>9</sup> /L) 48 h	13.6 (10.9, 16.5)	14.0 (10.2, 17.4)	11.8 (9.1, 14.9)	0.730	0.001	0.001	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	PT 0 h	13.2 (12.4, 14.1)	13.0 (12.3, 14.2)	13.3 (12.5, 14.4)	0.628	0.344	0.354	
APTT 24 h       30.4 (27.7, 33.9)       30.8 (28.6, 34.5)       31.0 (28.3, 34.1)       0.143       0.144       0.239         D-dimer (ng/mL) 0 h       2532.0 (1180.8, 4083.5)       987.0 (420.5, 2798.8)       186.0 (112.3, 339.0)       <0.001	PT 24 h	13.0 (12.1, 13.9)	13.0 (12.3, 14.0)	13.5 (12.9, 14.5)	0.345	< 0.001	< 0.001	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	APTT 0 h	33.9 (31.0, 40.3)	32.2 (29.2, 36.1)	31.5 (29.7, 35.4)	0.001	0.001	0.001	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	APTT 24 h	30.4 (27.7, 33.9)	30.8 (28.6, 34.5)	31.0 (28.3, 34.1)	0.143	0.144	0.239	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	D-dimer (ng/mL) 0 h	2532.0 (1180.8, 4083.5)	987.0 (420.5, 2798.8)	186.0 (112.3, 339.0)	< 0.001	< 0.001	< 0.001	
$\begin{array}{llllllllllllllllllllllllllllllllllll$	D-dimer (ng/mL) 24 h	2294.5 (1173.0, 3390.8)	917.5 (469.0, 2651.5)	234.0 (156.3, 416.0)	< 0.001	< 0.001	< 0.001	
$\begin{array}{llllllllllllllllllllllllllllllllllll$	FBG (g/L) 0 h	2.5 (2.0, 3.3)	2.3 (1.9, 2.9)	2.4 (1.8, 3.1)	0.043	0.091	0.091	
$\begin{array}{llllllllllllllllllllllllllllllllllll$	FBG (g/L) 24 h	3.7 (3.0, 4.5)	2.7 (2.3, 3.3)	3.1 (2.6, 3.7)	< 0.001	< 0.001	< 0.001	
VIS 24 h       6.0 (3.0, 12.8)       5.0 (2.0, 11.0)       7.0 (4.0, 12.0)       0.050       0.909       0.070         VIS 48 h       3.0 (0.0, 8.7)       0.0 (0.0, 7.0)       5.0 (0.0, 10.0)       0.001       0.815       0.001         SOFA 24 h       10.0 (7.0, 13.0)       6.0 (5.0, 8.0)       5.0 (4.0, 7.0)       <0.001	FDPs ( $\mu g/mL$ ) 0 h	20.5 (10.3, 38.1)	7.9 (3.0, 21.2)	1.4 (0.9, 2.3)	< 0.001	< 0.001	< 0.001	
VIS 48 h       3.0 (0.0, 8.7)       0.0 (0.0, 7.0)       5.0 (0.0, 10.0)       0.001       0.815       0.001         SOFA 24 h       10.0 (7.0, 13.0)       6.0 (5.0, 8.0)       5.0 (4.0, 7.0)       <0.001	FDPs ( $\mu g/mL$ ) 24 h	22.1 (11.8, 35.3)	8.9 (4.6, 21.9)	1.9 (1.2, 3.3)	< 0.001	< 0.001	< 0.001	
SOFA 24 h       10.0 (7.0, 13.0)       6.0 (5.0, 8.0)       5.0 (4.0, 7.0)       <0.001       <0.001       <0.001         SOFA48 h       8.0 (5.0, 12.8)       5.0 (3.0, 6.0)       4.0 (3.0, 5.0)       <0.001	VIS 24 h	6.0 (3.0, 12.8)	5.0 (2.0, 11.0)	7.0 (4.0, 12.0)	0.050	0.909	0.070	
SOFA48 h 8.0 (5.0, 12.8) 5.0 (3.0, 6.0) 4.0 (3.0, 5.0) <b>&lt;0.001 &lt;0.001 &lt;0.001</b> ARDS (%) 31 (20.7) 20 (13.3) 11 (7.3) 0.124 <b>0.002 0.004</b>	VIS 48 h	3.0 (0.0, 8.7)	0.0 (0.0, 7.0)	5.0 (0.0, 10.0)	0.001	0.815	0.001	
ARDS (%) 31 (20.7) 20 (13.3) 11 (7.3) 0.124 <b>0.002 0.004</b>	SOFA 24 h	10.0 (7.0, 13.0)	6.0 (5.0, 8.0)	5.0 (4.0, 7.0)	< 0.001	< 0.001		
	SOFA48 h		5.0 (3.0, 6.0)	4.0 (3.0, 5.0)				
ICU duration (b) $68.5 (40.0 - 151.8)$ $42.0 (10.0 -60.0)$ $25.5 (10.0 -49.0)$ $< 0.001 < 0.001$			` /	` /	0.124	0.002	0.004	
1CU duration (n) 68.5 (40.0, 151.8) 42.0 (19.0, 69.0) 25.5 (19.0, 48.0) < 0.001 < 0.001	ICU duration (h)	68.5 (40.0, 151.8)	42.0 (19.0, 69.0)	25.5 (19.0, 48.0)	< 0.001	< 0.001	< 0.001	

<sup>1</sup>p, p value of ATAAD group vs AA group; <sup>2</sup>p, p value of ATAAD group vs UA group; <sup>3</sup>p, p value of one-way ANOVA among three groups; CPB, cardiopulmonary bypass; DHCA, deep hypothermic circulatory arrest; ACC, aortic cross-clamp; HR, heart rate at admission in intensive care unit (ICU); MAP, mean arterial pressure at admission in ICU; ACT, activated clotting time at admission in ICU; Lac, lactic acid; P/F, PaO<sub>2</sub>/FiO<sub>2</sub>, oxygenation index; NE, neutrophil count; VIS, vasoactive inotrope score; SOFA, Sequential Organ Failure Assessment; 0 h, Patients at admission in ICU after surgery; 24 h, Patients in ICU at 24 h after surgery.

lyze the associations between variables for inferring coagulation activity and the oxygenation index.

All analyses were performed using R software 4.3.1 (R Foundation, Vienna, Austria) (https://www.r-project.org/) and Prism 10.2 (GraphPad Corp, San Diego, CA, USA). p-value < 0.05 indicates statistical significance (p < 0.017 indicates statistical significance after Bonferroni correction).

### 3. Results

## 3.1 Subject Preoperative Data

Baseline and preoperative laboratory values of the participants are presented in Table 1. Patients in the ATAAD group were younger than those in the AA group (49.1  $\pm$  11.5 years), whereas the patients with ATAAD exhibited a higher incidence of elevated blood pressure (84% vs 62.7%,



 $<sup>^{1}</sup>p$  and  $^{2}p < 0.017$  indicates statistical significance;  $^{3}p < 0.05$  indicates statistical significance. The bolded data indicate statistical significance.

Table 3. Baseline among ARDS and non-ARDS patients within ATAAD cohort.

Variables	Non-ARDS	ARDS	p					
variables	(n = 119)	(n = 31)						
Demographic info	rmation and preoperative of	complications						
Age (yrs)	$48.5 \pm 12.0$	$51.6 \pm 9.1$	0.179					
Sex (male%)	87 (73.1)	27 (87.1)	0.165					
BMI $(kg/m^2)$	$27.1 \pm 3.8$	$27.6 \pm 3.8$	0.526					
Smoking	51 (42.9)	14 (45.2)	0.978					
Alcohol	14 (11.8)	5 (16.1)	0.728					
HTN	101 (84.9)	25 (80.6)	0.766					
Diabetes	10 (8.4)	3 (9.7)	1					
AKI	22 (18.5)	1 (3.2)	0.069					
LD	10 (8.4)	1 (3.2)	0.550					
HF	37 (31.1)	12 (38.7)	0.555					
CLD	3 (2.5)	1 (3.2)	1					
ND	1 (0.8)	0 (0.0)	1					
Prior CS	1 (0.8)	1 (3.2)	0.879					
Shock	5 (4.2)	2 (6.5)	0.959					
AC	12 (10.1)	3 (9.7)	1					
EuroSCORE	5.0 (5.0, 6.5)	5.0 (5.0, 6.0)	0.622					
Preoperative labor	Preoperative laboratory values							
$PaCO_2$	36.7 (33.3, 39.6)	35.6 (33.4, 39.4)	0.492					
$PaO_2$	100.0 (80.7, 123.5)	92.6 (69.8, 111.0)	0.138					
Lac	1.4 (1.0, 2.2)	1.9 (1.2, 3.0)	0.094					
PT	12.0 (11.5, 12.6)	11.9 (11.3, 12.7)	0.922					
APTT	30.2 (28.6, 32.2)	31.1 (26.9, 32.4)	0.873					
D-dimer	2403.0 (942.0, 7524.5)	5302.0 (1184.0, 9419.0)	0.134					
FBG	2.3 (1.8, 3.0)	2.0 (1.7, 2.5)	0.100					
FDPs	22.7 (10.2, 65.9)	49.3 (12.5, 80.8)	0.098					

p < 0.05 indicates statistical significance. The bolded data indicate statistical significance.

respectively). Preoperatively, patients with ATAAD presented with significantly higher D-dimer levels than did those in either the AA group (2589 [978.5–7919] vs 180 [68.3–670.8] ng/mL; p < 0.001) or the UA group (2589 [978.5–7919] vs 92.5 [56.0–157.0] ng/mL; p < 0.001). The levels of FDPs in the ATAAD group increased significantly at admission (27.5 [10.9–69.3] vs 1.4 [0.5–4.6] µg/mL, p < 0.001; 27.5 [10.9–69.3] vs 0.8 [0.5–1.2] µg/mL, p < 0.001). However, the levels of fibrinogen (FBG) in the ATAAD group decreased significantly before surgery (2.3 [1.8–2.9] vs 2.9 [2.4–3.4] g/L, p < 0.001; 2.3 [1.8–2.9] vs 3.1 [2.6–3.6] g/L, p < 0.001).

### 3.2 Comparison of Coagulation Biomarker Levels Between the ATAAD, AA, and UA Groups

The duration of the CPB procedure was significantly longer in the patients with ATAAD (179 [158–206] min) than it was in the other two groups (p < 0.001 for ATAAD vs. AA and for ATAAD vs. UA), as was the aortic cross-clamping time (96.5 [86–116] min; p < 0.001 for ATAAD vs. AA and for ATAAD vs. UA). Immediately after surgery, the PaO<sub>2</sub> values (110.0 [85.5–160.5] vs. 151.5

[141.3–206.5] mmHg, ATAAD vs. AA group, p < 0.001; vs. 158.0 [120.5–205.8] mmHg, ATAAD vs UA group, p < 0.001) decreased and lactic acid (Lac) level increased more significantly in the ATAAD group than in the other two groups. The activated partial thromboplastin time (APTT) at the end of surgery was longer (33.9 [31.0–40.3] s) in the patients with ATAAD than that in the other groups (p = 0.001 for ATAAD vs. AA and for ATAAD vs UA). The D-dimer level in the ATAAD group increased significantly over the 24 hours post-surgery. Similarly, the trend in the levels of FDPs was consistent with that of the D-dimer levels (Table 2). More importantly, ARDS represented 13.8% of total patients and 20.7% of patients with ATAAD.

## 3.3 Coagulation Activity in the Patients With and Without ARDS in the ATAAD Cohort

The levels of coagulation factors in the patients with ARDS from the ATAAD group are shown in Tables 3,4. Preoperative D-dimer levels were similar between the non-ARDS and ARDS groups (2403.0 [942.0–7524.5] vs 5302 [1184.0–9419.0] ng/mL, p=0.134). The concentration of the FDPs also tended to rise in the ARDS group before



Table 4. Intraoperative and postoperative clinical factors among ARDS and non-ARDS patients within ATAAD cohort.

Variables	Non-ARDS	ARDS	- <i>p</i>					
variables	(n = 119)	(n = 31)						
Surgery-related variables								
Surgical duration (h)	8.0 (7.0, 9.0)	8.0 (7.5, 9.3)	0.086					
CPB time (min)	176.0 (158.0, 202.0)	192.0 (162.0, 222.5)	0.078					
DHCA time (min)	19.0 (12.5, 29.0)	24.0 (15.0, 36.5)	0.055					
ACC time (min)	94.0 (84.5, 112.5)	113.0 (93.5, 122.5)	0.019					
Heparin input (mL)	10.0 (10.0, 10.0)	10.0 (10.0, 11.0)	0.431					
Plasma input (mL)	0.0 (0.0, 400.0)	0.0(0.0, 0.0)	0.184					
Postoperative laboratory value	es							
HR	88.0 (75.0, 98.0)	90.0 (79.5, 103.5)	0.342					
MAP (mm/Hg)	94.0 (83.4, 103.8)	90.5 (78.3, 100.3)	0.256					
ACT	161.0 (147.0, 178.0)	160.0 (149.0, 173.0)	0.705					
$PaCO_2 \ 0 \ h$	46.5 (42.7, 50.0)	47.3 (42.4, 53.1)	0.462					
PaCO <sub>2</sub> 24 h	39.4 (35.5, 43.3)	39.4 (34.8, 42.2)	0.279					
$PaO_2 \ 0 \ h$	110.0 (85.6, 161.5)	109.0 (86.6, 141.0)	0.876					
$PaO_2$ 24 h	94.5 (78.6, 117.5)	90.6 (72.3, 110.5)	0.393					
Lac 0 h	2.3 (1.5, 3.3)	2.6 (1.8, 4.0)	0.224					
Lac 24 h	2.2 (1.5, 3.2)	2.2 (1.9, 3.2)	0.202					
P/F 0 h	182.8 (131.1, 273.4)	162.3 (137.3, 235.4)	0.598					
P/F 24 h	213.8 (158.6, 298.8)	188.3 (122.4, 265.0)	0.154					
P/F 48 h	275.7 (193.3, 333.4)	187.3 (129.7, 218.2)	< 0.001					
NE 24 h	10.2 (7.8, 12.4)	9.5 (7.3, 12.3)	0.590					
NE 48 h	13.9 (11.2, 16.5)	12.3 (10.7, 16.4)	0.415					
PT 0 h	13.2 (12.4, 14.0)	13.3 (12.6, 14.8)	0.288					
PT 24 h	13.0 (12.2, 14.0)	13.1 (12.2, 13.7)	0.688					
PT 48 h	12.3 (11.6, 13.0)	12.5 (11.7, 13.5)	0.328					
APTT 0 h	33.9 (31.0, 40.1)	33.6 (31.9, 40.3)	0.897					
APTT 24 h	30.4 (27.3, 33.7)	30.7 (28.1, 35.7)	0.426					
APTT 48 h	31.0 (27.8, 31.4)	31.0 (27.4, 33.7)	0.825					
D-dimer 0 h	2281.0 (1135.0, 3787.5)	3326.0 (2270.0, 5989.0)	0.017					
D-dimer 24 h	2222.0 (1095.0, 3358.5)	2823.0 (1881.5, 3558.5)	0.213					
D-dimer 48 h	935.0 (869.5, 2617.0)	2017.0 (935.0, 2955.0)	0.046					
FBG 0 h	2.7 (2.1, 3.6)	2.3 (1.6, 2.9)	0.013					
FBG 24 h	3.7 (2.9, 4.5)	3.7 (3.3, 4.5)	0.809					
FBG 48 h	4.2 (3.8, 4.6)	4.2 (3.3, 4.9)	0.696					
FDPs 0 h	16.3 (9.7, 33.3)	33.5 (17.4, 57.9)	0.008					
FDPs 24 h	20.8 (11.3, 33.8)	29.6 (17.8, 37.2)	0.128					
FDPs 48 h	8.9 (8.5, 23.6)	15.4 (8.8, 28.1)	0.037					
VIS 24 h	6.0 (3.0, 11.0)	10.0 (3.5, 20.0)	0.157					
VIS 48 h	3.0 (0.0, 8.0)	5.0 (3.0, 11.0)	0.054					
SOFA 24 h	10.0 (7.0, 13.0)	11.0 (8.5, 15.0)	0.069					
SOFA 48 h	7.0 (5.0, 11.0)	10.0 (8.0, 14.5)	0.001					
MV duration (h)	19.0 (12.0, 61.0)	39.0 (20.0, 108.5)	0.014					
ICU duration (h)	65.0 (37.0, 138.0)	106.0 (64.5, 201.0)	0.027					
In-hospital mortality (%)	5 (4.2)	8 (25.8)	0.001					
MV Machanical vantilation: 0 h. Patients at admission in ICU after surgery: 24 h. Patients in								

MV, Mechanical ventilation; 0 h, Patients at admission in ICU after surgery; 24 h, Patients in ICU at 24 h after surgery; 48 h, Patients in ICU at 48 h after surgery.

surgery (22.7 [10.2–65.9] vs 49.3 [12.5–80.8]  $\mu$ g/mL, p = 0.098). At the end of surgery, D-dimer levels in the ARDS group were significantly higher than those in the non-ARDS

group (3326 [2270.0–5989.0] vs 2281.0 [1135.0–3787.5] ng/mL, p = 0.017). Twenty-four hours after surgery, there was no significant difference between the groups, although



p < 0.05 indicates statistical significance. The bolded data indicate statistical significance.



Table 5. Correlation between coagulation activity and postoperative ARDS in patients in three groups.

	ATAAD				AA			UA				
Variables	Model 1		Model 2		Model 1 Mo		Model 2	lel 2 Model 1		Model 2		
	OR (95% CI)	p	OR (95% CI)	p	OR (95% CI)	p						
z-pre-PT	0.93 (0.58, 1.49)	0.756	0.87 (0.45, 1.65)	0.661	1.07 (0.77, 1.49)	0.698	1.42 (0.88, 2.28)	0.151	0.38 (0.10, 1.39)	0.143	0.39 (0.08, 2.00)	0.261
z-pre-APTT	1.02 (0.70, 1.50)	0.900	1.02 (0.66, 1.57)	0.942	0.81 (0.39, 1.70)	0.577	1.41 (0.55, 3.61)	0.474	0.85 (0.43, 1.68)	0.638	0.76 (0.21, 2.72)	0.668
z-pre-D-dimer	1.51 (0.89, 2.59)	0.130	1.39 (0.72, 2.69)	0.328	1.21 (0.67, 2.16)	0.525	1.19 (0.61, 2.33)	0.608	1.63 (0.52, 5.17)	0.403	3.81 (0.56, 25.8)	0.170
z-pre-FBG	0.75 (0.51, 1.09)	0.130	0.76 (0.48, 1.20)	0.237	0.96 (0.60, 1.54)	0.863	0.85 (0.46, 1.56)	0.594	1.67 (0.55, 5.05)	0.364	5.04 (0.69, 36.5)	0.110
z-pre-FDPs	1.54 (0.95, 2.51)	0.083	1.41 (0.78, 2.55)	0.252	1.16 (0.60, 2.23)	0.659	0.97 (0.44, 2.16)	0.943	1.67 (0.41, 6.75)	0.472	18.8 (1.08, 32.8)	0.044
z-PT 0 h	1.05 (0.75, 1.47)	0.782	1.09 (0.76, 1.57)	0.642	1.84 (0.82, 4.14)	0.139	2.38 (0.89, 6.36)	0.085	0.84 (0.34, 2.06)	0.705	1.31 (0.36, 4.85)	0.684
z-PT 24 h	1.08 (0.72, 1.47)	0.719	1.42 (0.87, 2.31)	0.161	1.13 (0.68, 1.87)	0.636	1.33 (0.70, 2.50)	0.384	0.87 (0.47, 1.62)	0.670	0.82 (0.36, 1.86)	0.629
z-PT 48 h	1.25 (0.86, 1.81)	0.236	1.35 (0.87, 2.09)	0.175	/		/		/		/	
z-APTT 0 h	1.02 (0.63, 1.65)	0.951	1.07 (0.61, 1.87)	0.820	1.15 (0.71, 1.87)	0.558	1.04 (0.58, 1.88)	0.893	1.28 (0.59, 2.78)	0.540	0.97 (0.37, 2.55)	0.958
z-APTT 24 h	1.36 (0.84, 2.21)	0.210	1.41 (0.78, 2.54)	0.257	0.77 (0.33, 1.81)	0.551	0.49 (0.19, 1.23)	0.129	1.16 (0.73, 1.82)	0.530	0.99 (0.45, 2.17)	0.980
z-APTT 48 h	1.00 (0.67, 1.48)	0.991	1.02 (0.66, 1.57)	0.933	/		/		/		/	
z-D-dimer 0 h	1.95 (1.06, 3.59)	0.032	2.03 (0.96, 4.28)	0.063	2.04 (1.11, 3.50)	0.008	1.63 (0.89, 2.98)	0.111	1.62 (0.59, 4.43)	0.346	2.82 (0.53, 15.2)	0.230
z-D-dimer 24 h	1.26 (0.68, 2.33)	0.455	1.24 (0.63, 2.47)	0.533	1.73 (0.99, 3.01)	0.053	1.41 (0.72, 2.74)	0.319	1.59 (0.63, 4.02)	0.362	1.95 (0.65, 5.88)	0.234
z-D-dimer 48 h	1.57 (1.06, 2.32)	0.026	1.55 (0.97, 2.47)	0.066	/		/		/		/	
z-FBG 0 h	0.43 (0.23, 0.81)	0.008	0.40 (0.19, 0.83)	0.014	0.58 (0.26, 1.32)	0.196	0.46 (0.17, 1.22)	0.117	1.05 (0.74, 1.50)	0.768	1.10 (0.50, 2.44)	0.814
z-FBG 24 h	1.02 (0.63, 1.65)	0.934	0.99 (0.59, 1.68)	0.978	0.64 (0.33, 1.23)	0.181	0.60 (0.28, 1.29)	0.189	1.18 (0.67, 2.09)	0.559	0.99 (0.46, 2.11)	0.979
z-FBG 48 h	0.88 (0.60, 1.30)	0.521	0.85 (0.51, 1.39)	0.510	/		/		/		/	
z-FDPs 0 h	2.05 (1.17, 3.60)	0.012	2.26 (1.13, 4.54)	0.022	1.92 (1.19, 3.09)	0.008	1.60 (0.91, 2.81)	0.105	1.12 (0.27, 4.65)	0.875	1.83 (0.22, 15.3)	0.575
z-FDPs 24 h	1.45 (0.78, 2.69)	0.235	1.42 (0.70, 2.84)	0.329	1.56 (0.96, 2.55)	0.075	1.40 (0.74, 2.65)	0.302	1.79 (0.61, 5.25)	0.287	2.37 (0.52, 10.8)	0.265
z-FDPs 48 h	1.50 (1.02, 2.21)	0.040	1.51 (0.95, 2.40)	0.080	/		/		/		/	

z: All biomarkers were normalized and standardized by log transformation and z-score transformation.

Model 1: Univariate logistic regression.

Model 2: Multivariate logistic regression. Adjusted covariates: age, sex, BMI, Hypertension, Diabetes, AKI, LD, CLD, ND, HF, Smoking, Alcohol, Prior CS, preoperative AST, preoperative oxygenation index. These covariates that showed significant differences between the ARDS group and Non-ARDS group in the baseline characteristics (**Supplementary Table 1**). Pre, preoperative.

p < 0.05 indicates statistical significance. The bolded data indicate statistical significance.

the D-dimer levels were significantly elevated in the ARDS group 48 hours after surgery (2017 [935.0–2955.0] vs. 935.0 [869.5–2617.0] ng/mL, p=0.046). The trend of FDPs levels was consistent with that of the D-dimer levels. In-hospital mortality was significantly higher postoperatively in the patients with ARDS (25.8% vs. 4.2%, p=0.001).

After all coagulation biomarkers were normalized and standardized using log- and z-score transformations, the results indicated that the levels of FDPs at the end of surgery (OR: 2.26, 95% CI: 1.13–4.54; p = 0.022) was the independent risk factor for the development of ARDS among the patients with ATAAD (Table 5).

### 3.4 Serum Concentrations of Coagulation Biomarkers

The effects of the different groups and phases on the concentrations of FXII, FXIIa, FVIII-Ag, and PAP are presented in Fig. 1. Preoperatively (immediately prior to anesthesia induction), compared with those in the other two groups, patients with ATAAD presented with significantly higher levels of FXIIa (p=0.014 and p<0.0001 for ATAAD vs. AA and for ATAAD vs. UA groups, respectively) and FVIII-Ag (p=0.0002 and p<0.0001 for ATAAD vs. AA and for ATAAD vs. UA groups, respectively). After surgery, FXII, FXIIa, and FVIII-Ag levels were significantly higher in the ATAAD group compared to those in the other groups (Fig. 1a–c), whereas the perioperative PAP levels were similar between the ATAAD and AA groups (p>0.05, Fig. 1d).

After conducting statistical analysis on the same indicators of the same group at different time points, it was found that the FXII level at 0 h after was significantly higher than that immediately prior to anesthesia induction (p < 0.001, Fig. 2a) and the TNF level immediately prior to anesthesia induction was significantly higher than that after surgery (p < 0.001, Fig. 2f) in patients with ATAAD. The level of PAP at immediately prior to anesthesia induction was significantly higher than that after surgery (p < 0.05, Fig. 2d) in patients with AA. The levels of other biomarkers did not show significant differences at each time point (Fig. 2).

There was a negative correlation between FXII levels and the oxygenation index 24 h post-surgery (Pearson correlation coefficient r = -0.682, p = 0.043), whereas no significant correlations were observed for the other markers or at other time points (Fig. 3). However, a trend was observed in which the oxygenation index decreased as FVIII-Ag levels increased, suggesting that FVIII-Ag may affect lung function.

### 4. Discussion

The aim of this study was to investigate if coagulation activity could be induced by ATAAD onset or CPB, and to determine the impact of coagulation activity on the development of post-operative ARDS. The results revealed sig-

nificantly elevated serum concentrations of perioperative coagulation biomarkers in patients with ATAAD who had undergone surgery involving CPB. Meanwhile, the results indicate that the elevation of coagulation biomarkers is a risk factor for the occurrence of ARDS.

## 4.1 Activation of Coagulation System in Patients With ATAAD

The present study, which used a control group of patients with AA, verified that ATAAD itself can significantly increase the activation of the coagulation system. ATAAD is caused by injury to the aortic intima, where blood gains access to the false lumen formed between the intima and media. The activation of the coagulation system could be induced by contact between blood and the false lumen and the release of coagulant material from the aortic wall into the systemic circulation [26,27].

Previous studies [28,29] have demonstrated that ATAAD and the surgical interventions to treat it can cause an increase in the levels of biomarkers that induce the activation of coagulation. The preoperative levels of FDPs and D-dimer become sharply elevated in patients with ATAAD [27,30]. An increased D-dimer level reflects both the augmented formation of fibrin as well as the degree of subsequent fibrinolysis, and it has been shown to be associated with postoperative adverse events in patients with ATAAD [31,32]. However, a small number of studies [33,34] have compared the perioperative coagulation function of patients with ATAAD undergoing emergency surgery and that of patients with AA undergoing elective aortic surgery.

In the present study, intergroup comparisons were performed between patients with ATAAD and those with AA or UA; the results showed that the preoperative (immediately prior to anesthesia induction) levels of FXIIa and FVIII-Ag were significantly higher in the ATAAD group than in the other two groups, whereas the preoperative fibrinogen level was significantly lower, indicating that patients with ATAAD consumed more fibrinogen before surgery. Although patients experiencing chronic aneurysms also presented with higher D-dimer levels, the upregulation was more pronounced in patients with ATAAD. However, whether there is a causal relationship between ATAAD itself and ARDS still requires further research to verify.

# 4.2 Activation of Coagulation System in Patients After CPB Surgery

In this study, the levels of FXII, FXIIa, FVIII-Ag, D-dimer, and FDPs were elevated in patients with ATAAD and AA, and the elevation was more pronounced in the former group. Meanwhile, the CPB duration was significantly prolonged in ATAAD group. We hypothesize that the coagulation disorder after CPB is related to the activation of the "contact system".



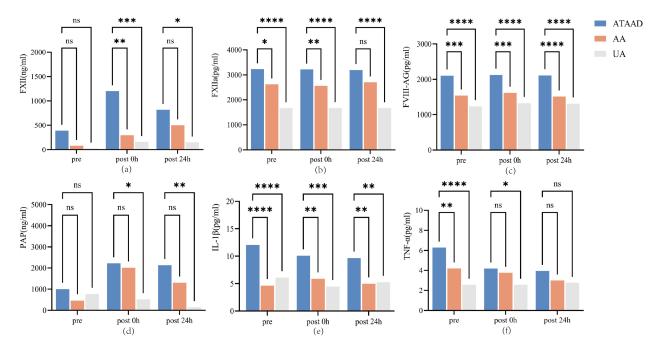


Fig. 1. The serum concentration of coagulation and inflammation biomarkers among three groups. (a) FXII levels: Intergroup comparisons across time points; (b) FXIIa levels: Intergroup comparisons across time points; (c) FVIII-Ag levels: Intergroup comparisons across time points; (e) IL-1 $\beta$  levels: Intergroup comparisons across time points; (f) TNF $\alpha$  levels: Intergroup comparisons across time points. FXII, factor XII; FXIIa, factor XIIa; FVIII-Ag, factor VIII-related antigen; PAP, plasmin-antiplasmin complex; IL-1 $\beta$ , interleukin one beta, and TNF $\alpha$ : tumor necrosis factor-alpha. Pre, immediately prior to anesthesia induction; post 0 h, immediately after surgery; post 24 h, 24 h after surgery. \*\*\*\*: p < 0.0001; \*\*\*: p < 0.001; \*\*: p < 0.005; \*: p < 0.05; ns: p > 0.05.

The "contact system" is a plasma protease cascade that plays an important role in the activation of the coagulation systems; it includes FXII, FXIIa, factor XI (FXI), highmolecular-weight kininogen (HMWK), and prekallikrein, which are associated proteins in the plasma [17]. Contact system activity increases dramatically as blood comes into contact with the artificial materials comprising CPB circuits [35]. Mechanistically, FXII cleaves itself upon contact with various anionic surfaces, resulting in the production of FXIIa, which subsequently converts prekallikrein into active kallikrein. In the plasma, kallikrein generates a positive feedback loop by cleaving additional FXII, which produces more bradykinin from HMWK [17]. Previous studies have shown that active thrombin can induce the release of tissue plasminogen activator in vitro, with bradykinin potentially serving as the predominant stimulus [36,37]. Meanwhile, the binding of soluble fibrin to CPB circuits promotes plasminogen activation [38,39].

Previous studies have reported that coagulation disorders can be deteriorated by use of CPB during surgery, especially in conditions that promote deep hypothermia [20,33,40,41]. For example, some groups have reported that CPB causes an increase in the levels of FXIIa and FDPs [17,20]. A study conducted by Boisclair *et al.* [42] suggested that FXIIa can serve as a marker of contact system activation in patients undergoing CPB procedures. In ad-

dition to producing kallikrein, FXIIa has been reported to convert FXI into FXIa, thereby initiating the activation of the intrinsic pathway [20]. Therefore, the contact system exacerbates the development of coagulation system disorders when CPB is performed, and this effect may be related to the length of time required to complete CPB procedures. FXIIa, D-dimer and FDPs may serve as early biomarkers for identifying high-risk ARDS patients. We should also pay attention to the coagulation system of patients with prolonged CPB duration.

# 4.3 Correlation Between the Activation of Coagulation and Postoperative ARDS

In the ATAAD group, the concentrations of D-dimer and FDPs increased immediately after surgery in the patients who developed ARDS, indicating that coagulation activity was elevated; however, the PAP concentration did not differ between the ATAAD and AA groups and was lower in patients with hypoxemia. The pathophysiology of ARDS is complex, and the underlying mechanisms have yet to be fully elucidated, especially during acute critical illness [43].

Matthay et al. [44] have speculated that one of the mechanisms underlying ARDS pathogenesis involves the activation and dysregulation of coagulation, both in the lungs and systemically. As a result, increased fibrinolytic activity may be insufficient to counteract the amplified co-



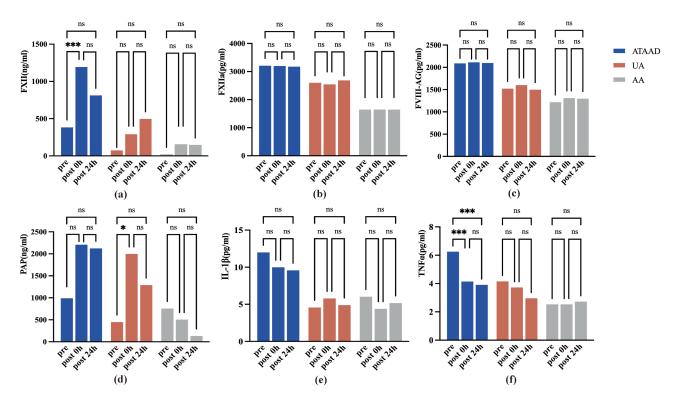


Fig. 2. The serum concentration of coagulation and inflammation biomarkers at different time points for the three groups. (a) FXII levels: Intragroup temporal variations; (b) FXIIa levels: Intragroup temporal variations; (c) FVIII-Ag levels: Intragroup temporal variations; (d) PAP levels: Intragroup temporal variations; (e) IL-1 $\beta$  levels: Intragroup temporal variations; (f) TNF $\alpha$  levels: Intragroup temporal variations. FXII, factor XII; FXIIa, factor XIIa; FVIII-Ag, factor VIII-related antigen; PAP, plasmin-antiplasmin complex; IL-1 $\beta$ , interleukin one beta, and TNF $\alpha$ , tumor necrosis factor-alpha. Pre, immediately prior to anesthesia induction; post 0 h, immediately after surgery; post 24 h, 24 h after surgery. \*\*\*: p < 0.001; \*: p < 0.05; ns: p > 0.05.

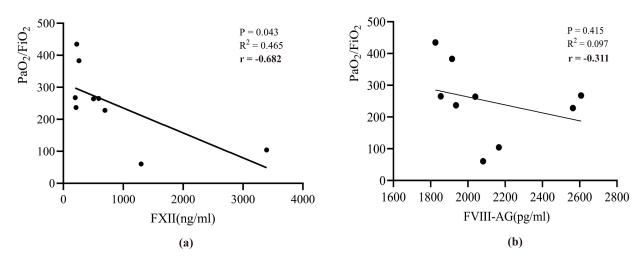


Fig. 3. The correlation between coagulation factors and oxygenation index in patients with ATAAD. (a) Association of FXII concentration with PaO<sub>2</sub>/FiO<sub>2</sub>; (b) Association of FVIII-Ag concentration with PaO<sub>2</sub>/FiO<sub>2</sub>. PaO<sub>2</sub>/FiO<sub>2</sub>, oxygenation index; FXII, factor XII; FVIII-Ag, factor VIII-related antigen.

agulation activity in patients with ARDS. This disequilibrium may lead to fibrin deposition along the denuded alveolar basement membrane, prompting hyaline membrane formation; such lesions can, in turn, decrease lung compliance

and increase inspiratory pressures [43]. In addition, the activation of procoagulant pathways may cause microvascular thromboses in the lungs; the increased amount of dead space reduces blood flow and subsequently influences gas



exchange in those with ARDS [43,45,46]. In this study, the oxygenation index was negatively correlated with the concentration of FXII and was positively correlated with the concentration of PAP 24 hours post-surgery, manifesting as a change in coagulation activity that may be associated with ARDS. However, given the limited sample size, additional research is warranted to validate these findings.

The concentrations of the pro-inflammatory cytokines IL-1 $\beta$  and TNF $\alpha$  were also significantly elevated in the ATAAD group. Correlations between ALI and inflammatory responses have been extensively investigated [22,47]. Both local and systemic acute inflammation are prominent features of ARDS [43], and previous studies have reported that contact with the components of bypass circuits and ischemia–reperfusion injury may upregulate the expression of inflammatory cytokines, thereby leading to the release of coagulation factors [48,49]. Meanwhile, both FXII and contact activation may induce bradykinin release and complement system activation, further promoting the systemic inflammatory response [48–50]. These findings indicated that specific interactions between coagulation and inflammation pathways contribute to lung injury.

To date, there has been little evidence to support the possibility that coagulation activation at an early stage following cardiac surgery can increase the likelihood of ARDS development, especially in patients with ATAAD. One of the strengths of this study was the detailed comparison that was performed of the relationships between coagulation disorders and postoperative ARDS between the three groups; however, there were some limitations. Firstly, the study population was derived from a single center, and it was not possible to exclude other factors that may have affected oxygenation; further validation is required with a larger cohort. Secondly, ELISAs were used to test serum samples from just nine randomly selected patients in each group. Future investigations should examine other markers of coagulation and fibrinolysis in larger cohorts. Thirdly, local coagulation and inflammatory activity are also important, as they can influence the occurrence of hypoxemia; further analysis of the coagulation biomarkers will be performed based on bronchoalveolar lavage fluid.

## 5. Conclusion

In conclusion, the ATAAD- or CPB-induced activation of coagulation pathways can affect gas exchange in the lungs. The early increase in the levels of FXII, FXIIa, D-dimer, and FDPs and the decrease in PAP may play an important role in the mechanisms through which patients with ATAAD develop ARDS following surgical procedures involving CPB. This study offers new insights into the clinical treatment of ARDS using aggressive anticoagulant therapies. Further studies are needed to elucidate the underlying cellular and molecular mechanisms through which the activation of coagulation cascades drives the development of postoperative ARDS in patients with ATAAD.

### **Abbreviations**

AA, aortic aneurysm; ACI, acute lung injury; APTT, activated partial thromboplastin time; ARDS, acute respiratory distress syndrome; AST, aspartate aminotransferase; ATAAD, acute type A aortic dissection; BIC, Bayesian information criterion; BMI, body mass index; CIs, confidence intervals; CKD, chronic kidney disease; CLD, chronic liver disease; CPB, cardiopulmonary bypass; CS, cardiac surgery; CT, computed tomography; DHCA, deep hypothermic circulatory arrest; ELISA, enzyme-linked immunosorbent assay; ESICM, European Society of Intensive Care Medicine; EuroSCORE, European System for Cardiac Operative Risk Evaluation; FXI, factor XI; FXII, factor XII; FXIIa, factor XIIa; FVIII-Ag, factor VIII-related antigen; HF, heart failure; HMWK, high-molecular-weight kininogen; HTN, hypertension; ICU, intensive care unit; IL-1 $\beta$ , interleukin one beta; ND, neurological dysfunction; OPCABG, off-pump coronary artery bypass grafting; ORs, odds ratios; PAP, plasmin- $\alpha$ 2-antiplasmin complex; TNF- $\alpha$ , tumor necrosis factor alpha; UA, unstable angina; VIS, vasoactive inotrope score.

## Availability of Data and Materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

### **Author Contributions**

NL was involved in the conception and design of the study. ML and TW have made substantial dedication to acquisition and analysis of data and datadraft the article. YY, ZC, SC and LY conducted immunoassays and data acquisition and was involved in drafting the manuscript. LL was involved in the analysis of data and datadraft the article. XL and LW was involved in patient recruitment and reviewed the draft critically. NL undertook a critical revision of intellectual content. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

## **Ethics Approval and Consent to Participate**

The study was carried out in accordance with the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of Beijing Anzhen Hospital, Capital Medical University (Protocol No. 20242001X). The patients randomly selected for ELISA testing need to sign informed consent form, while the others were exempted from signing it by the committee.

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### **Conflict of Interest**

The authors declare no conflict of interest.

## **Supplementary Material**

Supplementary material associated with this article can be found, in the online version, at https://doi.org/10.31083/RCM36372.

### References

- [1] Fanelli V, Vlachou A, Ghannadian S, Simonetti U, Slutsky AS, Zhang H. Acute respiratory distress syndrome: new definition, current and future therapeutic options. Journal of Thoracic Disease. 2013; 5: 326–334. https://doi.org/10.3978/j.issn.2072-1439.2013.04.05.
- [2] Villar J, Sulemanji D, Kacmarek RM. The acute respiratory distress syndrome: incidence and mortality, has it changed? Current Opinion in Critical Care. 2014; 20: 3–9. https://doi.org/10.1097/MCC.0000000000000057.
- [3] Stephens RS, Shah AS, Whitman GJR. Lung injury and acute respiratory distress syndrome after cardiac surgery. The Annals of Thoracic Surgery. 2013; 95: 1122–1129. https://doi.org/10.1016/j.athoracsur.2012.10.024.
- [4] Fernández-Pérez ER, Sprung J, Afessa B, Warner DO, Vachon CM, Schroeder DR, et al. Intraoperative ventilator settings and acute lung injury after elective surgery: a nested case control study. Thorax. 2009; 64: 121–127. https://doi.org/10.1136/thx. 2008.102228.
- [5] Grasselli G, Calfee CS, Camporota L, Poole D, Amato MBP, Antonelli M, et al. ESICM guidelines on acute respiratory distress syndrome: definition, phenotyping and respiratory support strategies. Intensive Care Medicine. 2023; 49: 727–759. https://doi.org/10.1007/s00134-023-07050-7.
- [6] Welty-Wolf KE, Carraway MS, Ortel TL, Piantadosi CA. Coagulation and inflammation in acute lung injury. Thrombosis and Haemostasis. 2002; 88: 17–25.
- [7] Milot J, Perron J, Lacasse Y, Létourneau L, Cartier PC, Maltais F. Incidence and predictors of ARDS after cardiac surgery. Chest. 2001; 119: 884–888. https://doi.org/10.1378/chest.119.3.884.
- [8] Messent M, Sullivan K, Keogh BF, Morgan CJ, Evans TW. Adult respiratory distress syndrome following cardiopulmonary bypass: incidence and prediction. Anaesthesia. 1992; 47: 267– 268. https://doi.org/10.1111/j.1365-2044.1992.tb02134.x.
- [9] Sanfilippo F, Palumbo GJ, Bignami E, Pavesi M, Ranucci M, Scolletta S, et al. Acute Respiratory Distress Syndrome in the Perioperative Period of Cardiac Surgery: Predictors, Diagnosis, Prognosis, Management Options, and Future Directions. Journal of Cardiothoracic and Vascular Anesthesia. 2022; 36: 1169–1179. https://doi.org/10.1053/j.jvca.2021.04.024.
- [10] Bronicki RA, Hall M. Cardiopulmonary Bypass-Induced Inflammatory Response: Pathophysiology and Treatment. Pediatric Critical Care Medicine: a Journal of the Society of Critical Care Medicine and the World Federation of Pediatric Intensive and Critical Care Societies. 2016; 17: S272–S278. https://doi.org/10.1097/PCC.0000000000000759.
- [11] Rubenfeld GD, Caldwell E, Peabody E, Weaver J, Martin DP, Neff M, *et al.* Incidence and outcomes of acute lung injury. The New England Journal of Medicine. 2005; 353: 1685–1693. https://doi.org/10.1056/NEJMoa050333.

- [12] Bellani G, Pham T, Laffey J, LUNG-SAFE Investigators, ES-ICM Trials Group. Incidence of Acute Respiratory Distress Syndrome–Reply. JAMA. 2016; 316: 347. https://doi.org/10.1001/jama.2016.6471.
- [13] Herridge MS, Tansey CM, Matté A, Tomlinson G, Diaz-Granados N, Cooper A, et al. Functional disability 5 years after acute respiratory distress syndrome. The New England Journal of Medicine. 2011; 364: 1293–1304. https://doi.org/10.1056/NE JMoa1011802.
- [14] Paparella D, Yau TM, Young E. Cardiopulmonary bypass induced inflammation: pathophysiology and treatment. An update. European Journal of Cardio-thoracic Surgery: Official Journal of the European Association for Cardiothoracic Surgery. 2002; 21: 232–244. https://doi.org/10.1016/ s1010-7940(01)01099-5.
- [15] Massoudy P, Zahler S, Becker BF, Braun SL, Barankay A, Meisner H. Evidence for inflammatory responses of the lungs during coronary artery bypass grafting with cardiopulmonary bypass. Chest. 2001; 119: 31–36. https://doi.org/10.1378/chest.119.1.31.
- [16] Rasmussen BS, Maltesen RG, Pedersen S, Kristensen SR. Early coagulation activation precedes the development of acute lung injury after cardiac surgery. Thrombosis Research. 2016; 139: 82–84. https://doi.org/10.1016/j.thromres.2016.01.015.
- [17] Sniecinski RM, Chandler WL. Activation of the hemostatic system during cardiopulmonary bypass. Anesthesia and Analgesia. 2011; 113: 1319–1333. https://doi.org/10.1213/ANE.0b 013e3182354b7e.
- [18] Liu Z, Liu D, Wang Z, Zou Y, Wang H, Li X, et al. Association between inflammatory biomarkers and acute respiratory distress syndrome or acute lung injury risk: A systematic review and meta-analysis. Wiener Klinische Wochenschrift. 2022; 134: 24– 38. https://doi.org/10.1007/s00508-021-01971-3.
- [19] Welty-Wolf KE, Carraway MS, Miller DL, Ortel TL, Ezban M, Ghio AJ, et al. Coagulation blockade prevents sepsis-induced respiratory and renal failure in baboons. American Journal of Respiratory and Critical Care Medicine. 2001; 164: 1988–1996. https://doi.org/10.1164/ajrccm.164.10.2105027.
- [20] Paparella D, Brister SJ, Buchanan MR. Coagulation disorders of cardiopulmonary bypass: a review. Intensive Care Medicine. 2004; 30: 1873–1881. https://doi.org/10.1007/s00134-004-2388-0.
- [21] Bartoszko J, Karkouti K. Managing the coagulopathy associated with cardiopulmonary bypass. Journal of Thrombosis and Haemostasis: JTH. 2021; 19: 617–632. https://doi.org/10.1111/jth.15195.
- [22] Gao Z, Pei X, He C, Wang Y, Lu J, Jin M, et al. Oxygenation impairment in patients with acute aortic dissection is associated with disorders of coagulation and fibrinolysis: a prospective observational study. Journal of Thoracic Disease. 2019; 11: 1190– 1201. https://doi.org/10.21037/jtd.2019.04.32.
- [23] Nienaber CA, Clough RE. Management of acute aortic dissection. Lancet (London, England). 2015; 385: 800–811. https://doi.org/10.1016/S0140-6736(14)61005-9.
- [24] Nashef SA, Roques F, Michel P, Gauducheau E, Lemeshow S, Salamon R. European system for cardiac operative risk evaluation (EuroSCORE). European Journal of Cardio-thoracic Surgery: Official Journal of the European Association for Cardio-thoracic Surgery. 1999; 16: 9–13. https://doi.org/10.1016/s1010-7940(99)00134-7.
- [25] Vincent JL, Moreno R, Takala J, Willatts S, De Mendonça A, Bruining H, et al. The SOFA (Sepsis-related Organ Failure Assessment) score to describe organ dysfunction/failure. On behalf of the Working Group on Sepsis-Related Problems of the European Society of Intensive Care Medicine. Intensive



- Care Medicine. 1996; 22: 707–710. https://doi.org/10.1007/BF
- [26] Evangelista A, Isselbacher EM, Bossone E, Gleason TG, Eusanio MD, Sechtem U, et al. Insights From the International Registry of Acute Aortic Dissection: A 20-Year Experience of Collaborative Clinical Research. Circulation. 2018; 137: 1846–1860. https://doi.org/10.1161/CIRCULATIONAHA.117. 031264.
- [27] ten Cate JW, Timmers H, Becker AE. Coagulopathy in Ruptured or Dissecting Aortic Aneurysms. The American Journal of Medicine. 1975; 59: 171–176. https://doi.org/10.1016/0002-9343(75)90351-4.
- [28] Paparella D, Rotunno C, Guida P, Malvindi PG, Scrascia G, De Palo M, et al. Hemostasis alterations in patients with acute aortic dissection. The Annals of Thoracic Surgery. 2011; 91: 1364– 1369. https://doi.org/10.1016/j.athoracsur.2011.01.058.
- [29] Guan XL, Wang XL, Liu YY, Lan F, Gong M, Li HY, et al. Changes in the Hemostatic System of Patients With Acute Aortic Dissection Undergoing Aortic Arch Surgery. The Annals of Thoracic Surgery. 2016; 101: 945–951. https://doi.org/10.1016/ j.athoracsur.2015.08.047.
- [30] Albini P, Barshes NR, Russell L, Wu D, Coselli JS, Shen YH, et al. D-dimer levels remain elevated in acute aortic dissection after 24 h. The Journal of Surgical Research. 2014; 191: 58–63. https://doi.org/10.1016/j.jss.2014.03.074.
- [31] Liu T, Zheng J, Zhang YC, Zhu K, Gao HQ, Zhang K, et al. Association Between D-dimer and Early Adverse Events in Patients With Acute Type A Aortic Dissection Undergoing Arch Replacement and the Frozen Elephant Trunk Implantation: A Retrospective Cohort Study. Frontiers in Physiology. 2020; 10: 1627. https://doi.org/10.3389/fphys.2019.01627.
- [32] Wu Q, Li J, Chen L, Yan LL, Qiu Z, Shen Y, *et al.* Efficacy of interleukin-6 in combination with D-dimer in predicting early poor postoperative prognosis after acute stanford type a aortic dissection. Journal of Cardiothoracic Surgery. 2020; 15: 172. https://doi.org/10.1186/s13019-020-01206-y.
- [33] Zindovic I, Sjögren J, Bjursten H, Ingemansson R, Ingimarsson J, Larsson M, *et al.* The Coagulopathy of Acute Type A Aortic Dissection: A Prospective, Observational Study. Journal of Cardiothoracic and Vascular Anesthesia. 2019; 33: 2746–2754. https://doi.org/10.1053/j.jyca.2019.02.013.
- [34] Sbarouni E, Georgiadou P, Marathias A, Geroulanos S, Kremastinos DT. D-dimer and BNP levels in acute aortic dissection. International Journal of Cardiology. 2007; 122: 170–172. https://doi.org/10.1016/j.ijcard.2006.11.056.
- [35] Campbell DJ, Dixon B, Kladis A, Kemme M, Santamaria JD. Activation of the kallikrein-kinin system by cardiopulmonary bypass in humans. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology. 2001; 281: R1059–70. https://doi.org/10.1152/ajpregu.2001.281.4.R1059.
- [36] Booyse FM, Bruce R, Dolenak D, Grover M, Casey LC. Rapid release and deactivation of plasminogen activators in human endothelial cell cultures in the presence of thrombin and ionophore A23187. Seminars in Thrombosis and Hemostasis. 1986; 12: 228–230. https://doi.org/10.1055/s-2007-1003558.
- [37] Fuhrer G, Gallimore MJ, Heller W, Hoffmeister HE. Aprotinin in cardiopulmonary bypass-effects on the Hageman fac-

- tor (FXII)–Kallikrein system and blood loss. Blood Coagulation & Fibrinolysis: an International Journal in Haemostasis and Thrombosis. 1992; 3: 99–104. https://doi.org/10.1097/00001721-199202000-00014.
- [38] Zhao X, Courtney JM, Yin HQ, West RH, Lowe GDO. Blood interactions with plasticised poly (vinyl chloride): influence of surface modification. Journal of Materials Science. Materials in Medicine. 2008; 19: 713–719. https://doi.org/10.1007/ s10856-007-3191-6.
- [39] van den Goor JM, van Oeveren W, Rutten PM, Tijssen JG, Eijsman L. Adhesion of thrombotic components to the surface of a clinically used oxygenator is not affected by Trillium coating. Perfusion. 2006; 21: 165–172. https://doi.org/10.1191/ 0267659106pf859oa.
- [40] Van Poucke S, Stevens K, Marcus AE, Lancé M. Hypothermia: effects on platelet function and hemostasis. Thrombosis Journal. 2014; 12: 31. https://doi.org/10.1186/s12959-014-0031-z.
- [41] Rohrer MJ, Natale AM. Effect of hypothermia on the coagulation cascade. Critical Care Medicine. 1992; 20: 1402–1405. https://doi.org/10.1097/00003246-199210000-00007.
- [42] Boisclair MD, Lane DA, Philippou H, Esnouf MP, Sheikh S, Hunt B, et al. Mechanisms of thrombin generation during surgery and cardiopulmonary bypass. Blood. 1993; 82: 3350– 3357.
- [43] Bos LDJ, Ware LB. Acute respiratory distress syndrome: causes, pathophysiology, and phenotypes. Lancet (London, England). 2022; 400: 1145–1156. https://doi.org/10.1016/ S0140-6736(22)01485-4.
- [44] Matthay MA, Zemans RL, Zimmerman GA, Arabi YM, Beitler JR, Mercat A, et al. Acute respiratory distress syndrome. Nature Reviews. Disease Primers. 2019; 5: 18. https://doi.org/10.1038/s41572-019-0069-0.
- [45] Shetty S, Padijnayayveetil J, Tucker T, Stankowska D, Idell S. The fibrinolytic system and the regulation of lung epithelial cell proteolysis, signaling, and cellular viability. American Journal of Physiology. Lung Cellular and Molecular Physiology. 2008; 295: L967–75. https://doi.org/10.1152/ajplung.90349.2008.
- [46] MacLaren R, Stringer KA. Emerging role of anticoagulants and fibrinolytics in the treatment of acute respiratory distress syndrome. Pharmacotherapy. 2007; 27: 860–873. https://doi.org/ 10.1592/phco.27.6.860.
- [47] Guan X, Li J, Gong M, Lan F, Zhang H. The hemostatic disturbance in patients with acute aortic dissection: A prospective observational study. Medicine. 2016; 95: e4710. https://doi.org/10.1097/MD.0000000000004710.
- [48] Hall RI, Smith MS, Rocker G. The systemic inflammatory response to cardiopulmonary bypass: pathophysiological, therapeutic, and pharmacological considerations. Anesthesia and Analgesia. 1997; 85: 766–782. https://doi.org/10.1097/ 00000539-199710000-00011.
- [49] Aljure OD, Fabbro M, 2nd. Cardiopulmonary Bypass and Inflammation: The Hidden Enemy. Journal of Cardiothoracic and Vascular Anesthesia. 2019; 33: 346–347. https://doi.org/10.1053/j.jvca.2018.05.030.
- [50] Baumann Kreuziger L, Karkouti K, Tweddell J, Massicotte MP. Antithrombotic therapy management of adult and pediatric cardiac surgery patients. Journal of Thrombosis and Haemostasis: JTH. 2018; 16: 2133–2146. https://doi.org/10.1111/jth.14276.

